

TITLE OF THE INVENTION

NUCLEIC ACID SEQUENCES ENCODING NOVEL POINT MUTATIONS ON mGluR2
AND mGluR3, POLYPEPTIDES WITH SAID MUTATIONS, AND METHODS OF USING
SAID NUCLEIC ACID SEQUENCES, AND SAID POLYPEPTIDES, TO IDENTIFY,
5 PREDICT AND EVALUATE SPECIFIC, SELECTIVE MODULATORS WHOSE
ASSOCIATION TO mGlu2 OR mGluR3 IS EFFECTED BY SAID MUTATIONS

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Application No.
10 60/60/409,910, filed September 9, 2002, the contents of which are incorporated herein by
reference in their entirety.

STATEMENT REGARDING FEDERALLY-SPONSORED R&D

Non applicable.

REFERENCE TO MICROFICHE APPENDIX

Not applicable.

BACKGROUND OF THE INVENTION

20 In the mammalian central nervous system (CNS), the transmission of nerve
impulses is controlled by the interaction between a neurotransmitter, released by a sending
neuron, and a surface receptor on a receiving neuron. This interaction causes excitation of the
receiving neuron. L-Glutamate, which is the most abundant neurotransmitter in the CNS,
mediates the major excitatory pathway in mammals, and is referred to as an excitatory amino
25 acid (EAA). The receptors that respond to glutamate are called excitatory amino acid receptors
(EAA receptors). See Watkins & Evans, Annual Reviews in Pharmacology and Toxicology,
21:165 (1981); Monaghan, Bridges, and Cotman, Annual Reviews in Pharmacology and
Toxicology, 29:365 (1989); Watkins, Krogsgaard-Larsen, and Honore, Transactions in
Pharmaceutical Science, 11:25 (1990). The excitatory amino acids are of great physiological
30 importance, playing a role in a variety of physiological processes, such as long-term potentiation
(learning and memory), the development of synaptic plasticity, motor control, respiration,
cardiovascular regulation, and sensory perception.

Excitatory amino acid receptors are classified into two general families –
ionotropic and metabotropic. Ionotropic receptors are directly coupled to the opening of cation
35 channels in the cell membrane of the neurons. This type of receptor has been subdivided into at

least four subtypes, which are defined by the depolarizing actions of the selective agonists γ -aminobutyric acid (GABA), N-methyl-D-aspartate (NMDA), α -amino-3-hydroxy-5-methylisoxazole-4-propionic acid (AMPA), and nicotinic acetylcholine (ACh). Kandel et al., Principles of Neural Science, 3d ed., Elsevier Science Press, 1991.

5 Metabotropic, the second general EAA receptor family, includes the G-protein (guanosine nucleotide-binding protein) or second messenger-linked "metabotropic" excitatory amino acid receptor (also known as a "G Protein-Coupled Receptor", or a "GPCR"). This second type is coupled to multiple second messenger systems that lead to enhanced phosphoinositide hydrolysis, activation of phospholipase D, increases or decreases in cAMP
10 formation, or changes in ion channel function. Schoepp and Conn, Trends in Pharmacological Science, 14:13 (1993). Both types of receptors appear not only to mediate normal synaptic transmission along excitatory pathways, but also participate in the modification of synaptic connections during development and throughout life. Schoepp, Bockaert, and Sladeczek, Trends in Pharmacological Science, 11:508 (1990); McDonald and Johnson, Brain Research Reviews,
15 15:41 (1990).

The excessive or inappropriate stimulation of excitatory amino acid receptors leads to neuronal cell damage or loss by way of a mechanism known as excitotoxicity. This process has been suggested to mediate neuronal degeneration in a variety of conditions. The medical consequences of such neuronal degeneration makes the abatement of these degenerative
20 neurological processes an important therapeutic goal. In that one effect of a positive mGluR2 modulator (e.g., an mGluR2 potentiator) is to decrease activity at a synapse near mGluR2 receptors (by stimulating mGluR2 activity), the development of specific positive modulators for mGluR2 appears to be an appropriate approach to control neuronal pathways and, in doing so, reducing neural cell damage. A key to the specificity, and expected greater effectiveness, of
25 such modulators, whether for mGluR2, mGluR3, or other GPCRs, is to be able to evaluate and identify modulators that, at a pharmacologically acceptable and effective dose, have a positive effect on one such GPCR, while not having the same positive effect on other GPCRs which have opposite or undesired effects on the cell.

In this regard, it is noted that the metabotropic glutamate receptors ("mGluRx,"
30 where x is an integer) are a highly heterogeneous family of glutamate receptors. All are linked to multiple second-messenger pathways. Presently at least eight identified subtypes of such receptors that fall into three classes based on second-messenger association, sequence homology, and agonist selectivity. See BOND, A. et al., "Neuroprotective Effects of LY379268, a Selective mGlu2/3 Receptor Agonist: Investigations into Possible Mechanism of Action In Vivo", JPET,
35 2000, 800-809: 294, USA. Also see "G Protein-Coupled Receptor Allosterism and

Complexing" Pharmacological Reviews, 54:2, 323-374. These receptors function to modulate the presynaptic release of glutamate, and the postsynaptic sensitivity of the neuronal cell to glutamate excitation. It is generally recognized that agonists and antagonists of these receptors may be useful for the treatment of acute and chronic neurodegenerative conditions, and as
5 antipsychotic, anticonvulsant, analgesic, anxiolytic, antidepressant, and anti-emetic agents. The present invention discloses at least one allosteric binding site which would be useful for the identification and development of mGluR2- and mGluR3-specific modulators that associate with such site.

In recent years, the role of allosteric modulators for the metabotropic glutamate
10 receptors has been a subject of increasing interest and research. An advantage of developing a specific allosteric modulator for one type of metabotropic glutamate receptor is that the use of such modulator may provide a desirable pharmacological and behavioral result at lower doses, with less toxicity and side effects due to its specificity.

The present invention advances the art by identifying a site of allosteric
15 modulation in mGluR2 and mGluR3, and by characterizing critical components of said site, through point mutations, where such mutations have a dramatic effect on potentiation of the glutamate cell receptor.

SUMMARY OF THE INVENTION

The present invention provides novel forms of mGluR2 and mGluR3 comprising
20 at least one allosteric binding site which has been modified by one, two, or three single amino acid point mutations. The loss of potentiation of receptors bearing such mutations in mGluR2 is clear and dramatic. The present invention provides nucleic acid sequences, expressible in isolated cells, useful for the identification of novel modulators and for the rational development
25 of specific modulators of metabotropic glutamate receptors. The present invention also provides for polypeptides encoded by the herein disclosed isolated nucleic acid molecules that will find use in the identification of novel modulators and for the rational development of specific modulators of metabotropic glutamate receptors. The identification of the herein described nucleic acid and their respective mutant gene products will aid in the characterization and
30 treatment of physiological disorders is hereby furthered.

In particular, the present invention advances the art by identifying at least one site of allosteric modulation in mGluR2 and mGluR3, by characterizing the said site through such mutations. These mutations result in a dramatic effect on potentiation by modulators at said site. In particular, certain modulators that have a dramatic effect on wild type of mGluR2 are shown

to have far less effect on variant mGluR2 receptors in which one or more such point mutations are introduced. Critically, such effect is not observed by the other specific subtype, mGluR3.

The present invention also provides for assays to identify and determine the efficacy and reaction profile of modulators acting at such newly identified allosteric binding sites, which proves useful in the treatment or prevention of disorders associated with an excess or deficiency in the amount of glutamate present. Modulators developed with a high specificity to a single type of metabotropic G-Protein Receptor is expected to have greater utility and efficacy because: 1) as a modulator, it can function at lower concentrations than an agonist or antagonist; and 2) with great specificity, a modulator can precisely target the desired metabotropic G-Protein Receptor without affecting the activity of related receptors having different, potentially undesired cellular effects. This is perceived to provide for development of modulators whose use in a patient in need thereof results in less overall toxicity to said patient.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 provides a summary chart of the normalized response to glutamate in HEK-293 cells transfected with wild-type mGluR2 receptors, with some treatments exposed to two mGluR2 modulators, A and B.

Figure 2 provides a summary chart of the normalized response to glutamate in HEK-293 cells transfected with a mutant mGluR2 receptor of the present invention, with some treatments exposed to two mGluR2 modulators, A and B.

Figure 3 provides a summary chart of the normalized response to glutamate in HEK-293 cells transfected with another mutant mGluR2 receptor of the present invention, with some treatments exposed to two mGluR2 modulators, A and B.

Figure 4 provides a summary chart of the normalized response to glutamate in HEK-293 cells transfected with another mutant mGluR2 receptor of the present invention, with some treatments exposed to two mGluR2 modulators, A and B.

Figure 5 provides a summary chart of the normalized response to glutamate in HEK-293 cells transfected with another mutant mGluR2 receptor of the present invention, with some treatments exposed to two mGluR2 modulators, A and B.

Figure 6 provides a summary chart of the normalized response to glutamate in HEK-293 cells transfected with another mutant mGluR2 receptor of the present invention, with some treatments exposed to two mGluR2 modulators, A and B.

DETAILED DESCRIPTION OF THE INVENTION

Definitions and Terms of Art

The terms and abbreviations used in this document have their normal meanings unless otherwise designated. For example "°C" refers to degrees Celsius; "N" refers to normal or normality; "mmol" refers to millimole or millimoles; "g" refers to gram or grams; "ml" means milliliter or milliliters; "M" refers to molar or molarity; "µg" and "ug" refer to microgram or micrograms; "µM" and "uM" refer to microMole or micromoles; and "µl" and "ul" refer to microliter or microliters. It is noted that some definitions are located outside this section.

All nucleic acid sequences, unless otherwise designated, are written in the direction from the 5' end to the 3' end, frequently referred to as "5' to 3".

All amino acid or protein sequences, unless otherwise designated, are written commencing with the amino terminus ("N-terminus") and concluding with the carboxy terminus ("C-terminus").

"Base pair" or "bp" as used herein refers to DNA or RNA. The abbreviations A, C, G, and T correspond to the 5'-monophosphate forms of the deoxyribonucleosides (deoxy)adenosine, (deoxy)cytidine, (deoxy)guanosine, and (deoxy)thymidine, respectively, when they occur in DNA molecules. The abbreviations U, C, G, and A correspond to the 5'-monophosphate forms of the ribonucleosides uridine, cytidine, guanosine, and adenosine, respectively when they occur in RNA molecules. In double stranded DNA, base pair may refer to a pairing of A with T or C with G. In a DNA/RNA, heteroduplex base pair may refer to a pairing of A with U or C with G. (See the definition of "complementary", *infra*.)

The terms "cleavage" or "restriction digestion" of DNA refers to the catalytic cleavage of the DNA with a restriction enzyme that acts only at certain sequences in the DNA ("sequence-specific endonucleases"). The various restriction enzymes used herein are commercially available and their reaction conditions, cofactors, and other requirements were used as would be known to one of ordinary skill in the art. Appropriate buffers and substrate amounts for particular restriction enzymes are specified by the manufacturer or can be readily found in the literature.

"Ligation" refers to the process of forming phosphodiester bonds between two nucleic acid fragments (T. Maniatis, et al., *supra*, p. 146). Unless otherwise provided, ligation may be accomplished using known buffers and conditions with a DNA ligase, such as T4 DNA ligase.

The term "plasmid" refers to an extrachromosomal (usually) self-replicating genetic element. Plasmids are generally designated by a lower case "p" followed by letters and/or numbers. The starting plasmids herein are either commercially available, publicly

available on an unrestricted basis, or can be constructed from available plasmids in accordance with published procedures. In addition, equivalent plasmids to those described are known in the art and will be apparent to the ordinarily skilled artisan.

5 The term "reading frame" means the nucleotide sequence from which translation occurs "read" in triplets by the translational apparatus of transfer RNA (tRNA) and ribosomes and associated factors, each triplet corresponding to a particular amino acid. A frameshift mutation occurs when a base pair is inserted or deleted from a DNA segment. When this occurs, the result is a different protein from that coded for by the DNA segment prior to the frameshift mutation. To insure against this, the triplet codons corresponding to the desired polypeptide
10 must be aligned in multiples of three from the initiation codon, i.e. the correct "reading frame" being maintained.

"Recombinant DNA cloning vector" as used herein refers to any autonomously replicating agent, including, but not limited to, plasmids and phages, comprising a DNA molecule to which one or more additional DNA segments can or have been added.

15 The term "recombinant DNA expression vector" as used herein refers to any recombinant DNA cloning vector in which a promoter and other regulatory elements to control transcription of the inserted DNA.

The term "expression vector system" as used herein refers to a recombinant DNA expression vector in combination with one or more trans-acting factors that specifically influence
20 transcription, stability, or replication of the recombinant DNA expression vector. The trans-acting factor may be expressed from a co-transfected plasmid, virus, or other extrachromosomal element, or may be expressed from a gene integrated within the chromosome.

"Transcription" as used herein refers to the process whereby information contained in a nucleotide sequence of DNA is transferred to a complementary RNA sequence.

25 The term "transfection" as used herein refers to the taking up of an expression vector by a host cell whether or not any coding sequences are in fact expressed. Numerous methods of transfection are known to the ordinarily skilled artisan, for example, lipofectamine, calcium phosphate co-precipitation, and electroporation. Successful transfection is generally recognized when any indication of the operation of this vector occurs within the host cell.

30 The term "transformation" as used herein means the introduction of DNA into an organism so that the DNA is replicable, either as an extrachromosomal element or by chromosomal integration. Methods of transforming bacterial and eukaryotic hosts are well known in the art, many of which methods are summarized in J. Sambrook, et al., "Molecular Cloning: A Laboratory Manual" (1989).

The term "translation" as used herein refers to the process whereby the genetic information of messenger RNA is used to specify and direct the synthesis of a polypeptide chain.

The term "vector" as used herein refers to a nucleic acid compound used for the transformation of cells with polynucleotide sequences corresponding to appropriate protein molecules which when combined with appropriate control sequences confer specific properties on the host cell to be transformed. Plasmids, viruses, and bacteriophage are suitable vectors. Artificial vectors are constructed by joining DNA molecules from different sources. The term "vector" as used herein includes Recombinant DNA cloning vectors and Recombinant DNA expression vectors.

The terms "complementary" or "complementarity" as used herein refers to the pairing of bases, purines and pyrimidines, that associate through hydrogen bonding in double stranded nucleic acid. The following base pairs are complementary: guanine and cytosine; adenine and thymine; and adenine and uracil.

"Isolated amino acid sequence" refers to any amino acid sequence, however constructed or synthesized, which is locationally distinct from the naturally occurring sequence.

"Isolated DNA compound" refers to any DNA sequence, however constructed or synthesized, which is locationally distinct from its natural location in genomic DNA.

"Isolated nucleic acid compound" refers to any RNA or DNA sequence, however constructed or synthesized, which is locationally distinct from its natural location.

A "primer" is a nucleic acid fragment which functions as an initiating substrate for enzymatic or synthetic elongation.

The term "promoter" refers to a DNA sequence which directs transcription of DNA to RNA.

As used herein, a nucleic acid "probe" is single-stranded DNA or RNA, or analog thereof, that has a sequence of nucleotides that includes at least 14, preferably at least 20, more preferably at least 50, contiguous bases that are the same as or the complement of any 14 or more contiguous bases set forth in SEQ.ID.NO.:1. In addition, the entire cDNA-encoding region of the entire sequence corresponding to SEQ.ID.NO.:1 may be used as a probe.

Presently preferred probe-based screening conditions comprise a temperature of about 37°C, a formamide concentration of about 20%, and a salt concentration of about 5X standard saline citrate (SSC; 20X SSC contains 3M sodium chloride, 0.3M sodium citrate, pH 7.0). Such conditions will allow the identification of sequences which have a substantial degree of similarity with the probe sequence, without requiring perfect homology.

"Hybridization" refers to the binding of complementary strands of nucleic acid (i.e., sense:antisense strands or probe:target-DNA) to each other through hydrogen bonds, similar

to the bonds that naturally occur in chromosomal DNA. Stringency levels used to hybridize a given probe with target-DNA can be readily varied by those of skill in the art.

The phrase "stringent hybridization conditioned" is used herein to refer to conditions under which polynucleic acid hybrids are stable. As known to those of skill in the art, the stability of hybrids is reflected in the melting temperature (T_m) of the hybrids. T_m can be approximated by the formula:

$$81.5^{\circ}\text{C} - 16.6(\log_{10} [\text{Na}^+]) + 0.41(\%G+C) - 600/l,$$

where l is the length of the hybrids in nucleotides. T_m decreases approximately 1° - 1.5°C with every 1% decrease in sequence homology. In general, the stability of a hybrid is a function of sodium ion concentration and temperature. Typically, the hybridization reaction is performed under conditions of lower stringency, followed by washes of varying, but higher, stringency.

Reference to hybridization stringency relates to such washing conditions.

As used herein, the phrase "moderately stringent hybridization" refers to conditions that permit target-DNA to bind a complementary nucleic acid that has about 60% identity, preferably about 75% identity, more preferably about 85% identity to the target DNA; with greater than about 90% identity to target-DNA being especially preferred. Preferably, moderately stringent conditions are conditions equivalent to hybridization in 50% formamide, 5X Denhart's solution, 5X SSPE, 0.2% SDS at 42°C , followed by washing in 0.2X SSPE, 0.2% SDS, at 65°C .

The phrase "high stringency hybridization" refers to conditions that permit hybridization of only those nucleic acid sequences that form stable hybrids in 0.018M NaCl at 65°C (i.e., if a hybrid is not stable in 0.018M NaCl at 65°C , it will not be stable under high stringency conditions, as contemplated herein). High stringency conditions can be provided, for example, by hybridization in 50% formamide, 5X Denhart's solution, 5X SSPE, 0.2% SDS at 42°C ., followed by washing in 0.1X SSPE, and 0.1% SDS at 65°C .

The phrase "low stringency hybridization" refers to conditions equivalent to hybridization in 10% formamide, 5X Denhart's solution, 6X SSPE, 0.2% SDS at 42°C ., followed by washing in 1X SSPE, 0.2% SDS, at 50°C .

The term "antigenically distinct" as used herein refers to a situation in which antibodies raised against an epitope of the proteins of the present invention, or a fragment thereof, may be used to differentiate between the proteins of the present invention and other glutamate receptor subtypes. This term may also be employed in the sense that such antibodies

may be used to differentiate between the mutant human mGluR2 and mGluR3 receptor proteins and analogous proteins derived from other species.

The term "PCR" as used herein refers to the widely known polymerase chain reaction employing a thermally-stable polymerase.

5 Further, as used herein, "modulator" may be any molecule, compound, or any other composition which is suspected of being capable of modulating the rate or other substantive characteristic of binding of glutamate to an mGluRx receptor in vivo or in vitro by acting at a site (an allosteric site) on the receptor that is not the agonist binding site.

10 "Modulators" that are screened in the present invention can be any substances that are generally screened in the pharmaceutical industry during the drug development process. The substances may be macromolecules, such as biological polymers, including proteins, polysaccharides, nucleic acids, or the like. More usually, a substance will be a small molecule having a molecular weight below about 2 kD, more usually below 1.5 kD, frequently below 1 kD, and usually in the range from 100 to 1,000 D, and even more usually in the range from 200 D to 750 D. One or
15 more substances may be pre-selected based on a variety of criteria. For example, suitable substances may be selected based upon SAR analysis based upon the calculated or predicted three-dimensional structures of the allosteric binding site discovered herein. Alternatively, the substances may be selected randomly and tested by the screening methods of the present invention. Substances which are able to up-modulate or down-modulate glutamate binding to an
20 mGluRx receptor in vitro are considered as candidates for further screening of their ability to affect the activity of the tested mGluRx in cells and/or animals. Substances are often tested in the methods of the present invention as large collections of substances, e.g. libraries of low molecular weight organic compounds, peptides, or natural products.

25 In the scientific literature, the term "allosteric" has taken somewhat different meanings with different scopes (see "G Protein-Coupled Receptor Allostereism and Complexing" Pharmacological Reviews, 54:2, 323-374, at 326-327). As used herein, "allosteric" is taken to mean a site other than the ligand binding site where a non-ligand molecule (one that is neither an agonist nor an antagonist at the reactive binding site of the molecule) may bind, and wherein such binding affects the binding of the ligand at the ligand binding site (as expressed by rate,
30 association constant, affinity, etc.). In contrast to some definitions in the field, as used herein the binding of a molecule at the ligand binding site need not affect the binding of a molecule at the allosteric site. An "allosteric modulator" is a modulator, as defined above, that binds to an allosteric site on a molecule of interest. It is noted that the same molecule may act as an allosteric modulator on one protein or receptor, and act as an agonist or antagonist with regard to
35 a second protein or receptor.

With regard to the effect of a mutation of an mGluRx polypeptide as described herein, the terms “depotentiate(s)” and “depotentialiation” are taken to mean that the effect of such mutation results in a measurable decrease in the effect of a modulator in combination with said mutant polypeptide in comparison to a non-mutated wild-type mGluRx of the same species and subtype. Preferably, the measurable decrease is substantial as that term is defined herein. Likewise, the terms “potentiate(s)” and “potentiation” are taken to mean that the effect of such mutation results in a measurable increase in the effect of a modulator in combination with said mutant polypeptide in comparison to a non-mutated wild-type mGluRx of the same species and subtype. Preferably, the measurable increase is substantial as that term is defined herein.

A “structure activity relationship” (SAR) refers to the relationship between a given chemical structure or series of chemical structures and the pharmacological activity that series of compounds has on the given target or action of the compound. Compounds can be classed together based on a number of characteristics including but not limited to such structural characteristics as shape, size, stereochemical arrangement, and distribution of functional groups. Other factors contributing to structure- activity relationships can include chemical reactivity, electronic effects, resonance, and inductive effects. The utilization of various assays that can differentiate between structural modifications that produce increased affinity or specificity of a given compound on its target compared to the activity seen on closely related targets or on targets that may be associated with off-target or unwanted activity of the compounds. The refinement of this SAR process can lead to pharmacologically active compounds that are more specific and less toxic than the initially identified compound or series of compounds.

A “conservative amino acid substitution” refers to the replacement of one amino acid residue by another, chemically similar, amino acid residue. Examples of such conservative substitutions are: substitution of one hydrophobic residue (isoleucine, leucine, valine, or methionine) for another; substitution of one polar residue for another polar residue of the same charge (e.g., arginine for lysine; glutamic acid for aspartic acid); substitution of one aromatic amino acid (tryptophan, tyrosine, or phenylalanine) for another.

A “conservative amino acid substitution” as defined above is but one type of variation of an amino acid sequence listing encompassed by the broader term, “conservatively modified variants thereof.” For instance, the latter is taken to have the meaning ascribed to the term in M.P.E.P. § 2422.03, Eighth Edition, 2001, which can include, without being limited to this example, deletions such as “at the C-terminus by 1, 2, 3, 4, or 5 residues.” Where appropriate within this specification, conservative amino acid substitutions that are known or reasonably predicted to not adversely alter the desired functionality of the novel sequences disclosed herein are disclosed. Such disclosed conservative amino acid substitutions are

considered to fall within the scope of the sequence listings that include the novel polypeptide sequences disclosed and claimed herein. The same principal of "conservatively modified variants" applies to nucleotide sequences as well, additionally taking into account the redundancy of codons for a particular amino acid, and the optimization of codons for expression in particular species.

For instance, but not meant to be limiting, an amino acid sequence or a nucleotide sequence is considered "identical" to a reference sequence if the two sequences are the same when aligned for maximum correspondence over a comparison window. Optimal alignment of nucleotide and amino acid sequences for aligning comparison window may be conducted by the local homology algorithm of Smith & Waterman, 1981, Adv. Appl. Math. 2:482, by the homology alignment algorithm of Needleman & Wunsch, 1970, J. Mol. Biol. 48:443, by the search for similarity method of Pearson & Lipman, 1988, Proc. Natl. Acad. Sci., U.S.A. 85:2444-2448, by computerized implementations of these algorithms (GAP, BESFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, Genetics Computer Group, 575 Science Dr., Madison, WI), or by inspection. Such determination of identity can be considered to indicate a "conservatively modified variant" of a particular amino acid sequence or nucleotide sequence so long as the variant continues to function as a polypeptide or nucleic acid sequence according to the present invention.

"Consists essentially," with respect to a mutant polypeptide, indicates that the reference sequence can be modified by N-terminal and/or C-terminal additions or deletions that do not cause a substantial decrease in the ability of the mutant polypeptide to function to affect the binding of an allosteric modulator at the site of the mutation compared to the reference sequence lacking such additions or deletions. An example of a deletion is the removal of an N-terminal methionine.

A "substantial change" in the ability of the mutant mGluR2 and mGluR3 proteins of the present invention to affect the modulation relative to a wild-type control, is defined to be a change of at least about 20%, more usually at least about 50%, preferably at least about 75%, and often at least about 90% or higher compared to the response of said wild-type control. This change may be a decrease or an increase relative to the wild-type control, e.g., a "substantial decrease" or a "substantial increase" per the definition of "substantial change" relative to such control.

The term "antibody" refers to a polypeptide substantially encoded by an immunoglobulin gene or immunoglobulin genes, or fragments thereof, which specifically bind and recognize an analyte (antigen). The recognized immunoglobulin genes include the kappa, lambda, alpha, gamma, delta, epsilon and mu constant region genes, as well as the myriad

immunoglobulin variable region genes. Antibodies exist, e.g., as intact immunoglobulins or as a number of well-characterized fragments produced by digestion with various peptidases. These include, e.g., Fab' and F(ab)'₂ fragments. The term "antibody" also includes antibody fragments either produced by the modification of whole antibodies or those synthesized de novo using recombinant DNA methodologies, and further includes "humanized" antibodies made by conventional techniques.

The term "immunoassay" is an assay that utilizes an antibody to specifically bind an analyte. The immunoassay is characterized by the use of specific binding properties of a particular antibody to isolate, target, and/or quantify the analyte.

An antibody "specifically binds to" or "is specifically immunoreactive with" a protein, polypeptide, or peptide when the antibody functions in a binding reaction which is determinative of the presence of the protein, polypeptide, or peptide in the presence of a heterogeneous population of proteins and other biologics. Thus, under designated immunoassay conditions, the specified antibodies bind preferentially to a particular protein, polypeptide, or peptide and do not bind in a significant amount to other proteins, polypeptides, or peptides present in the sample. Specific binding to a protein, polypeptide, or peptide under such conditions requires an antibody that is selected for specificity for a particular protein, polypeptide, or peptide. As used herein, the term "recognize" as it regards an antibody's association to an a particular protein, polypeptide, or peptide, or an epitope therein, is taken to mean that said antibody "specifically binds to" or "is specifically immunoreactive with that protein, polypeptide, or peptide."

A variety of immunoassay formats may be used to select antibodies specifically immunoreactive with a particular protein, polypeptide, or peptide. For example, solid-phase ELISA immunoassays are routinely used to select monoclonal antibodies specifically immunoreactive with a protein, polypeptide, or peptide. See Harlow & Lane, 1988, Antibodies, A Laboratory Manual, Cold Spring Harbor Publications, New York, for a description of immunoassay formats and conditions that can be used to determine specific immunoreactivity.

"Transfection" refers to any of the methods known in the art for introducing DNA into a cell, for example, but not limited to, the methods of lipofectamine, calcium phosphate or calcium chloride mediated transfection, electroporation, and infection with a retroviral vector.

Experiments Leading to Invention

The identification of the site of action of the mGluR2 potentiators was initiated based on the fact that compounds had been identified that positively modulated mGluR2 in the presence of EC₁₀ of glutamate but had no potentiator activity on mGluR3 (patent references,

Lilly and Merck). The subtype specificity and selectivity of these compounds implied that a subset of the amino acid residues that differed between mGluR2 receptor and mGluR3 were most likely involved in the specific binding of the compound. Initial alignments of the amino acid sequences of human mGluR2 (hmGluR2) and human mGluR3 (hmGluR3) revealed an overall amino acid homology of 66.3%. In order to identify the site of action of the mGluR2 positive modulators, various regions that differed between the hmGluR2 and hmGluR3 receptors were identified and selected for chimeric substitution between sequences contained in hmGluR2 and hmGluR3.

The initial region selected for chimeric substitution was a region in the amino terminal domain of hmGluR2 that was 5' of transmembrane one (Arginine-425 to Glutamic Acid-569). This domain contained a very cysteine rich region. It had recently been reported (Schweitzer et al. (2000)) that zinc selectively inhibited the agonist activity of glutamate on mGluR2 compared to mGluR3. This apparent subtype selectivity of the effect of zinc on mGluR2 compared to mGluR3 could be due to the presence of a zinc finger domain on mGluR2 that was not present on mGluR3. Schweitzer et al. (2000) had not identified the site of action of this zinc effect. We targeted the substitution of the cysteine rich region of mGluR2 for the homologous region in hmGluR3, since it has been widely reported that zinc binds to various cysteine and histidine motifs. Clearly the effect seen by the mGluR2 potentiators was the inverse effect seen by zinc however, it was thought that identification of the site of action of this zinc effect might lead to identification/understanding of the region involved in the potentiator effect.

After the construction of the cysteine-rich chimera was initiated, a series of subsequent chimeric molecules were made to various regions of the transmembrane domains of hmGluR2 and hmGluR3. The rationale for these substitutions was based on the fact that many sites of action for agonists or antagonist have been localized on various subtypes of G-protein coupled receptors (GPCRs) to particular transmembrane domains of these receptors (insert references). However, to our knowledge, the site of action of a receptor specific potentiator had not been localized to a transmembrane domain. The site of action of a mGluR5-specific antagonist, MPEP, was recently localized to three specific residues contained in the metabotropic receptor subtype 5 cDNA, mGluR5a (insert reference) and since the metabotropic receptors represent a family of closely related GPCRs, it was thought that a transmembrane binding domain(s) could be involved in the binding of the mGluR2 potentiators. A series of chimeric constructs were subsequently generated that exchanged regions from TM1-3, TM3-5 or a single amino acid substitution in TM7 from hmGluR2 with homologous regions of hmGluR3.

Detailed Disclosure of Invention, Protocols, and Applications

First, it is noted that the disclosure herein, of mutant forms of mGluR2 and mGluR3, describes and specifies sequences for human forms of these mutant receptors.

However, it is appreciated by those skilled in the art that the invention likewise applies to

5 orthologues of such mutant forms in other species, including but not limited to murine species, in particular the rat and mouse, and also for monkey, ape, and other higher primates. Also, chimeric forms incorporating the novel sequences are prepared by combining genetic material from two or more species, for instance, combining a mutant form of human mGluR2 into a rat model genome.

10 In certain embodiments, the invention provides a mutant polypeptide, where the mutant polypeptide comprises substitution of at least one, two or three amino acids at positions 688, 689 and 735 relative to the wild-type human mGluR2 molecule, and specifically comprising any one, two in combination, or all three of the amino acid substitutions indicated below:

- for mGluR2: leucine for serine at position 688; valine for glycine at position
15 689; and aspartic acid for asparagine at position 735.

- for mGluR3: serine for leucine at position 688; glycine for valine at position
689; and asparagine for aspartic acid at position 735.

That is, based on the research described in the section above, it was learned that the substitutions of the amino acids in wild-type mGluR3 at homologous positions 688, 689 and
20 735, into mGluR2, substantially affected the potentiation of the resultant mutant mGluR2 molecules when said molecules were evaluated in the presence of an mGluR2 modulator that appears to bind to an allosteric binding site involving these amino acid positions. It is noted that said positions 688 and 689 are recognized to reside in transmembrane region 4 (TM4) of mGluR2, and said position 735 is recognized to reside in transmembrane region 5 (TM5) of
25 mGluR2. While not being bound to a particular theory, it is hypothesized that, for mGluR2, the substitution of larger and/or more acidic amino acids at such positions results in a configurational change in an allosteric binding site situated in or near TM4 and TM5. It is further hypothesized that such configurational change makes binding by certain allosteric modulators at such site more difficult, or less effective, resulting in a depotentiation compared to a wild-type mGluR2.

30 Likewise, while not being bound to a particular theory, it is postulated that the "reverse" substitutions, of the wild-type mGluR2 amino acids at these positions in mGluR3, to form mGluR3 mutants, results in potentiation by allosteric modulators at an analogous site on the mutant mGluR3. Further, although this disclosure may state that modulators bind to an allosteric site associated with the amino acids at the above three positions, it is emphasized that this is
35 merely based on a theory hypothesized from the evidence at hand. Critically, the novelty and the

utility of the invention disclosed herein, particularly the mGluR2 and mGluR3 mutants and related methods, are not bound to this particular theory. Instead, the invention relies on the merits of the discoveries and results presented herein, independently of the ultimate accuracy of any of the above hypotheses and theories.

5 Particularly, this invention provides mutant mGluR2 polypeptide sequences selected from the group consisting of SEQ.ID.NOS.:1-8, and conservatively modified variants of such sequences. The data presented herein, particularly in Figure 4 (see below), indicates that four single amino acid mutations together provide a substantial decrease in the effect of two allosteric modulators. However, it is believed that the three mutations noted above are the
10 mutations with the greater effect relative to the fourth mutation, A733T. This belief is based on certain data (not shown) that appears to indicate that the A733T mutation (threonine replacing alanine at position 733) by itself provides an effect similar to that of a wild-type control. In contrast, the testing of the other single mutations by themselves (see Fig. 3 for N735D, other data not shown) indicates a greater difference from the wild-type control.

15 In other embodiments, the invention provides an isolated nucleic acid or a nucleic acid compound that comprises a nucleic acid sequence which encodes for a mutant polypeptide, where the mutant polypeptide comprises a substitution of any one, two in combination, or all three of the amino acid substitutions indicated below:

20 for mGluR2: leucine for serine at position 688; valine for glycine at position 689; and aspartic acid for asparagine at position 735.

 for mGluR3: serine for leucine at position 688; glycine for valine at position 689; and asparagine for aspartic acid at position 735.

25 Particularly, this invention provides isolated nucleic acid sequences selected from the group consisting of SEQ.ID.NOS.:9-16, and conservatively modified variants of such sequences.

 In the above description of the mutations of mGluR3, for convenience of this disclosure, the same numbering used for mGluR2 is used for the mGluR3. It is noted that there is an offset of nine amino acids in the numbering. For example, where a substitution at position 688 in mGluR3 is noted above, the actual position on the mGluR3 molecule is at position 697.

30 Skilled artisans will recognize that the polypeptide, peptides, and fusion proteins of the present invention can be synthesized by any number of different methods. The amino acid compounds of the invention can be made by chemical methods well known in the art, including solid phase peptide synthesis or recombinant methods. Both methods are described in U.S. Pat. No. 4,617,149, incorporated herein by reference.

The principles of solid phase chemical synthesis of polypeptides are well known in the art and may be found in general texts in the area. See. e.g., H. Dugas and C. Penney, *Bioorganic Chemistry* (1981) Springer-Verlag, New York, 54-92. For example, peptides may be synthesized by solid-phase methodology utilizing an Applied Biosystems 430A peptide synthesizer (commercially available from Applied Biosystems, Foster City Calif.) and synthesis cycles supplied by Applied Biosystems. Protected amino acids, such as t-butoxycarbonyl-protected amino acids, and other reagents are commercially available from many chemical supply houses.

Sequential t-butoxycarbonyl chemistry using double couple protocols are applied to the starting p-methyl benzhydryl amine resins for the production of C-terminal carboxamides. For the production of C-terminal acids, the corresponding pyridine-2-aldoxime methiodide resin is used. Asparagine, glutamine, and arginine are coupled using preformed hydroxy benzotriazole esters. The following side chain protection may be used:

Arg, Tosyl
Asp, cyclohexyl
Glu, cyclohexyl
Ser, Benzyl
Thr, Benzyl
Tyr, 4-bromo carbobenzoxy

Removal of the t-butoxycarbonyl moiety (deprotection) may be accomplished with trifluoroacetic acid (TFA) in methylene chloride. Following completion of the synthesis the peptides may be deprotected and cleaved from the resin with anhydrous hydrogen fluoride containing 10% meta-cresol. Cleavage of the side chain protecting group(s) and of the peptide from the resin is carried out at zero degrees Celcius or below, preferably -20°C for thirty minutes followed by thirty minutes at 0°C.

After removal of the hydrogen fluoride, the peptide/resin is washed with ether, and the peptide extracted with glacial acetic acid and then lyophilized. Purification is accomplished by size-exclusion chromatography on a Sephadex G-10 (Pharmacia) column in 10% acetic acid.

The proteins of the present invention alternatively are produced by recombinant methods. Recombinant methods are preferred if a high yield is desired. A general method for the construction of any desired DNA sequence is provided in J. Brown, et al., *Methods in Enzymology*, 68:109 (1979). See also, J. Sambrook, et al., *supra*.

The basic steps in the recombinant production of desired proteins are:

- a) construction of a natural, synthetic or semi-synthetic DNA encoding the protein of interest;
- b) integrating said DNA into an expression vector in a manner suitable for the expression of the protein of interest, either alone or as a fusion protein;
- 5 c) transforming an appropriate eukaryotic or prokaryotic host cell with said expression vector,
- d) culturing said transformed or transfected host cell in a manner to express the protein of interest; and
- 10 e) recovering and purifying the recombinantly produced protein of interest.

In general, prokaryotes are used for cloning of DNA sequences and constructing the vectors of the present invention. Prokaryotes may also be employed in the production of the protein of interest. For example, the *Escherichia coli* K12 strain 294 (ATCC No. 31446) is useful for the prokaryotic expression of foreign proteins. Other strains of *E. coli* (and their
15 relevant genotypes) are well known and commonly used in the art.

Further, in addition strains of *E. coli*, bacilli such as *Bacillus subtilis*, other enterobacteriaceae such as *Salmonella typhimurium* or *Serratia marcescans*, and various *Pseudomonas* species may be used. In addition to these gram-negative bacteria, other bacteria, especially *Streptomyces*, spp., may be employed in the prokaryotic cloning and expression of the
20 proteins of this invention.

Thus, suitable *E. coli* strains, as well as many other suitable prokaryote species and strains are known in the art, and are commercially available from suppliers such as: Bethesda Research Laboratories, Gaithersburg, Md. 20877 and Stratagene Cloning Systems, La Jolla, Calif. 92037; or are readily available to the public from sources such as the American Type
25 Culture Collection, 12301 Parklawn Drive, Rockville, Md., 10852-1776. U.S. Patents 6,017,697 and 6,387,655 disclose additional information regarding suitable species for use, and are incorporated by reference generally and for this specific purpose.

Except where otherwise noted or recognized in the art, the bacterial strains can be used interchangeably. Any particular type of bacterial host stated herein is not meant to limit the
30 invention in any way. Genotype designations are in accordance with standard nomenclature. See, for example, J. Sambrook, et al., supra.

Promoters suitable for use with prokaryotic hosts include the β -lactamase [vector pGX2907 (ATCC 39344) contains the replicon and β -lactamase gene] and lactose promoter systems [Chang et al., *Nature* (London), 275:615 (1978); and Goeddel et al., *Nature* (London),
35 281:544 (1979)], alkaline phosphatase, the tryptophan (trp) promoter system [vector pATH1

(ATCC 37695) is designed to facilitate expression of an open reading frame as a trpE fusion protein under control of the trp promoter] and hybrid promoters such as the tac promoter (isolatable from plasmid pDR540 ATCC-37282). However, other functional bacterial promoters, whose nucleotide sequences are generally known, enable one of skill in the art to
5 ligate them to DNA encoding the proteins of the instant invention using linkers or adapters to supply any required restriction sites. Promoters for use in bacterial systems also will contain a Shine-Dalgarno sequence operably linked to the DNA encoding the desired polypeptides. These examples are illustrative rather than limiting.

The proteins of this invention may be synthesized either by direct expression or as
10 a fusion protein comprising the protein of interest as a translational fusion with another protein or peptide which may be removable by enzymatic or chemical cleavage. It is often observed in the production of certain peptides in recombinant systems that expression as a fusion protein prolongs the lifespan, increases the yield of the desired peptide, or provides a convenient means of purifying the protein of interest. A variety of peptidases (e.g. enterokinase and thrombin)
15 which cleave a polypeptide at specific sites or digest the peptides from the amino or carboxy termini (e.g. diaminopeptidase) of the peptide chain are known. Furthermore, particular chemicals (e.g. cyanogen bromide) will cleave a polypeptide chain at specific sites. The skilled artisan will appreciate the modifications necessary to the amino acid sequence (and synthetic or semi-synthetic coding sequence if recombinant means are employed) to incorporate site-specific
20 internal cleavage sites. See e.g., P. Carter, "Site Specific Proteolysis of Fusion Proteins", Chapter 13 in Protein Purification: From Molecular Mechanisms to Large Scale Processes, American Chemical Society, Washington, D.C. (1990).

In addition to cloning and expressing the genes of interest in the prokaryotic systems discussed above, the proteins of the present invention may also be produced in
25 eukaryotic systems. The present invention is not limited to use in a particular eukaryotic host cell. A variety of eukaryotic host cells are available from depositories such as the American Type Culture Collection (ATCC) and are suitable for use with the vectors of the present invention. The choice of a particular host cell depends to some extent on the particular expression vector used to drive expression of the human glutamate receptor-encoding nucleic
30 acids of the present invention.

A preferred host cell line employed in this invention is the widely available cell line HEK293 (hereinafter referred to as "HEK293" or "293"). This cell line is available from the American Type Culture Collection under the accession number CRL-1573.

Cell lines, such as HEK293, produce glutamate endogenously. As a result, some
35 amounts of glutamate are secreted into the culture medium, thereby making it somewhat difficult

to express and study glutamate receptors in such cell lines due to the activation of the transfected receptor. However, measures can be taken to reduce the effects of this glutamate release on the functional characterization of the receptor. These include but are not limited to the use of neurotransport protein. For example, cell lines can be produced to express a plasmid in which the rat glutamate transporter gene (GLAST) is expressed. The glutamate levels in 24 hour medium of a cell line expressing GLAST will reduce the amount of glutamate in the medium to less than 3 micromolar, thus reducing the basal activation of the receptor and/or desensitization or the requirement for extensive washing to remove residual glutamate before assay procedures. See Storck, et al, Proc. Nat'l Acad. Sci. USA, 89:10955-59 (Nov. 1992) and Desai et al, Molecular Pharmacology, 48:648-657 (1995). Other measures also are helpful in dealing with the endogenous glutamate production. For instance, plating the cells on fibronectin coated plates and exchanging out the media 16 hours before the assay is performed are helpful in reducing and normalizing the effects of the release of glutamate to the media.

In addition to the lipid transfection method for incorporating the novel nucleic acid sequences (see Example 2 below), a wide variety of vectors, some of which are discussed below, exist for the transformation of mammalian host cells such as those described above. Thus, the present invention also relates to vectors which comprise a polynucleotide or polynucleotides of the present invention, and host cells which are genetically engineered with vectors of the invention and to the production of polypeptides of the invention by recombinant techniques. In another embodiment, the present invention relates to a recombinant DNA molecule comprising, 5' to 3', a promoter effective to initiate transcription in a host cell and the above-described invention nucleic acid molecule(s).

Cell-free translation systems can also be employed to produce such proteins using RNAs derived from the DNA constructs of the present invention.

Incorporation of cloned DNA into a suitable expression vector, transfection of eukaryotic cells with a plasmid vector or a combination of plasmid vectors, each encoding one or more distinct genes or with linear DNA, and selection of transfected cells are well known in the art (see, e.g., Sambrook et al. (1989) Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor Laboratory Press). Suitable means for introducing (transducing) expression vectors containing invention nucleic acid constructs into host cells to produce transduced recombinant cells (i.e., cells containing recombinant heterologous nucleic acid) are well known in the art (see, for review, Friedmann, 1989, Science, 244:1275-1281; Mulligan, 1993, Science, 260:926-932, each of which are incorporated herein by reference in their entirety).

As referred to above, exemplary methods of transduction include, e.g., infection employing viral vectors (see, e.g., U.S. Pat. No. 4,405,712 and 4,650,764), calcium phosphate

transfection (U.S. Pat. Nos. 4,399,216 and 4,634,665), dextran sulfate transfection, electroporation, lipofection (see, e.g., U.S. Pat. Nos. 4,394,448 and 4,619,794), cytofection, particle bead bombardment, and the like. The heterologous nucleic acid can optionally include sequences which allow for its extrachromosomal (i.e., episomal) maintenance, or the
5 heterologous nucleic acid can be donor nucleic acid that integrates into the genome of the host. Recombinant cells can then be cultured under conditions whereby the mutant mGluR2 and mGluR3 sequences encoded by the DNA is (are) expressed. Preferred cells include mammalian cells (e.g., HEK 293, CHO and Ltk⁻ cells), yeast cells (e.g., methylotrophic yeast cells, such as *Pichia pastoris*), bacterial cells (e.g., *Escherichia coli*), and the like.

10 Expression vectors for use in carrying out the present invention will comprise a promoter capable of directing the transcription of a cloned DNA and a transcriptional terminator.

Also contained in the expression vectors is a polyadenylation signal located downstream of the coding sequence of interest. Polyadenylation signals include the early or late polyadenylation signals from SV40 (Kaufman and Sharp, *ibid.*), the polyadenylation signal from
15 the Adenovirus 5 E1B region and the human growth hormone gene terminator (DeNoto et al., *Nuc. Acid Res.* 9: 3719-3730, 1981). The expression vectors may include a noncoding viral leader sequence, such as the Adenovirus 2 tripartite leader, located between the promoter and the RNA splice sites. Preferred vectors may also include enhancer sequences, such as the SV40 enhancer and the mouse μ enhancer (Gillies, *Cell* 33: 717-728, 1983). Expression vectors may
20 also include sequences encoding the adenovirus VA RNAs.

Suitable expression vectors are well known in the art, and include vectors capable of expressing DNA operatively linked to a regulatory sequence, such as a promoter region that is capable of regulating expression of such DNA. Thus, an expression vector refers to a recombinant DNA or RNA construct, such as a plasmid, a phage, recombinant virus or other
25 vector that, upon introduction into an appropriate host cell, results in expression of the inserted DNA. Appropriate expression vectors are well known to those of skill in the art and include those that are replicable in eukaryotic cells and/or prokaryotic cells and those that remain episomal or those which integrate into the host cell genome.

Exemplary expression vectors for transformation of *E. coli* prokaryotic cells
30 include the pET expression vectors (Novagen, Madison, Wis., see U.S. Pat. No. 4,952,496), e.g., pET11a, which contains the T7 promoter, T7 terminator, the inducible *E. coli* lac operator, and the lac repressor gene; and pET 12a-c, which contains the T7 promoter, T7 terminator, and the *E. coli* ompT secretion signal. Another such vector is the pIN-IIIompA2 (see Duffaud et al., *Meth. in Enzymology*, 153:492-507, 1987), which contains the lpp promoter, the lacUV5 promoter
35 operator, the ompA secretion signal, and the lac repressor gene.

Exemplary eukaryotic expression vectors include eukaryotic cassettes, such as the pSV-2 gpt system (Mulligan et al., 1979, Nature, 277:108-114); the Okayama-Berg system (Mol. Cell Biol., 2:161-170), and the expression-cloning vector described by Genetics Institute (1985, Science, 228:810-815). Each of these plasmid vectors is capable of promoting expression of the invention chimeric protein of interest.

Also provided are nucleic acid molecule(s) comprising a transcriptional region functional in a cell, a sequence complimentary to an RNA sequence encoding an amino acid sequence corresponding to the herein-disclosed mutant mGluR2 and mGluR3 sequences, and a transcriptional termination region functional in a suitable host cell.

A wide variety of transcriptional and translational regulatory sequences may be employed, depending upon the nature of the host. The transcriptional and translational regulatory signals may be derived from viral sources, such as adenovirus, bovine papilloma virus, cytomegalovirus, simian virus, or the like, where the regulatory signals are associated with a particular gene sequence which has a high level of expression. Alternatively, promoters from mammalian expression products, such as actin, collagen, myosin, and the like, may be employed. Transcriptional initiation regulatory signals may be selected which allow for repression or activation, so that expression of the gene sequences can be modulated. Of interest are regulatory signals which are temperature-sensitive so that by varying the temperature, expression can be repressed or initiated, or are subject to chemical (such as metabolite) regulation. A favored promoter is the promoter from the Cytomegalovirus (CMV).

Thus, an embodiment provides are transformed host cells that recombinantly express the herein disclosed mutant mGluR2 and mGluR3 sequences of the invention.

As used herein, a cell is said to be "altered to express a desired peptide" when the cell, through genetic manipulation, is made to produce a protein which it normally does not produce or which the cell normally produces at lower levels. One skilled in the art can readily adapt procedures for introducing and expressing either genomic, cDNA, or synthetic sequences into either eukaryotic or prokaryotic cells.

A nucleic acid molecule, such as DNA, is said to be "capable of expressing" a polypeptide if it contains nucleotide sequences which contain transcriptional and translational regulatory information and such sequences are "operably linked" to nucleotide sequences which encode the polypeptide. An operable linkage is a linkage in which the regulatory DNA sequences and the DNA sequence sought to be expressed are connected in such a way as to permit gene sequence expression. The precise nature of the regulatory regions needed for gene sequence expression may vary from organism to organism, but shall in general include a promoter region which, in prokaryotes, contains both the promoter (which directs the initiation

of RNA transcription) as well as the DNA sequences which, when transcribed into RNA, will signal synthesis initiation. Such regions will normally include those 5'-non-coding sequences involved with initiation of transcription and translation, such as the TATA box, capping sequence, CAAT sequence, and the like.

5 If desired, the non-coding region 3' to the sequence encoding an mGluR2 or mGluR3 gene may be obtained by the above-described methods. This region may be retained for its transcriptional termination regulatory sequences, such as termination and polyadenylation. Thus, by retaining the 3'-region naturally contiguous to the DNA sequence encoding a mutant mGluR2 and mGluR3 sequences, the transcriptional termination signals may be provided.

10 Where the transcriptional termination signals are not satisfactorily functional in the expression host cell, then a 3' region functional in the host cell may be substituted.

Two DNA sequencers (such as a promoter region sequence and an a mutant mGluR2 or mGluR3 encoding sequence) are said to be "operably linked" if the nature of the linkage between the two DNA sequences does not (1) result in the introduction of a frame-shift mutation, (2) interfere with the ability of the promoter region sequence to direct the transcription of the mutant mGluR2 or mGluR3 encoding gene sequence, or (3) interfere with the ability of the mutant mGluR2 or mGluR3 encoding sequence to be transcribed by the promoter region sequence. Thus, a promoter region would be operably linked to a DNA sequence if the promoter were capable of effecting transcription of that DNA sequence.

20 The selection of control sequences, expression vectors, transformation methods, and the like, are dependent on the type of host cell used to express the gene. As used herein, "cell", "cell line", and "cell culture" may be used interchangeably and all such designations include progeny.

25 The term "transformants" or "transformed cells" include the primary subject cell and cultures derived therefrom, without regard to the number of transfers. It is also understood that all progeny may not be precisely identical in DNA content, due to deliberate or inadvertent mutations. However, as defined, mutant progeny have the same functionality as that of the originally transformed cell.

30 To express the human mutant mGluR2 and mGluR3 sequences-encoding gene of the invention, transcriptional and translational signals recognized by an appropriate host are necessary. The present invention encompasses the expression of the mutant mGluR2 or mGluR3 sequences in either prokaryotic or eukaryotic cells.

Host cells which may be used in the expression systems of the present invention are not strictly limited, provided that they are suitable for use in the expression of the novel

human mutant mGluR2 and mGluR3 sequences of the invention. Suitable hosts may often include eukaryotic cells, such as HEK-293 and other suitable mammalian cell lines.

Also, as noted above, representative examples of appropriate host cells for use in practicing the present invention include bacterial cells, such as streptococci, staphylococci, E. coli, Streptomyces and Bacillus subtilis cells. Also, fungal cells, such as yeast cells and Aspergillus cells; insect cells such as Drosophila S2 and Spodoptera Sf9 cells; animal cells such as CHO, COS, HeLa, C127, 3T3, BHK, HEK 293 and Bowes melanoma cells; and plant cells may be used.

Fungal cells, including species of yeast (e.g., Saccharomyces spp., particularly S. cerevisiae, Schizosaccharomyces spp.) or filamentous fungi (e.g., Aspergillus spp., Neurospora spp.) may be used as host cells within the present invention. Suitable yeast vectors for use in the present invention include YRp7 (Struhl et al., Proc. Natl. Acad. Sci. USA. 76: 1035-1039, 1978), YEpl3 (Broach et al., Gene 8: 121-133, 1979), POT vectors (Kawasaki et al, U.S. Pat. No. 4,931,373, which is incorporated by reference herein), pJDB249 and pJDB219 (Beggs, Nature 275:104-108, 1978) and derivatives thereof. Such vectors will generally include a selectable marker, which may be one of any number of genes that exhibit a dominant phenotype for which a phenotypic assay exists to enable transformants to be selected. Preferred selectable markers are those that complement host cell auxotrophy, provide antibiotic resistance or enable a cell to utilize specific carbon sources, and include LEU2 (Broach et al., ibid.), URA3 (Botstein et al., Gene 8: 17, 1979), HIS3 (Struhl et al., ibid.) or POT1 (Kawasaki et al., ibid.). Another suitable selectable marker is the CAT gene, which confers chloramphenicol resistance on yeast cells.

Any of a series of yeast gene sequence expression systems can be utilized which incorporate promoter and termination elements from the actively expressed gene sequences coding for glycolytic enzymes are produced in large quantities when yeast are grown in mediums rich in glucose. Known glycolytic gene sequences can also provide very efficient transcriptional control signals.

Yeast provides substantial advantages in that it can also carry out post-translational peptide modifications. A number of recombinant DNA strategies exist which utilize strong promoter sequences and high copy number of plasmids which can be utilized for production of the desired proteins in yeast. Yeast recognizes leader sequences on cloned mammalian gene sequence products and secretes peptides bearing leader sequences (i.e., pre-peptides). For a mammalian host, several possible vector systems are available for the expression of the mutant mGluR2 and mGluR3 sequences.

A variety of higher eukaryotic cells may serve as host cells for expression of the polypeptides of the invention, although not all cell lines will be capable of functional coupling of

the receptor to the cell's second messenger systems. Cultured mammalian cells, such as BHK, CHO, Y1 (Shapiro et al., TIPS Suppl. 43-46 (1989)), NG108-15 (Dawson et al., Neuroscience Approached Through Cell Culture, Vol. 2, pages 89-114 (1989)), N1E-115 (Liles et al., J. Biol. Chem. 261:5307-5313 (1986)), PC 12 and COS-1 (ATCC CRL 1650) are preferred. Preferred
5 BHK cell lines are the tk.sup.-- ts13 BHK cell line (Waechter and Baserga, Proc. Natl. Acad. Sci. USA 79:1106-1110 (1982)) and the BHK 570 cell line (deposited with the American Type Culture Collection, 12301 Parklawn Dr., Rockville, Md. under accession number CRL 10314). A tk.sup.-- BHK cell line is available from the ATCC under accession number CRL 1632.

Prokaryotic hosts are, generally, very efficient and convenient for the production
10 of recombinant proteins and are, therefore, one type of preferred expression system for the expressing the mutant mGluR2 and mGluR3 sequences encoding gene.

Prokaryotes most frequently are represented by various strains of *E. coli*. However, other microbial strains may also be used, including other bacterial strains. In prokaryotic systems, plasmid vectors that contain replication sites and control sequences derived
15 from a species compatible with the host may be used. Examples of suitable plasmid vectors may include pBR322, pUC-118, pUC119 and the like; suitable phage or bacteriophage vectors may include .gamma.gt10, .gamma.gt11 and the like; and suitable virus vectors may include pMAM-neo, pKRC and the like. Preferably, the selected vector of the present invention has the capacity to replicate in the selected host cell.

Recognized prokaryotic hosts include bacteria such as *E. coli*, *Bacillus*, *Streptomyces*, *Pseudomonas*, *Salmonella*, *Serratia*, and the like. However, under such
20 conditions, the peptide will not be glycosylated. The prokaryotic host must be compatible with the replicon and control sequences in the expression plasmid.

To express the mutant mGluR2 and mGluR3 sequences (or a functional derivative
25 thereof) in a prokaryotic cell, it is necessary to operably link the mutant mGluR2 or mGluR3 encoding nucleotide sequence to a functional prokaryotic promoter. Such promoters may be either constitutive or, more preferably, regulatable (i.e., inducible or derepressible). Examples of constitutive promoters and inducible promoters are well known to a skilled artisan. Prokaryotic promoters are reviewed by Cenatiempo (Biochimie 68:505-516 (1986)); and Gottesman (Ann.
30 Rev. Genet. 18:415-442 (1984)). Proper expression in a prokaryotic cell also requires the presence of a ribosome-binding site upstream of the gene sequence-encoding sequence. Such ribosome binding sites are disclosed, for example, by Gold et al., Ann. Rev. Microbiol. 35:365-404 (1981).

As used herein, the term "promoter" refers to a polynucleotide sequence,
35 preferably a DNA sequence, that regulates expression of a selected DNA sequence operably

linked to the promoter, and which effects expression of the selected DNA sequence in cells. The term encompasses "tissue specific" promoters, i.e. promoters, which effect expression of the selected DNA sequence only in specific cells (e.g. cells of a specific tissue). The term also covers so-called "leaky" promoters, which regulate expression of a selected DNA primarily in one tissue, but cause expression in other tissues as well. The term also encompasses non-tissue specific promoters and promoters that constitutively express or that are inducible (i.e. expression levels can be controlled).

A mutant mGluR2 and mGluR3 sequences encoding nucleic acid molecule and an operably linked promoter may be introduced into a recipient prokaryotic or eukaryotic cell either as a nonreplicating DNA (or RNA) molecule, which may either be a linear molecule or, more preferably, a closed covalent circular molecule. Since such molecules are incapable of autonomous replication, the expression of the gene may occur through the transient expression of the introduced sequence. Alternatively, permanent expression may occur through the integration of the introduced DNA sequence into the host chromosome.

In one embodiment, a vector is employed which is capable of integrating the desired gene sequences into the host cell chromosome. Cells which have stably integrated the introduced DNA into their chromosomes can be selected by also introducing one or more markers which allow for selection of host cells which contain the expression vector. The marker may provide for prototrophy to an auxotrophic host, biocide resistance, e.g., antibiotics, or heavy metals, such as copper, or the like. The selectable marker gene sequence can either be directly linked to the DNA gene sequences to be expressed, or introduced into the same cell by co-transfection. Additional elements may also be needed for optimal synthesis of single chain binding protein mRNA. These elements may include splice signals, as well as transcription promoters, enhancers, and termination signals. cDNA expression vectors incorporating such elements include those described by Okayama, *Molec. Cell. Biol.* 3:280 (1983).

In a preferred embodiment, the introduced nucleic acid molecule will be incorporated into a plasmid or viral vector capable of autonomous replication in the recipient host. Any of a wide variety of vectors may be employed for this purpose. Factors of importance in selecting a particular plasmid or viral vector include: the ease with which recipient cells that contain the vector may be recognized and selected from those recipient cells which do not contain the vector; the number of copies of the vector which are desired in a particular host; and whether it is desirable to be able to "shuttle" the vector between host cells of different species.

Preferred prokaryotic vectors include plasmids such as those capable of replication in *E. coli* (such as, for example, pBR322, ColEI, pSC101, pACYC 184, .pi.VX. Such plasmids are, for example, disclosed by Sambrook (cf. *Molecular Cloning: A Laboratory*

Manual, second edition, edited by Sambrook, Fritsch, & Maniatis, Cold Spring Harbor Laboratory, (1989)). *Bacillus* plasmids include pC194, pC221, pT127, and the like. Such plasmids are disclosed by Gryczan (In: *The Molecular Biology of the Bacilli*, Academic Press, N.Y. (1982), pp. 307-329). Suitable *Streptomyces* plasmids include p1J101 (Kendall et al., J. Bacteriol. 169:4177-4183 (1987)), and *streptomyces* bacteriophages such as .phi.C31 (Chater et al., In: *Sixth International Symposium on Actinomycetales Biology*, Akademiai Kiado, Budapest, Hungary (1986), pp. 45-54). *Pseudomonas* plasmids are reviewed by John et al. (Rev. Infect. Dis. 8:693-704 (1986)), and Izaki (Jpn. J. Bacteriol. 33:729-742 (1978)).

As noted, supra, expression of the mutant mGluR2 and mGluR3 sequences in eukaryotic hosts requires the use of eukaryotic regulatory regions. Such regions will, in general, include a promoter region sufficient to direct the initiation of RNA synthesis. Preferred eukaryotic promoters include, for example, the Cytomegalovirus Promoter (CMV), the promoter of the mouse metallothionein I gene sequence (Hamer et al., J. Mol. Appl. Gen. 1:273-288 (1982)); the TK promoter of Herpes virus (McKnight, Cell 31:355-365 (1982)); the SV40 early promoter (Benoist et al., Nature (London) 290:304-310(1981)); the yeast gal4 gene sequence promoter (Johnston et al., Proc. Natl. Acad. Sci. (USA) 79:6971-7975 (1982); Silver et al., Proc. Natl. Acad. Sci. (USA) 81:5951-5955 (1984)).

As is widely known, translation of eukaryotic mRNA is initiated at the colon which encodes the first methionine. For this reason, it is preferable to ensure that the linkage between a eukaryotic promoter and a DNA sequence which encodes the mutant mGluR2 and mGluR3 sequences (or a functional derivative thereof) does not contain any intervening codons which are capable of encoding a methionine (i.e., AUG). The presence of such codons results either in a formation of a fusion protein (if the AUG codon is in the same reading frame as the mutant mGluR2 or mGluR3 coding sequence) or a frame-shift mutation (if the AUG codon is not in the same reading frame as the mutant mGluR2 or mGluR3 coding sequence).

Preferred eukaryotic plasmids include, for example, BPV, vaccinia, SV40, 2-micron circle, and the like, or their derivatives. Such plasmids are well known in the art (Botstein et al., Miami Wntr. Symp. 19:265-274 (1982); Broach, In: *The Molecular Biology of the Yeast Saccharomyces: Life Cycle and Inheritance*, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., p. 445-470 (1981); Broach, Cell 28:203-204 (1982); Bollon et al., J. Clin. Hematol. Oncol. 10:39-48 (1980); Maniatis, In: *Cell Biology: A Comprehensive Treatise*, Vol. 3, Gene Sequence Expression, Academic Press, N.Y., pp. 563-608 (1980).

Once the vector or nucleic acid molecule containing the construct(s) has been prepared for expression, the DNA construct(s) may be introduced into an appropriate host cell by any of a variety of suitable means, i.e., transformation, transfection, conjugation, protoplast

fusion, electroporation, particle gun technology, calcium phosphate-precipitation, direct microinjection, and the like. After the introduction of the vector, recipient cells are grown in a selective medium, which selects for the growth of vector-containing cells. Expression of the cloned gene molecule(s) results in the production of the herein-disclosed mutant mGluR2 and mGluR3 sequences or biologically active fragments thereof. This can take place in the transformed cells as such, or following the induction of these cells to differentiate (for example, by administration of bromodeoxyuracil to neuroblastoma cells or the like).

A variety of incubation conditions can be used to form the peptide of the present invention. Preferred conditions include those which mimic physiological conditions.

An example of the means for preparing the mutant mGluR2 and mGluR3 sequences of the invention is to express nucleic acids encoding the mutant mGluR2 and mGluR3 sequences in a suitable host cell, such as a bacterial cell, a yeast cell, an amphibian cell (i.e., oocyte), or a mammalian cell, using methods well known in the art, and recovering the expressed polypeptide, again using well-known methods.

Using methods such as northern blot or slot blot analysis, transfected cells that contain mutant mGluR2 and mGluR3 sequences encoding DNA or RNA can be selected. Transfected cells can also be analyzed to identify those that express the mutant mGluR2 and mGluR3 sequences. Analysis can be carried out, for example, by using any of well known screening assays attending a functional receptor, and comparing the values obtained to a control, untransfected host cells by electrophysiologically monitoring the currents through the cell membrane in response to mutant mGluR2 and mGluR3 sequences, and the like. Mutant mGluR2 and mGluR3 sequences(s) can be isolated directly from cells that have been transformed with expression vectors comprising nucleic acid encoding the mutant mGluR2 and mGluR3 sequences or fragments/portions thereof.

Nucleic acid molecules may be stably incorporated into cells or may be transiently introduced using methods known in the art. Stably transfected mammalian cells may be prepared by transfecting cells with an expression vector comprising a sequence of nucleotides that encodes the mutant mGluR2 and mGluR3 sequences in conjunction with a selectable marker gene (such as, for example, the gene for thymidine kinase, dihydrofolate reductase, neomycin resistance, and the like), and growing the transfected cells under conditions selective for cells expressing the marker gene. To prepare transient transfectants, mammalian cells are transfected with a reporter gene (such as the *E. coli* β -galactosidase gene) to monitor transfection efficiency. The precise amounts and ratios of DNA encoding the mutant mGluR2 and mGluR3 sequences may be empirically determined and optimized for a particular cells and assay conditions. Selectable marker genes are typically not included in the transient transfections because the

transfectants are typically not grown under selective conditions, and are usually analyzed within a few days after transfection.

In order to identify cells that have integrated the cloned DNA, a selectable marker is generally introduced into the cells along with the gene or cDNA of interest. Preferred
5 selectable markers for use in cultured mammalian cells include genes that confer resistance to drugs, such as neomycin, hygromycin, and methotrexate. The selectable marker may be an amplifiable selectable marker. Preferred amplifiable selectable markers are the DHFR gene and the neomycin resistance gene. Selectable markers are reviewed by Thilly (Mammalian Cell Technology, Butterworth Publishers, Stoneham, MA, which is incorporated herein by reference).
10 The choice of selectable markers is well within the level of ordinary skill in the art.

Selectable markers may be introduced into the cell on a separate plasmid at the same time as the gene of interest, or they may be introduced on the same plasmid. If on the same plasmid, the selectable marker and the gene of interest may be under the control of different promoters or the same promoter, the latter arrangement producing a dicistronic message.

15 Constructs of this type are known in the art (for example, Levinson and Simonsen, U.S. Pat. No. 4,713,339). It may also be advantageous to add additional DNA, known as "carrier DNA" to the mixture which is introduced into the cells.

In particularly preferred aspects, eukaryotic cells which contain heterologous DNAs express such DNA and form recombinant mutant mGluR2 and mGluR3 sequences. In
20 more preferred aspects, recombinant mutant mGluR2 and mGluR3 sequences activity is readily detectable because it is a type that is absent from the untransfected host cell.

Heterologous DNA may be maintained in the cell as an episomal element or may be integrated into chromosomal DNA of the cell. The resulting recombinant cells may then be cultured or subcultured (or passaged, in the case of mammalian cells) from such a culture or a
25 subculture thereof. Methods for transfection, injection and culturing recombinant cells are known to the skilled artisan. Similarly, the mutant mGluR2 and mGluR3 sequences(s) may be purified using protein purification methods known to those of skill in the art. For example, antibodies or other ligands that specifically bind to the mutant mGluR2 and mGluR3 sequences may be used for affinity purification of the mutant mGluR2 and mGluR3 sequences.

30 As used herein, "heterologous or foreign DNA and/or RNA" are used interchangeably and refer to DNA or RNA that does not occur naturally as part of the genome of the cell in which it is present or to DNA or RNA which is found in a location or locations in the genome that differ from that in which it occurs in nature. Typically, heterologous or foreign DNA and RNA refer to DNA or RNA that is not endogenous to the host cell and has been

artificially introduced into the cell. Examples of heterologous DNA include DNA disclosed herein.

In other embodiments, mRNA may be produced by in vitro transcription of DNA encoding the mutant mGluR2 and mGluR3 sequences. This mRNA can then be injected into
5 Xenopus oocytes where the RNA directs the synthesis of the mutant mGluR2 and mGluR3 sequences. Alternatively, the invention-encoding DNA can be directly injected into oocytes for expression of a functional mutant mGluR2 and mGluR3 sequences. The transfected mammalian cells or injected oocytes may then be used in the methods of drug screening provided herein.

Alternatively, the invention DNA sequences can be transcribed into RNA, which
10 can then be transfected into amphibian cells for translation into protein. Suitable amphibian cells include Xenopus oocytes.

Practitioners of this invention realize that, in addition to the above-mentioned expression systems, the cloned cDNA may also be employed in the production of transgenic animals in which a test mammal, usually a mouse, in which the effects of the expression of the
15 proteins of the present invention can be assessed, particularly with regard to the interaction of such proteins and modulators or portions of modulators. For instance, without being limited by such example, the nucleic acids of the present invention may be employed in the construction of "knockin" animals in which the expression of a mutant mGluR2 or mGluR3 is expressed. Once such test animals are prepared, evaluations are conducted that evaluate the effects of modulator
20 compounds. For instance, a modulator development program, based on rational drug design using SAR, may have a putative highly selective modulator design. Structural variations of this design, and the hypothetical optimal design, each are evaluated in a number of different types of knock-in mice, where each knock-in mouse type carries a different mutation combination of the S688L, G689V and N735D mutants. This data is used to verify, refine the design of, and
25 evaluate the putative highly selective modulator in a whole organism model. An important feature of such evaluations is the ability to evaluate and assess "off-target" effects of the modulators, i.e., effects at other organs, or unexpected behavioral effects.

Skilled artisans also recognize that some alterations of SEQ.ID.NOS.:1-8 will fail to change the function of the respective amino acid compound. For instance, some hydrophobic
30 amino acids may be exchanged for other hydrophobic amino acids. Those altered amino acid compounds which confer substantially the same function in substantially the same manner as the exemplified amino acid compound are also encompassed within the present invention. Typically such conservative substitutions attempt to preserve the: (a) secondary or tertiary structure of the polypeptide backbone; (b) the charge or hydrophobicity of the residue; or (c) the bulk of the side
35 chain. Some examples of such conservative substitutions of amino acids, resulting in the

production of proteins which may be functional equivalents of the protein of SEQ.ID.NOS.:1-8 are shown in TABLE II, *infra*.

TABLE II

	Original Residue	Exemplary Substitutions
5	Ala	Ser, Gly
	Arg	Lys
	Asn	Gln, His
	Asp	Glu
10	Cys	Ser
	Gln	Asn
	Glu	Asp
	Gly	Pro, Ala
	His	Asn, Gln
15	Ile	Leu, Val
	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	Met, Leu, Tyr
20	Ser	Thr
	Thr	Ser
	Trp	Tyr
	Tyr	Trp, Phe
	Val	Ile, Leu

These substitutions may be introduced into the protein in a variety of ways, such as during the chemical synthesis or by chemical modification of an amino acid side chain after the protein has been prepared.

Alterations of the protein having a sequence which corresponds to the sequence of SEQ.ID.NOS.:1-8 also may be induced by alterations of the nucleic acid compounds which encode these proteins, i.e., SEQ.ID.NOS.:9-16. These mutations of the nucleic acid compound may be generated by either random mutagenesis techniques, such as those techniques employing chemical mutagens, or by site-specific mutagenesis employing oligonucleotides. Those nucleic acid compounds which confer substantially the same function in substantially the same manner as the exemplified nucleic acid compounds are also encompassed within the present invention.

Other embodiments of the present invention are nucleic acid compounds which comprise isolated nucleic acid sequences which encode any of the polypeptide sequences of SEQ.ID.NOS.:1-8. As skilled artisans will recognize, the amino acid compounds of the invention can be encoded by a multitude of different nucleic acid sequences because most of the amino acids are encoded by more than one nucleic acid triplet due to the degeneracy of the amino acid code. Because these alternative nucleic acid sequences would encode the same amino acid sequences, the present invention further comprises these alternate nucleic acid sequences.

The nucleic acid sequences encoding the mutant mGluR α receptor molecules of the present invention may be produced using synthetic methodology. This synthesis of nucleic acids is well known in the art. See, e.g., E. L. Brown, R. Belagaje, M. J. Ryan, and H. G. Khorana, *Methods in Enzymology*, 68:109-151 (1979). The DNA segments corresponding to the receptor gene are generated using conventional DNA synthesizing apparatus such as the Applied Biosystems Model 380A or 380B DNA synthesizers (commercially available from Applied Biosystems, Inc., 850 Lincoln Center Drive, Foster City, Calif. 94404) which employ phosphoramidite chemistry. In the alternative, the more traditional phosphotriester chemistry may be employed to synthesize the nucleic acids of this invention. [(See, e.g., M. J. Gait, ed., *Oligonucleotide Synthesis, A Practical Approach*, (1984).]

Any of such synthetic human mGluR α nucleic acid sequences (e.g., synthetic genes) may be designed to possess restriction endonuclease cleavage sites at either end of the transcript to facilitate isolation from and integration into expression and amplification plasmids. The choice of restriction sites are chosen so as to properly orient the coding sequence of the receptor with control sequences to achieve proper in-frame reading and expression of the human mGluR α receptor molecule. A variety of other such cleavage sites may be incorporated depending on the particular plasmid constructs employed and may be generated by techniques well known in the art.

In an alternative methodology, the desired DNA sequences can be generated using the polymerase chain reaction as described in U.S. Pat. No. 4,889,818, which is incorporated herein by reference.

In addition to the deoxyribonucleic acid of SEQ.ID.NOS.:9-16, this invention also provides ribonucleic acids (RNA) that likewise express the mGluR2 mutants designated as SEQ.ID.NOS.:1-8, and conservatively modified variants thereof. As is known to those of skill in the art, the DNA sequences provided herein are convertible to corresponding RNA sequences via pyrimidine base substitution of uracil (U) for thymine (T).

The ribonucleic acids of the present invention may be prepared using the polynucleotide synthetic methods discussed supra or they may be prepared enzymatically using RNA polymerases to transcribe a DNA template.

One system for preparing the ribonucleic acids of the present invention employs the RNA polymerase from the bacteriophage T7 or the bacteriophage SP6. Both of these RNA polymerases are highly specific and require the insertion of bacteriophage-specific sequences at the 5' end of the message to be read. See, J. Sambrook, et al., supra, at 18.82-18.84.

This invention also provides nucleic acids, RNA or DNA, which are complementary to SEQ.ID.NOS.:9-16.

The present invention also provides probes and primers useful for molecular biology techniques. A compound which encodes for SEQ.ID.NOS.:9-16, or a complementary sequence of SEQ.ID.NOS.: 9-16, or a fragment thereof, and which is at least 20-50 base pairs in length, and which will selectively hybridize to human genomic DNA or messenger RNA encoding a human glutamate receptor, is provided. Preferably, the 25 or more base pair compound is DNA or a length sufficient to hybridize.

The term "selectively hybridize" as used herein may refer to either of two situations. In the first such embodiment of this invention, the nucleic acid compounds described supra hybridize to a human glutamate receptor under more stringent hybridization conditions than these same nucleic acid compounds would hybridize to an analogous glutamate receptor of another species, e.g. rodent. In the second such embodiment of this invention, these probes hybridize to the mutant mGluR2 or mGluR3 receptor molecules under more stringent hybridization conditions than other related compounds, including nucleic acid sequences encoding other glutamate receptors of the same species. A more detailed explanation of hybridization and varying stringency of hybridization, which pertain to the two types of hybridization described here, is provided in the "Definitions and Terms of Art" section supra.

These probes and primers can be prepared enzymatically as described supra. These probes and primers also are synthesized using chemical means as described supra. Alternately, these probes and primers of defined structure may also be purchased commercially.

Another aspect of the present invention is recombinant DNA cloning vectors and expression vectors comprising the nucleic acid sequences of the present invention. Many of the vectors encompassed within this invention are described above. The preferred nucleic acid vectors are those which are DNA. A preferred recombinant DNA vector comprises the isolated DNA sequence SEQ.ID.NO.:14, bearing the codons for all three single amino acid substitutions on the respective mutant mGluR2 sequence. This sequence has been shown to provide the strongest depotentiation. However, it is fully appreciated that when using the compositions of

the present invention to help identify desired structures and sequences for effective modulators of mGluR2, the isolated nucleic acid sequences encoding a mutant mGluR2 with a single amino acid substitution, e.g., SEQ.ID.NO.:8-10, and the sequences encoding a mutant mGluR2 with two amino acid substitutions, e.g., SEQ.ID.NO.:11-13, are valuable when used in concert. Any
5 of these are suitable for transfection into cells by use of vectors, as described herein, and by other means of transfection, such as by lipid transfection as in Example 2 below.

The skilled artisan understands that the type of cloning vector or expression vector employed depends upon the availability of appropriate restriction sites, the type of host cell in which the vector is to be transfected or transformed, the purpose of the transfection or
10 transformation (e.g., transient expression in an oocyte system, stable transformation as an extrachromosomal element, or integration into the host chromosome), the presence or absence of readily assayable markers (e.g., antibiotic resistance markers, metabolic markers, or the like), and the number of copies of the gene to be present in the cell.

The type of vector employed to carry the nucleic acids of the present invention
15 may be RNA viruses, DNA viruses, lytic bacteriophages, lysogenic bacteriophages, stable bacteriophages, plasmids, viroids, and the like. The most preferred vectors of the present invention are those derived from plasmids.

When preparing an expression vector the skilled artisan understands that there are many variables to be considered. One such example is the use of a constitutive promoter, i.e. a
20 promoter which is functional at all times, instead of a regulatable promoter which may be activated or inactivated by the artisan using heat, addition or removal of a nutrient, addition of an antibiotic, and the like. The practitioner also understands that the amount of nucleic acid or protein to be produced dictates, in part, the selection of the expression system. For experiments examining the amount of the protein expressed on the cell membrane or for experiments
25 examining the biological function of an expressed membrane protein, for example, it may be unwise to employ an expression system which produces too much of the protein. The addition or subtraction of certain sequences, such as a signal sequence preceding the coding sequence, may be employed by the practitioner to influence localization of the resulting polypeptide. Such sequences added to or removed from the nucleic acid compounds of the present invention are
30 encompassed within this invention.

The plasmid pcDNA3.1 can be readily modified to construct expression vectors that produce mutant mGluR2 and mGluR3 receptors of the present invention in a variety of cells, including, for example, HEK-293.

One of the most widely employed techniques for altering a nucleic acid sequence
35 is by way of oligonucleotide-directed site-specific mutagenesis. B. Comack, "Current Protocols

in Molecular Biology", 8.01-8.5.9, (F. Ausubel, et al., eds. 1991). In this technique an oligonucleotide, whose sequence contains the mutation of interest, is synthesized as described supra. This oligonucleotide is then hybridized to a template containing the wild-type sequence. In a most preferred embodiment of this technique, the template is a single-stranded template. Particularly preferred are plasmids which contain regions such as the f1 intergenic region. This region allows the generation of single-stranded templates when a helper phage is added to the culture harboring the "phagemid".

After the annealing of the oligonucleotide to the template, a DNA-dependent DNA polymerase is then used to synthesize the second strand from the oligonucleotide, complementary to the template DNA. The resulting product is a heteroduplex molecule containing a mismatch due to the mutation in the oligonucleotide. After DNA replication by the host cell a mixture of two types of plasmid are present, the wild-type and the newly constructed mutant. This technique permits the introduction of convenient restriction sites such that the coding sequence may be placed immediately adjacent to whichever transcriptional or translational regulatory elements are employed by the practitioner.

The construction protocols utilized for *E. coli* can be followed to construct analogous vectors for other organisms, merely by substituting, if necessary, the appropriate regulatory elements using techniques well known to skilled artisans. Also, construction protocols for eukaryotic cells are widely known and employed by those skilled in the art.

Host cells which harbor the nucleic acids provided by the present invention are also provided. A preferred host cell is an *Xenopus* sp. oocyte which has been injected with RNA or DNA compounds of the present invention. Most preferred oocytes of the present invention are those which harbor a sense mRNA of the present invention. Other preferred host cells include HEK-293 cells which have been transfected and/or transformed with a vector which comprises a nucleic acid of the present invention.

The present invention also provides a method for constructing a recombinant host cell capable of expressing any of SEQ.ID.NOS.:9-16, said method comprising transforming a host cell with a recombinant DNA vector that comprises an isolated DNA sequence which encodes any of SEQ.ID.NOS.:9-16. The preferred host cell is HEK-293. A preferred vector for expression is one which comprises SEQ.ID.NO.:16. A preferred host cell for this method is HEK-293. An especially preferred expression vector in HEK-293 is one which comprises SEQ.ID.NO.:16.

Transformed host cells may be cultured under conditions well known to skilled artisans such that any of SEQ.ID.NOS.:9-16 is expressed, thereby producing a mutant of mGluR2 in the recombinant host cell, where such mutant alters the potentiation compared to

wild type mGluR2 with respect to modulators that normally bind to at least one allosteric binding site associated with the three amino acid substitutions believed to provide the most substantial alteration in effect.

Likewise, transformed host cells may be cultured under conditions well known to skilled artisans such that any of SEQ.ID.NOS.:41-48 is expressed, thereby producing a mutant of mGluR3 in the recombinant host cell, where such mutant alters the potentiation compared to wild type mGluR3 with respect to modulators that normally bind to at least one allosteric binding site associated with the three amino acid substitutions believed to provide the most substantial alteration in effect. Also with regard to mGluR3, mutant peptides of SEQ.ID.NOS.:33-40 are produced and used in manners disclosed herein for mutant mGluR2 peptides. With regard to the use of such mGluR3 mutant peptides and the nucleic acid sequences encoding them, it is believed that the polypeptide configuration of at least some of such mutants interacts with an allosteric modulator having its site in the general TM4/TM5 region in such a way as to increase the effect of such modulator.

The ability of glutamate to bind to an mGluRx receptor is essential in the development of a multitude of indications. In identifying and developing substances that act as allosteric modulators of a particular mGluRx receptor, such as mGluR2, it would be desirable, therefore, to determine those substances that bind to the allosteric binding site described herein. Generally, such an assay includes a method for determining whether a substance non-competitively affects the activity of the designated mGluRx receptor, and more specifically such an assay compares the effect of said substance on glutamate reception in assay systems using wild type mGluRx, and at least one of the mutant mGluRxs of the present invention. Typically a control of wild-type mGluRx with a known amount of glutamate, i.e., 1 mM, serves as a positive control against which other responses are normalized. For instance, a purported up-modulator of mGluR2 that acts at the allosteric binding site discovered herein is expected to raise the response with a small glutamate quantity (e.g., 3 or 10 μ M) in wild-type mGluR2, but is expected to have a diminished up-modulation effect in a treatment having a mutant mGluR2 where the mutant has at least one of any of the following amino acid substitutions: S688L; G689V; and N735D. This difference in modulation can be observed by comparing the effect of modulator Compounds A and B in Figure 1 with the lower effect of such modulators in Figures 2-6.

The instant invention provides such a screening system useful for discovering agents which allosterically modulate glutamate binding to the mGluR2 (or, alternately, the mGluR3) receptor, said screening system comprising the steps of:

a) preparing a human mutant mGluR2 mutant receptor, as with any of the sequences SEQ.ID.NOS.:9-16, and obtaining or preparing a wild-type human mGluR2 receptor;

b) transforming cells to express, separately, the mutant and wild-type mGluR2 receptors prepared in "a");

c) introducing glutamate at desired levels in each treatment and control;

d) exposing said human mGluR2 mutant and wild-type receptors (expressed per the transformations in "b)" above) to a potential allosteric modulator of the mGluR2 receptor complex (where said potential allosteric modulator may bind to the site(s) affected by the mutation in the mutant mGluR2); and,

e) quantifying the relative degree of responses among the treatments such as compared to a positive control provided a higher level of glutamate.

This allows one to rapidly screen for allosteric modulators of the glutamate/HmGluR2 receptor complex. This screening approach also is applied to mGluR3. Utilization of the screening system described above provides a sensitive and rapid means to determine compounds which modulate mGluR2 but not mGluR3 to the same extent, or vice versa. For example, a preferred modulator identified with the present invention modulates mGluR2 at a pharmacologically effective dose, but its impact on modulation of mGluR3 is small, for instance, an up-modulation of less than 20 percent, whereas the up-modulation for mGluR2 by said modulator is over 100 percent change under the same test conditions. This increase in specificity afforded by the present invention is seen to result in decrease toxicity by increasing targeted effectiveness at lower doses and eliminating undesired stimulation of non-targeted mGluRx receptors.

Another use of such mutant forms of mGluR2 and mGluR3 is to use as "negative controls" in studies that evaluate whether or not a modulator binds to the corresponding wild-type mGluR2 or mGluR3. In such studies, strong binding to mGluR2 wild-type receptors and substantially less or no binding to mGluR2 mutants at the 688, 689, and/or 735 sites indicates that said modulator has good specificity to an allosteric modulator site on wild-type mGluR2 (where the mutations disrupt the association that provides for the modulator's effect). It further suggests that there would not be cross-over effect to modulate mGluR3 at a corresponding site on mGluR3.

The following table provides a summary of desired levels of changes in the targeted mGluRx compared to the mGluRx that is most affected by the modulator but which is not the targeted subtype:

For a "positive" modulator – approx. percent change compared to no modulator			For a "negative" modulator – approx. percent change compared to no modulator		
Levels of Modulator Specificity (arbitrary scale)	Targeted mGluRx	Non-targeted mGluRx most affected by modulator	Levels of Modulator Specificity (arbitrary scale)	Targeted mGluRx	Non-targeted mGluRx most affected by modulator
1	+ 25%	+15%	1	- 25%	-15%
2	+25%	+10%	2	-25%	-10%
3	+50%	+15%	3	-50%	-15%
4	+50%	+10%	4	-50%	-10%
5	+100%	+10%	5	-100%	-10%
6	+200%	+15%	6	-200%	-15%
7	+500%	+25%	7	-500%	-25%
8	>500%	<50%	8	<500%	>50%

For example, considering a negative modulator targeted for mGluR2, where mGluR3 is the non-targeted mGluRx with the highest response of all non-targeted, identified mGluRxs, a level 4 response ("good") is when the modulator provides about a 50% decrease in glutamate response for mGluR2, and only about a 10% decrease in glutamate response for mGluR3. A better profile for a modulator, in that it is more specific, is a level 5 response, where the modulator provides about a 100% decrease in glutamate response for mGluR2, and about the same 10% decrease in glutamate response for mGluR3.

Screening systems, such as based on the above method, may also be adapted to automated procedures such as a PANDEX.RTM. (Baxter-Dade Diagnostics) system allowing for efficient high-volume screening of potential allosteric modulator therapeutic agents.

It is also noted that an oocyte transient expression system can be constructed according to the procedure described in S. Lubbert, et al, Proceedings of the National Academy of Sciences (USA), 84:4332 (1987).

In an especially preferred embodiment of this invention an assay measuring the production of phosphoinositides was performed. The production of phosphoinositides is known to positively correlated with the addition of glutamate to cells containing certain types of metabotropic receptors that are coupled to Gq or to Gs or Gi linked metabotropic GPCRs that are co-expressed with various promiscuous g-proteins that now couple these Gs or Gi linked receptors to Gq-linked second messenger systems.

In an especially preferred embodiment of this invention an assay measuring the inhibition of forskolin-stimulated cAMP synthesis was performed. The inhibition of cAMP synthesis is known to positively correlated with the addition of glutamate to cells containing certain types of metabotropic receptors.

5 In another embodiment this invention provides a method for identifying, in a test sample, DNA homologous to a probe of the present invention, wherein the test nucleic acid is contacted with the probe under hybridizing conditions and identified as being homologous to the probe. Hybridization techniques are well known in the art. See, e.g., J. Sambrook, et al., supra, at Chapter 11.

10 The nucleic acid compounds of the present invention may also be used to hybridize to genomic DNA which has been digested with one or more restriction enzymes and run on an electrophoretic gel. The hybridization of radiolabeled probes onto such restricted DNA, usually fixed to a membrane after electrophoresis, is well known in the art. See, e.g., J. Sambrook, supra. Such procedures may be employed in searching for persons with mutations in
15 these receptors by the well-known techniques of restriction fragment length polymorphisms (RFLP), the procedures of which are described in U.S. Pat. No. 4,666,828, issued May 19, 1987, the entire contents of which is incorporated herein by reference.

The determination of the DNA base sequence of the human genome is considered to have a major impact on biomedical science in the next century. It is believed that such
20 determinations will enhance a range of applications from genetic mapping of disease-associated genes to diagnostic tests for disease susceptibility and drug response. Briefly, the determination of base composition at specific, variable DNA sites known as single nucleotide polymorphisms (SNPs) is especially important. SNPs have a number of uses in mapping, disease gene identification, and diagnostic assays. All of these applications involve the determination of bases
25 composition at the SNP site.

Conventional techniques to determine base composition at a single site include minisequencing (See, e.g., "Minisequencing: A Specific Tool For DNA Analysis And Diagnostics On Oligonucleotide Arrays," by Tomi Pastinen et al., Genome Research 7, 606 (1997)), and oligo-ligation (See, e.g., "Single-Well Genotyping Of Diallelic Sequence Variations
30 By A Two-Color ELISA-Based Oligonucleotide Ligation Assay," by Vincent O. Tobe et al., Nuclear Acids Res. 24, 3728 (1996)). In minisequencing, a primer is designed to interrogate a specific site on a sample template, and polymerase is used to extend the primer with a labeled dideoxynucleotide. In oligo-ligation, a similar primer is designed, and ligase is used to covalently attach a downstream oligo that is variable at the site of interest. In each case, the
35 preference of an enzyme for correctly base-paired substrates is used to discriminate the base

identity that is revealed by the covalent attachment of a label to the primer. In most applications these assays are configured with the primer immobilized on a solid substrate, including microplates, magnetic beads and recently, oligonucleotides microarrayed on microscope slides. Detection strategies include direct labeling with fluorescence detection or indirect labeling using
5 biotin and a labeled streptavidin with fluorescent, chemiluminescent, or absorbance detection.

Oligonucleotide microarrays or "DNA chips" have also generated much attention for their potential for massively parallel analysis. The prospect of sequencing tens of thousands of bases of a small sample in just a few minutes is exciting. At present, this technology has limited availability in that arrays to sequence only a handful of genes are currently available,
10 with substantial hardware and consumable costs. In addition, the general approach of sequencing by hybridization is not particularly robust, with the requirement of significant sequence-dependent optimization of hybridization conditions. Nonetheless, the parallelism of an "array" technology is very powerful and multiplexed sequence determination is an important element of the new flow cytometry method.

15 Other methods for SNP analysis include those in U.S. Patent No. 6,287,766.

Briefly, a DNA sequence variation can be analyzed using the techniques outlined in the above referenced patent. In general, differences exist in similar DNA regions isolated from two individuals of a species. Such differences in DNA sequence are known as genetic sequence variation. Genetic sequence variation may result in phenotypic differences in the two
20 individuals or may have no phenotypic effect whatsoever. Similarly, the genetic sequence variation may have a profound effect on the host of the different genetic sequence or it may have no effect whatsoever.

Comparisons between two DNA samples can lead to useful genetic information. For example, with the various genome projects, reference or control sequences are available to
25 use for comparison purposes. New DNA samples isolated from similar or dissimilar organisms can be compared to the known sequences using the techniques of the invention. Similarly, two different unknown samples can be compared.

In general, a first DNA sample is attached to a solid support such as flow cytometry beads. The first DNA sample may be a known sample or an unknown DNA sample.
30 Next, a test sample of DNA is isolated. The test sample may be a PCR product. The test sample is then incubated with the DNA attached to the solid support under conditions suitable to permit DNA hybridization between the two DNA samples. DNA sequence differences are detected as DNA mismatches or other "mutations" in the hybrid DNA.

SNP Analysis

Single Nucleotide Polymorphisms (SNP) can be detected using the techniques disclosed in the 6,287,766 patent. Importantly, the SNPs can be known or unknown.

5 1. Known SNPs

Native or wild type DNA is attached to a solid support such as flow cytometry beads. Next, the test DNA sample is isolated. The DNA may be amplified by PCR using oligos that flank the SNP. The DNA sample is incubated with the native or wild type DNA attached to the solid support. Finally, the mixture is incubated with a labeled DNA mutation binding
10 protein. A detected mismatch is indicative of a SNP. The test sample may be isolated from a patient.

In an alternative embodiment, two solid supports are utilized. Native or wild type DNA is coupled to one support. DNA containing the known mutation is coupled to the other support. A DNA sample is incubated with both solid supports under conditions that allows DNA
15 hybridization. Finally, the mixture is incubated with a labeled DNA mutation binding protein. If the sample DNA has a SNP, the native DNA-support will show a mismatch and the mutant DNA-support will show no signal due to the match. If the sample DNA does not have the mutation, the native support will show a match and the mutant support will show a mismatch.

The most efficient solid support for known SNP analysis is flow cytometry. A
20 library of DNA molecules of interest can be coupled to the beads. Such DNA can be produced by PCR. For example, if one is attempting to identify a SNP in a particular gene such as a BRCA breast cancer gene, one would PCR a blood sample from a candidate to amplify the DNA using oligos that flank the region of interest. The PCR sample is then annealed to the beads containing the BRCA native DNA in sections of 100-200 bases with overlap to ensure SNP
25 detection in the entire gene. A DNA mutation binding protein such as thermophilic MutS protein which recognizes all DNA mismatches is labeled with a detectable label such as biotin and is then incubated with the beads. Streptavidin Phycoerythrin can be used as a reporter. If the DNA mutation binding protein (MutS) protein detects a DNA mutation, it binds to the DNA which is bound to the bead forming a DNA mutation binding protein--DNA--bead complex. The
30 biotin attached to the DNA mutation-binding protein (MutS) is detected by the reporter using flow cytometry.

The ability to assay 100 beads (each with a different form of DNA attached) per sample using flow cytometry in a matter of seconds makes the genomic approach to SNP
35 detection feasible. This approach makes it possible to narrow down the SNP to approximately 100 bases which can then be sequenced to identify the exact nature of the SNP or change in the

sequence. The DNA sequencing could be done from the initial PCR reaction used to anneal to the beads thus simplifying the SNP detection greatly.

2. Unknown SNPs

For genome wide search for SNPs, flow cytometry is the preferred form of detection. This method proposes attaching wild type DNA (in 200-500 base fragments where each fragment overlaps with the next by about 30 bases to insure that all DNA is read) to a library of beads. Preferably, smaller fragments such as 200 bases are used since it is estimated that a SNP occurs every 1000 base pairs and it is desirable to narrow down the SNP to limit the amount of sequencing needed. As above, the patient sample is amplified and annealed to the beads containing the wild type DNA sequence.

One hundred (100) beads (which can read in multiplex fashion--at the same time due to the ability to detect each bead by its fluorescent signature) can be read in one test well. Therefore in 96 well format, 9600 sections of 200 bases can be read in less than an hour which correlates with 1,920,000 bases checked for SNPs on each 96 well plate. Mismatches can be detected in minutes thereby reflecting a SNP in the respective 200 base fragment on that bead. If that fragment corresponds to a DNA sequence of interest, further sequencing (from the original PCR sample used to anneal to the beads) will identify the exact SNP sequence.

The identity of the herein disclosed nucleic acid molecules will also find use in the identification of individuals harboring identical or substantially similar sequences in their genomes. This, finding, in turn, will aid the practicing physician in identifying those individuals at risk of not responding to medicaments aimed at normalizing endogenous glutamate levels especially those seeking treatment of mGluR 2 or mGluR3 related disorders characterized with low or non-existent levels of endogenous glutamate.

Thus, a diagnostic method for screening an individual at risk for non-compliance with an mGluR2/3 related therapeutic regimen comprises:

- (a) providing a biological sample of human cell/tissue; and
- (b) determining, in the sample, levels of expression of a gene product expressed from a nucleotide sequence which hybridizes under stringent conditions with a nucleotide sequence corresponding to one or more of SEQ.ID.NOS.:9-16, or SEQ.ID.NOS.:41-48, or a complement thereof, wherein said step comprises identifying a complex formed between said gene product and an antibody having affinity for said gene product.

An alternative method proposes assaying a biological sample from a subject undergoing a therapeutic regiment to treat an mGluR2/3 related disorder characterized by

abnormal binding of glutamate to its respective receptor, for expression of mRNA of a gene comprising a sequence of nucleotides as set forth in SEQ.ID.NOS.:9-16, 25-32 or 41-48 or substantially identical thereto or encoding the polypeptide of SEQ.ID.NOS.:1-8, 17-24 or 33-40, wherein expression thereof is indicative of the presence of a mutant form of mGluR2 or 3 as disclosed herein. This finding, in turn, will aid in the identifications of individuals at risk of not responding to a therapeutic regiment aimed at correcting/normalizing levels of endogenous glutamate. In further thereto, it is believed that individuals having nucleotides sequences identical to or substantially similar to SEQ.ID.NOS.:9-16, 25-32 or 41-48 will have lower binding of a selective allosteric potentiator which would result in less potentiation by the endogenous glutamate which would eventually result in less inhibition of the presynaptic mGluR2 receptor resulting in increased synaptic activity of the presynaptic terminal.

The proteins of this invention as well as fragments of these proteins may be used as antigens for the synthesis of antibodies. The term "antibody" as used herein describes antibodies, fragments of antibodies (such as, but not limited, to Fab, Fab', Fab.sub.2', and Fv fragments), and chimeric, humanized, veneered, resurfaced, or CDR-grafted antibodies capable of binding antigens of a similar nature as the parent antibody molecule from which they are derived. The instant invention also encompasses single chain polypeptide binding molecules.

The term "antibody" as used herein is not limited by the manner in which the antibodies are produced, whether such production is in situ or not. The term "antibody" as used in this specification encompasses those antibodies produced by recombinant DNA technology means including, but not limited, to expression in bacteria, yeast, insect cell lines, or mammalian cell lines.

The production of antibodies, both monoclonal and polyclonal, in animals, especially mice, is well known in the art. See. e.g., C. Milstein, Handbook of Experimental Immunology, (Blackwell Scientific Pub., 1986); J. Goding, Monoclonal Antibodies: Principles and Practice, (Academic Press, 1983). For the production of monoclonal antibodies the basic process begins with injecting a mouse, or other suitable animal, with an immunogen. The mouse is subsequently sacrificed and cells taken from its spleen are fused with myeloma cells, resulting in a hybridoma that reproduces in vitro. The population of hybridomas is screened to isolate individual clones, each of which secretes a single antibody species, specific for the immunogen. The individual antibody species obtained in this way is each the product of a single B cell from the immune animal generated in response to a specific antigenic site, or epitope, recognized on the immunogenic substance.

Chimeric antibodies are described in U.S. Pat. No. 4,816,567, which issued Mar. 28, 1989 to S. Cabilly, et al. This reference discloses methods and vectors for the preparation of

chimeric antibodies. The entire contents of U.S. Pat. No. 4,816,567 are incorporated herein by reference. An alternative approach to production of genetically engineered antibodies is provided in U.S. Pat. No. 4,816,397, which also issued Mar. 28, 1989 to M. Boss, et al., the entire contents of which are incorporated herein by reference. The Boss patent teaches the simultaneous co-expression of the heavy and light chains of the antibody in the same host cell.

The approach of U.S. Pat. No. 4,816,397 has been further refined as taught in European Patent Publication No. 0 239 400, which published Sep. 30, 1987. The teachings of this European patent publication (Winter) are a preferred format for the genetic engineering of the reactive monoclonal antibodies of this invention. The Winter technology involves the replacement of complementarity determining regions (CDRS) of a human antibody with the CDRs of a murine monoclonal antibody thereby converting the specificity of the human antibody to the specificity of the murine antibody which was the source of the CDR regions. This "CDR grafting" technology affords a molecule containing minimal murine sequence and thus is less immunogenic.

Single chain antibody technology is yet another variety of genetically engineered antibody which is now well known in the art. See. e.g. R. E. Bird, et al., Science 242:423-426 (1988); PCT Publication No. WO 88/01649, which was published Mar. 10, 1988. The single chain antibody technology involves joining the binding regions of heavy and light chains with a polypeptide sequence to generate a single polypeptide having the binding specificity of the antibody from which it was derived.

The aforementioned genetic engineering approaches provide the skilled artisan with numerous means to generate molecules which retain the binding characteristics of the parental antibody while affording a less immunogenic format.

These antibodies are used in diagnostics, therapeutics or in diagnostic/therapeutic combinations. By "diagnostics" as used herein is meant testing that is related to either the in vitro or in vivo diagnosis of disease states or biological status in mammals, preferably in humans. By "therapeutics" and "therapeutic/diagnostic combinations" as used herein is respectively meant the treatment or the diagnosis and treatment of disease states or biological status by the in vivo administration to mammals, preferably humans, of the antibodies of the present invention. The antibodies of the present invention are used in the screening methods and in rational drug development, as well as in other applications where the immunological binding of an antibody to one of the novel polypeptides, peptides or fused proteins is desired.

Numerous other assay systems are also readily adaptable to detect agents which bind the novel polypeptides, peptides or fused proteins of the present invention. Examples of these aforementioned assay systems are discussed in Methods in Enzymology, (J. Langone. and

H. Vunakis, eds. 1981), Vol. 73, Part B, the contents of which are incorporated herein by reference. Skilled artisans are directed to Section II of Methods in Enzymology, Vol. 73, Part B, supra, which discusses labeling of antibodies and antigens, and Section IV, which discusses immunoassay methods. Also, regarding the development of other antibodies, which are specific for the hypervariable regions of the mGluR α receptor antibodies, wherein some such anti-idiotypic antibodies would resemble the original epitope, and, therefore, would be useful in evaluating the effectiveness of compounds which are potential antagonists, agonists, or partial agonists of the mGluR α receptor itself (as opposed to the allosteric site), see. e.g., Cleveland, et al., Nature (London), 305:56 (1983); Wasserman, et al., Proceedings of the National Academy of Sciences (USA), 79:4810 (1982).

Several methods for preparing and using the compounds, and applying the methods, of this invention are illustrated in the following Examples. Starting materials are made according to procedures known in the art or as illustrated herein.

EXAMPLE 1

Site Directed Mutagenesis Using the QuickChange Protocol of StratageneTM:

The following summarizes the procedure that is employed to generate mutant forms of mGluR2 and mGluR3 at the specific amino acid locations indicated above. This is based on the protocol developed by StratageneTM.

1. Resuspend all primers in sterile water with a final concentration of 1 μ g/ μ l. Make working stocks of each primer by diluting to 125ng/ μ l.

2. Prepare test reaction and positive control reaction according to table below, using components provided within the kit:

Reactants:	Test reaction	Control
10X Buffer	5 μ l	5 μ l
Template 20ng/ μ l	1 μ l hmGluR2	1 μ l pWhitescript
Primer 1 125ng/ μ l	1 μ l hmG2 Primer 1	1.25 μ l primer 1
Primer 2 125ng/ μ l	1 μ l hmG2 Primer 2	1.25 μ l primer 2
dNTP	1 μ l	1 μ l
Water	40 μ l	38.5 μ l
PfuTurbo	1 μ l	1 μ l

- Conditions:
1. Place samples in PCR thermo cycler and set the following PCR Cycling
 - 1 Cycle: 95°C - 30 seconds
 - 18 Cycles: 95°C - 30 seconds
55°C - 60 seconds
68°C - 12 minutes
 - Hold: 4°C
 2. Add 1ul DpnI enzyme to each reaction and incubate for 1 hour at 37°C.
 3. Prepare LB/AMP plates.
 4. Add 10ul .1M IPTG and 50ul 40mg/ml XGal per plate and spread evenly.
 5. Let plates sit in 37°C-incubator 1 hour before using.
 6. Thaw competent cells on ice.
 7. Transfer 50uL of competent cells to pre-chilled falcon tubes, 1 tube per reaction.
 8. Add 1uL of DpnI treated reactions to the separate tubes and incubate on ice for 30 minutes.
 9. Heat transformation in 42°C-water bath for 45 seconds, then cool on ice for 2 minutes.
 10. Add 500uL NZY+ broth (42°C). Incubate at 37°C for 1 hour at 225 rpm.
 11. Plate 250ul of transformations per plate. Incubate overnight at 37°C.
 12. Remove plates from 37°C and store at 4°C.
 13. Pick colonies from each clone and inoculated them into 4ml LB/AMP. Incubate overnight at 37°C, 300rpm.

Then DNA is isolated from overnight cultures using a mini-prep kit (Qiagen™ or equivalent) following the kit instructions. Samples of DNA are sequenced by standard sequencing protocols to determine if a desired, correct mutation has been incorporated into the sequence and no further mutations have been mis-incorporated.

EXAMPLE 2

Lipid Transfection to Introduce Mutant DNA into Cells Using MAGECTM Multi-well Plate-based Protocol:

For mGluR2 and mGluR3 mutants, the mGluR2/G α 16 nucleic acids are mixed with a lipid-based transfection reagent and are deposited into a well of a multi-well plate coated with fibronectin. The resulting DNA/lipid complexes are subsequently dried in vacuo then seeded with HEK293 cells. This is based on a method designated as MAGECTM, and disclosed in U.S. Provisional Application No. 60/372,476, filed April 15, 2002, and titled, "Matrix Analyses of Gene Expression in Cells."

The following summarizes the steps of the method:

1. In a 15 ml tube, add 0.35 μ g DNA of mGluR2 and G α 16 DNAs to 45 μ l of DNA-condensation buffer (Buffer EC from Effectene Kit, Qiagen, Inc.) in which sucrose has been dissolved to a concentration of 0.3 M.
2. Add 4.5 μ l of Enhancer solution, mix the contents by pipetting up vortex gently then incubate the mixture at room temperature for 5 minutes.
3. Add 11 μ l Effectene transfection reagent; mix the solution with gentle vortexing.
4. Incubate at room temperature for 10 minutes.
5. Add 90 μ l 0.25% glycogen, remix the solution and pipette 150 μ l into a well of multi-well fibronectin coated plate.
6. Spread solution to cover the whole bottom of the well and let sit overnight at 4°C.
7. Dry plate in a SpeedVac or vacuum chamber. Dried plates can be used immediately or stored at 4°C for an extended period.
8. One day prior to transfection, cells are passaged to maintain growth in log-phase.
9. On the day of transfection, cells are trypsinized and 600,000 cells/well are subsequently seeded into the previously coated and dried multi-well plate(s) in DMEM media containing glutamine and placed at 37°C with 6% CO₂.

10. Approximately 26-30 hours after plating, the media is removed from the cells and 1 ml of DMEM media minus glutamine is pipetted over the cells and the plates are placed back at 37°C with 6% CO₂.

11. Approximately forty hours after plating, the cells are analyzed in the phosphoinositide hydrolysis assay.

EXAMPLE 3

Measurement of Phosphoinositide Hydrolysis

Phosphoinositide hydrolysis was determined by measuring the accumulation of tritiated inositol monophosphate ([³H]-IP₁) in the presence of LiCl. The method is taken from Berridge: Biochem J (1983) 212: 849-58. This determination provides for comparative data on the level of potentiation and loss of potentiation when testing a purported or known allosteric modulator in the presence of glutamate and either a wild-type mGluR2 or mGluR3 and at least one mutant of the respective mGluR2 or mGluR3.

1. HEK293 cells that were previously transiently transfected with hmGluR2 and Gα16 were labeled overnight with 1 μCi/well myo-[2-³H]-inositol in a glutamine-free DMEM medium.

2. The following day, the medium was removed and the cells were washed two times with HBS buffer containing 125 mM NaCl, 5 mM KCl, 0.62 mM MgSO₄, 1.8 mM CaCl₂, 6 mM glucose and 20 mM HEPES pH 7.4 for 45 min at 37°C.

3. Following the wash, cells were then incubated with HBS-buffer containing 10 mM LiCl.

4. After 20 min incubation in the LiCl containing buffer, varying concentrations of glutamate alone (5 μM (EC₁₀) or 1 mM (EC₁₀₀)), or glutamate (5 μM (EC₁₀) in combination with varying concentrations of a mGluR2 potentiator were added to the wells and incubated for an additional hour at 37°C.

5. The reactions were terminated by aspiration of the media from the wells and the accumulated [³H]-inositol monophosphates were extracted by adding 1 ml of cold chloroform-methanol-HCl (4N) (200:100:2).

6. The mixtures were then transferred to glass tubes containing 300 μl of chloroform and 400 μl of H₂O and vortexing briefly.

7. The aqueous phase was separated from the organic phase, by centrifugation at 4000 rpm for 3 min or the samples can be allowed to settle for 15 min.

8. A 0.5 ml aliquot of the aqueous phase from each sample was added to an anion exchange column containing Dowex-1-X8 (200-300 mesh in the formate form).

9. After the application of the sample to the column, the columns were washed with 10 ml of a solution consisting of 60 mM ammonium formate and 5 mM borax.

10. [³H]- inositol monophosphates were eluted from the columns by the addition of 4 ml of a solution containing 800 mM ammonium formate and 0.1 M formic acid.

11. The column eluates were transferred to a 20 ml glass scintillation vial and 16 ml of scintillation cocktail (Ecolume, ICN) was added.

12. The sample was quantified in a scintillation counter after a two-hour waiting period.

EXAMPLE 4

The responses to novel nucleic acid sequences of the present invention were evaluated through comparisons with two known mGluR2 modulators. One of these modulators is designated herein as Compound A, has the chemical formula N-(3-(2-methoxyphenoxy)-phenyl-N-(2,2,2-trifluoroethylsulfonyl)pyrid-3-ylmethylamine, and is described in the PCT Patent Application No.: WO 01/56990. The second modulator is designated as Compound B.

The interactions between the novel mutant forms of mGluR2 and these modulators indicate the value of the novel nucleic acid sequences and polypeptides of the present invention for purposes including, but not limited to:

1. screening of potential mGluR2 modulators that are not active, or less active, for mGluR3;

2. screening of potential mGluR3 modulators that are not active, or less active, for mGluR2;

3. rational drug design using SAR to design specific modulators for mGluR2 that are not active, or less active, for mGluR3; and

4. rational drug design using SAR to design specific modulators for mGluR3 that are not active, or less active, for mGluR2.

Figure 1 shows the percent response of several treatments compared to a 1mM glutamate positive control with wild type mGluR2. This control is normalized and set to "100

percent on the y-axis. This figure shows the effect of the two modulators, Compound A and Compound B, in up-regulating the response of mGluR2 when provided a relatively low level, 5 uM of glutamate ("glu"), compared to the 1 mM level of the positive control (far right bar).

Figure 2 shows the effect of the SG-688-689-LV mutant of mGluR2, (SEQ.ID.NO.:5), when the same modulators are applied. A substantial decrease of glutamate response is evident in comparison to the wild-type mGluR2 results in Figure 1. This indicates that the modulators which were effective at up-regulating wild-type mGluR2 in the presence of 5 uM quantities of glutamate (relative to the 1 mM control) are both less effective at such up-modulation when acting on the SG-688-689-LV mutant of mGluR2.

Figure 3 shows the effect of the N735D mutant of mGluR2, (SEQ.ID.NO.:1), when the same modulators are applied. A substantial decrease of glutamate response is evident in comparison to the wild-type mGluR2 results in Figure 1. This indicates that the modulators which were effective at up-regulating wild-type mGluR2 in the presence of 5 uM quantities of glutamate (relative to the 1 mM control) are both less effective at such up-modulation when acting on the N735D mutant of mGluR2. It also is noted that the loss of effectiveness of these modulators is somewhat greater for this mutant than for the mutant of Figure 2.

Figure 4 shows the effect of four combined point mutations - the SG-688-689-LV and AN(733,735)TD mutant of mGluR2, (SEQ.ID.NO.:8) when the same modulators are applied. A more substantial decrease of glutamate response is evident in comparison to the wild-type mGluR2 results in Figure 1. This indicates that the modulators which were effective at up-regulating wild-type mGluR2 in the presence of 5 uM quantities of glutamate (relative to the 1 mM control) are both less effective at such up-modulation when acting on the SG-688-689-LV and AN(733,735)TD mutant of mGluR2. It also is noted that the loss of effectiveness of these modulators is greater for this mutant than for the mutants of Figures 2 and 3. This suggests that this combination of mutations is more effective at disrupting an allosteric binding site for these modulators.

Figure 5 shows the effect of two combined point mutations - the S-688-L and N735D mutant of mGluR2, (SEQ.ID.NO.:6), when the same modulators are applied. A more substantial decrease of glutamate response is evident in comparison to the wild-type mGluR2 results in Figure 1. This indicates that the modulators which were effective at up-regulating wild-type mGluR2 in the presence of 5 uM quantities of glutamate (relative to the 1 mM control) are both less effective at such up-modulation when acting on the S-688-L and N735D mutant of mGluR2. It also is noted that the loss of effectiveness of these modulators is greater for this mutant than for the mutants of Figures 2 and 3, and comparable to the results of Figure 4. This

suggests that this combination of mutations is more effective than the single point mutations, and as effective as the four-mutation combination depicted in Figure 4.

Figure 6 shows the effect of two combined point mutations - the G-689-V and N735D mutant of mGluR2, (SEQ.ID.NO.:2), when the same modulators are applied. A more substantial decrease of glutamate response is evident in comparison to the wild-type mGluR2 results in Figure 1. This indicates that the modulators which were effective at up-regulating wild-type mGluR2 in the presence of 5 uM quantities of glutamate (relative to the 1 mM control) are both less effective at such up-modulation when acting on the G-689-V and N735D mutant of mGluR2. It also is noted that the loss of effectiveness of these modulators is greater for this mutant than for the mutants of Figures 2 and 3, and comparable to the results in Figure 4. This suggests that this combination of mutations is more effective than the single point mutations, and as effective as the four-mutation combination depicted in Figure 4.

Without being bound to a specific theory, the above sets of results suggest that the amino acids at positions 688, 689, and 735 of mGluR2 are important in establishing an allosteric binding site for modulators that include the Compound A and Compound B modulators. The data indicate that substitution of amino acids in these positions dramatically affect the ability of such modulators to act on such mutated mGluR2 receptor molecules. Further, Figures 5 and 6 suggest that two mutations one at 735 and one at either 688 or 689, is essentially as effective as a mutant having four mutations. This adds to the knowledge of what structure is critical for the binding and activity of Compounds A and B, and similar modulators. This opens opportunities to exploit the discovery of these positions as critical to effectiveness of modulators, and to develop specific modulators.

Accordingly, the teachings of the present invention find application in a number of areas, some of which are described in this and the following paragraphs. For example, without being limiting of the scope of the present invention, the mutant forms of mGluR2 and mGluR3 disclosed herein are incorporated into chimeric nucleic acid sequences of human, rat, and mouse mGluR2 and/or mGluR3. The resultant chimeric mutant nucleic acids sequences, and the resultant polypeptides, are utilized in screening for modulators, testing of identified modulators for specificity, toxicity testing, and rational drug design and development. With regard to screening of substances for modulator properties, these mutant chimeric molecules, and the resultant polypeptides, are used in high-throughput functional drug screening assays in order to identify subtype-specific modulators, particularly positive modulators, of mGluR2, and of mGluR3.

Further regarding identification of the chimeric mutant nucleic acid sequences and polypeptides encoded by said sequences, novel polypeptides, peptides, and fusion proteins are

prepared wherein such polypeptides, peptides, and fusion proteins identify the polypeptides encoded by said sequences. Also, novel nucleic acid sequences are prepared that encode these novel polypeptides, peptides, and fusion proteins. Also, expression vectors expressing these novel nucleic acid sequences are prepared.

5 In certain embodiments, these novel polypeptides, peptides, and fusion proteins are designed and constructed to specifically identify a unique or rare sequence of a specific mutant chimeric polypeptide of the present invention. Alternately, in other embodiments, novel polyclonal or monoclonal antibodies are prepared and purified to immunologically bind to said chimeric polypeptide. The specificity of such antibodies is selected to meet the purpose of the
10 antibody/peptide binding requirements for a specific assay or other application. In more stringent specificity applications, the antibody is prepared to immunologically bind to the site of interaction of the modulator, e.g., at the site modified by the mutant forms of mGluR2 or mGluR3 of the present invention.

15 In other embodiments, the corresponding sequences in other species are utilized for assays, comparison and evaluations with in vitro, in vivo cellular, in vivo whole organism (e.g., chimeric), and in vivo site-specific use of mutants of the present invention. In any of such embodiments, the following sequences are of utility: mutant rat mGluR2 peptides, SEQ.ID.NOS.:17-24; mutant rat mGluR2 DNA nucleic acid sequences, SEQ.ID.NOS.:25-32; mutant human mGluR3 peptides, SEQ.ID.NOS.:33-40; and mutant human mGluR3 nucleic acid
20 sequences, SEQ.ID.NOS.:41-48.

In addition, the three sites of mutations in mGlu2 or mGluR3, and optionally nearby relevant amino acids that also are shown to play a role in the allosteric binding of a modulator in the TM4 and TM5 regions associated with these three sites of mutations, are candidates for Single Nucleotide Polymorphism (SNP) analyses. Such comparative
25 pharmacogenetics are conducted either in the general population (to obtain baseline data) and/or in sub-populations of subjects in need of modulators for prevention or treatment of certain neurological conditions, such as schizophrenia and general anxiety disorder.

The above additional application of the present invention are developed through techniques known to those skilled in the art using methods described in the present application
30 and supported by the information incorporating by reference from the patents, patent applications, publications, texts and references discussed or cited herein.

More generally, all patents, patent applications, publications, texts and references discussed or cited herein are incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually set forth in its entirety, and
35 any such incorporation is not limited in any way to the particular aspect or method for which a

patent, patent application, publication, text or reference was cited herein. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosures by virtue of prior invention. In addition, all terms not specifically defined are first taken to have the meaning given through usage in this disclosure, and if no such meaning is inferable, their normal meaning. Where a limitation is described but not given a specific term, a term corresponding to such limitation may be taken from any references, patents, applications, and other documents cited herein, or, for an application claiming priority to this application, additionally from an Invention Disclosure Statement, Examiner's Summary of Cited References, or a paper otherwise entered into the file history of this application.

Additional references useful in understanding aspects of the present disclosure and invention includes: D. D. Schoepp, "Glutamate receptors", Handbook of Receptors and Channels, Chapter 13 (S. J. Peroutka, ed., CRC Press, 1984); ESCRIBANO, A. et al., "(2S,4S)-2-Amino-4(2,2-Diphenylethyl) Pentanedioic Acid Selective Group 2 Metabotropic Glutamate Receptor Antagonist", Bioorg. Med. Chem. Lett., 1998, 765-770:8; HELTON, DAVID R., et al., "Anxiolytic and Side-Effect Profile of LY354740: A Potent, Highly Selective, Orally Active Agonist for Group II Metabotropic Glutamate Receptors", JPET, 1998, 651-760: 284, USA; KINGSTON, A.E., et al., "LY341495 is a Nanomolar Potent and Selective Antagonist of Group II Metabotropic Glutamate Receptors", Neuropharmacology, 1998, 1-12: 37; MUKHOPADHYAYA, J.K., et al., "Synthesis of N¹ - Substituted Analogues of (2R, 4R) - 4-Amino-pyrrolidine-2,4-dicarboxylic Acid as Agonists, Partial Agonists, and Antagonists of Group II Metabotropic Glutamate Receptors", Bioorg. Med. Chem. Lett., 2001, 1919-1924: 11; NAKAZATO, A. et al., "Synthesis, SARs, and Pharmacological Characterization of 2-Amino-3 or 6-fluorobicyclo[3.1.0] hexane-2,6-dicarboxylic Acid Derivatives as Potent, Selective, and Orally Active Group II Metabotropic Glutamate Receptor Agonists", J. Med. Chem., 2000, 4893-4909: 43, 25; NAPLES, M.A. & HAMPSON D.R., "Pharmacological Profiles of the Metabotropic Glutamate Receptor Ligands [³ H]L-AP4 and [³ H] CPPG", Neuropharmacology, 2001, 170-177: 40; ORNSTEIN, P.L. et al., "2-Substituted (2SR)-2-Amino-2-((1SR,2 SR)—carboxycycloprop-1-yl) glycines as Potent and Selective Antagonists of Group II Metabotropic Glutamate Receptors. 1. Effects of Alkyl, Arylalkyl, and Diarylalkyl Substitution", J. Med. Chem., 1998, 346-357: 41.

The present invention is not to be limited in scope by the specific embodiments described herein. Indeed, various modifications of the invention in addition to those described herein will become apparent to those skilled in the art from the foregoing description. Such modifications are intended to fall within the scope of the appended claims. Thus, for the above variations and in other regards, it should be understood that the examples and embodiments

described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and the scope of the appended claims.

Also, although only a few exemplary embodiments of this invention have been
5 described in detail above, those skilled in the art will readily appreciate that many modifications are possible without undue experimentation based on the methods and citations provided, and based on the exemplary embodiments herein, without materially departing from the novel teachings and advantages of this invention. Accordingly, all such modifications are intended to be included in the scope of this invention as defined in the following claims. In the claims,
10 means-plus-function clauses and step-plus-function clauses are intended to cover the structures described herein as effectuating or performing the recited function and to cover not only structural equivalents, but also to cover equivalent structures as one of ordinary skill in the art would understand equivalence with regard to a any means or any step that will achieve a stated function in an equivalent manner. For instance, a “means to detect a modulator of mGluR2”
15 should be taken to include methods now or later known to those of skill in the art regardless of differences in the exact steps and reagents required to achieve this function.

Seq ID 1: N735D

	5		10		15		20												
Met	Gly	Ser	Leu	Leu	Ala	Leu	Leu	Ala	Leu	Pro	Leu	Trp	Gly	Ala	Val	Ala	Glu	Gly	
	25		30		35		40												
Pro	Ala	Lys	Lys	Val	Leu	Thr	Leu	Glu	Gly	Asp	Leu	Val	Leu	Gly	Gly	Leu	Phe	Pro	Val
	45		50		55		60												
His	Gln	Lys	Gly	Gly	Pro	Ala	Glu	Asp	Cys	Gly	Pro	Val	Asn	Glu	His	Arg	Gly	Ile	Gln
	65		70		75		80												
Arg	Leu	Glu	Ala	Met	Leu	Phe	Ala	Leu	Asp	Arg	Ile	Asn	Arg	Asp	Pro	His	Leu	Leu	Pro
	85		90		95		100												
Gly	Val	Arg	Leu	Gly	Ala	His	Ile	Leu	Asp	Ser	Cys	Ser	Lys	Asp	Thr	His	Ala	Leu	Glu
	105		110		115		120												
Gln	Ala	Leu	Asp	Phe	Val	Arg	Ala	Ser	Leu	Ser	Arg	Gly	Ala	Asp	Gly	Ser	Arg	His	Ile
	125		130		135		140												
Cys	Pro	Asp	Gly	Ser	Tyr	Ala	Thr	His	Gly	Asp	Ala	Pro	Thr	Ala	Ile	Thr	Gly	Val	Ile
	145		150		155		160												
Gly	Gly	Ser	Tyr	Ser	Asp	Val	Ser	Ile	Gln	Val	Ala	Asn	Leu	Leu	Arg	Leu	Phe	Gln	Ile
	165		170		175		180												
Pro	Gln	Ile	Ser	Tyr	Ala	Ser	Thr	Ser	Ala	Lys	Leu	Ser	Asp	Lys	Ser	Arg	Tyr	Asp	Tyr
	185		190		195		200												
Phe	Ala	Arg	Thr	Val	Pro	Pro	Asp	Phe	Phe	Gln	Ala	Lys	Ala	Met	Ala	Glu	Ile	Leu	Arg
	205		210		215		220												
Phe	Phe	Asn	Trp	Thr	Tyr	Val	Ser	Thr	Val	Ala	Ser	Glu	Gly	Asp	Tyr	Gly	Glu	Thr	Gly
	225		230		235		240												
Ile	Glu	Ala	Phe	Glu	Leu	Glu	Ala	Arg	Ala	Arg	Asn	Ile	Cys	Val	Ala	Thr	Ser	Glu	Lys
	245		250		255		260												
Val	Gly	Arg	Ala	Met	Ser	Arg	Ala	Ala	Phe	Glu	Gly	Val	Val	Arg	Ala	Leu	Leu	Gln	Lys
	265		270		275		280												
Pro	Ser	Ala	Arg	Val	Ala	Val	Leu	Phe	Thr	Arg	Ser	Glu	Asp	Ala	Arg	Glu	Leu	Leu	Ala
	285		290		295		300												
Ala	Ser	Gln	Arg	Leu	Asn	Ala	Ser	Phe	Thr	Trp	Val	Ala	Ser	Asp	Gly	Trp	Gly	Ala	Leu
	305		310		315		320												
Glu	Ser	Val	Val	Ala	Gly	Ser	Glu	Gly	Ala	Ala	Glu	Gly	Ala	Ile	Thr	Ile	Glu	Leu	Ala
	325		330		335		340												
Ser	Tyr	Pro	Ile	Ser	Asp	Phe	Ala	Ser	Tyr	Phe	Gln	Ser	Leu	Asp	Pro	Trp	Asn	Asn	Ser
	345		350		355		360												
Arg	Asn	Pro	Trp	Phe	Arg	Glu	Phe	Trp	Glu	Gln	Arg	Phe	Arg	Cys	Ser	Phe	Arg	Gln	Arg
	365		370		375		380												
Asp	Cys	Ala	Ala	His	Ser	Leu	Arg	Ala	Val	Pro	Phe	Glu	Gln	Glu	Ser	Lys	Ile	Met	Phe
	385		390		395		400												
Val	Val	Asn	Ala	Val	Tyr	Ala	Met	Ala	His	Ala	Leu	His	Asn	Met	His	Arg	Ala	Leu	Cys

405	410	415	420
Pro Asn Thr Thr Arg Leu Cys Asp Ala Met Arg Pro Val Asn Gly Arg Arg Leu Tyr Lys			
425	430	435	440
Asp Phe Val Leu Asn Val Lys Phe Asp Ala Pro Phe Arg Pro Ala Asp Thr His Asn Glu			
445	450	455	460
Val Arg Phe Asp Arg Phe Gly Asp Gly Ile Gly Arg Tyr Asn Ile Phe Thr Tyr Leu Arg			
465	470	475	480
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln Lys Val Gly Tyr Trp Ala Glu Gly Leu Thr Leu			
485	490	495	500
Asp Thr Ser Leu Ile Pro Trp Ala Ser Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys			
505	510	515	520
Ser Glu Pro Cys Leu Gln Asn Glu Val Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp			
525	530	535	540
Leu Cys Ile Pro Cys Gln Pro Tyr Glu Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys			
545	550	555	560
Gly Leu Gly Tyr Trp Pro Asn Ala Ser Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr			
565	570	575	580
Ile Arg Trp Gly Asp Ala Trp Ala Val Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu			
585	590	595	600
Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala			
605	610	615	620
Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr			
625	630	635	640
Phe Ile Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly			
645	650	655	660
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile			
665	670	675	680
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val			
685	690	695	700
Ala Ile Cys Leu Ala Leu Ile Ser Gly Gln Leu Leu Ile Val Val Ala Trp Leu Val Val			
705	710	715	720
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg			
725	730	735	740
Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asp Val Leu Leu Ile Ala			
745	750	755	760
Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys			
765	770	775	780
Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe			
785	790	795	800
Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu			
805	810	815	820

Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln
 825 830 835 840
 Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Ala
 845 850 855 860
 Arg Ala Ser Ser Ser Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn
 865 870
 Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter

Seq ID 2: G689V N735D

5 10 15 20
 Met Gly Ser Leu Leu Ala Leu Leu Ala Leu Leu Pro Leu Trp Gly Ala Val Ala Glu Gly
 25 30 35 40
 Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val
 45 50 55 60
 His Gln Lys Gly Gly Pro Ala Glu Asp Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln
 65 70 75 80
 Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro
 85 90 95 100
 Gly Val Arg Leu Gly Ala His Ile Leu Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu
 105 110 115 120
 Gln Ala Leu Asp Phe Val Arg Ala Ser Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile
 125 130 135 140
 Cys Pro Asp Gly Ser Tyr Ala Thr His Gly Asp Ala Pro Thr Ala Ile Thr Gly Val Ile
 145 150 155 160
 Gly Gly Ser Tyr Ser Asp Val Ser Ile Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile
 165 170 175 180
 Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp Lys Ser Arg Tyr Asp Tyr
 185 190 195 200
 Phe Ala Arg Thr Val Pro Pro Asp Phe Phe Gln Ala Lys Ala Met Ala Glu Ile Leu Arg
 205 210 215 220
 Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly Asp Tyr Gly Glu Thr Gly
 225 230 235 240
 Ile Glu Ala Phe Glu Leu Glu Ala Arg Ala Arg Asn Ile Cys Val Ala Thr Ser Glu Lys
 245 250 255 260
 Val Gly Arg Ala Met Ser Arg Ala Ala Phe Glu Gly Val Val Arg Ala Leu Leu Gln Lys
 265 270 275 280
 Pro Ser Ala Arg Val Ala Val Leu Phe Thr Arg Ser Glu Asp Ala Arg Glu Leu Leu Ala
 285 290 295 300
 Ala Ser Gln Arg Leu Asn Ala Ser Phe Thr Trp Val Ala Ser Asp Gly Trp Gly Ala Leu
 305 310 315 320
 Glu Ser Val Val Ala Gly Ser Glu Gly Ala Ala Glu Gly Ala Ile Thr Ile Glu Leu Ala

325	330	335	340
Ser Tyr Pro Ile Ser Asp Phe Ala Ser Tyr Phe Gln Ser Leu Asp Pro Trp Asn Asn Ser			
345	350	355	360
Arg Asn Pro Trp Phe Arg Glu Phe Trp Glu Gln Arg Phe Arg Cys Ser Phe Arg Gln Arg			
365	370	375	380
Asp Cys Ala Ala His Ser Leu Arg Ala Val Pro Phe Glu Gln Glu Ser Lys Ile Met Phe			
385	390	395	400
Val Val Asn Ala Val Tyr Ala Met Ala His Ala Leu His Asn Met His Arg Ala Leu Cys			
405	410	415	420
Pro Asn Thr Thr Arg Leu Cys Asp Ala Met Arg Pro Val Asn Gly Arg Arg Leu Tyr Lys			
425	430	435	440
Asp Phe Val Leu Asn Val Lys Phe Asp Ala Pro Phe Arg Pro Ala Asp Thr His Asn Glu			
445	450	455	460
Val Arg Phe Asp Arg Phe Gly Asp Gly Ile Gly Arg Tyr Asn Ile Phe Thr Tyr Leu Arg			
465	470	475	480
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln Lys Val Gly Tyr Trp Ala Glu Gly Leu Thr Leu			
485	490	495	500
Asp Thr Ser Leu Ile Pro Trp Ala Ser Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys			
505	510	515	520
Ser Glu Pro Cys Leu Gln Asn Glu Val Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp			
525	530	535	540
Leu Cys Ile Pro Cys Gln Pro Tyr Glu Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys			
545	550	555	560
Gly Leu Gly Tyr Trp Pro Asn Ala Ser Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr			
565	570	575	580
Ile Arg Trp Gly Asp Ala Trp Ala Val Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu			
585	590	595	600
Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala			
605	610	615	620
Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr			
625	630	635	640
Phe Ile Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly			
645	650	655	660
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile			
665	670	675	680
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val			
685	690	695	700
Ala Ile Cys Leu Ala Leu Ile Ser Val Gln Leu Leu Ile Val Val Ala Trp Leu Val Val			
705	710	715	720
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg			

725	730	735	740
Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asp Val Leu Leu Ile Ala			
745	750	755	760
Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys			
765	770	775	780
Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe			
785	790	795	800
Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu			
805	810	815	820
Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln			
825	830	835	840
Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Ala			
845	850	855	860
Arg Ala Ser Ser Ser Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn			
865	870		
Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter			

Seq ID 3: S688L, G689V, N735D

5	10	15	20
Met Gly Ser Leu Leu Ala Leu Leu Ala Leu Leu Pro Leu Trp Gly Ala Val Ala Glu Gly			
25	30	35	40
Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val			
45	50	55	60
His Gln Lys Gly Gly Pro Ala Glu Asp Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln			
65	70	75	80
Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro			
85	90	95	100
Gly Val Arg Leu Gly Ala His Ile Leu Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu			
105	110	115	120
Gln Ala Leu Asp Phe Val Arg Ala Ser Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile			
125	130	135	140
Cys Pro Asp Gly Ser Tyr Ala Thr His Gly Asp Ala Pro Thr Ala Ile Thr Gly Val Ile			
145	150	155	160
Gly Gly Ser Tyr Ser Asp Val Ser Ile Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile			
165	170	175	180
Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp Lys Ser Arg Tyr Asp Tyr			
185	190	195	200
Phe Ala Arg Thr Val Pro Pro Asp Phe Phe Gln Ala Lys Ala Met Ala Glu Ile Leu Arg			
205	210	215	220
Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly Asp Tyr Gly Glu Thr Gly			
225	230	235	240
Ile Glu Ala Phe Glu Leu Glu Ala Arg Ala Arg Asn Ile Cys Val Ala Thr Ser Glu Lys			

245	250	255	260
Val Gly Arg Ala Met Ser Arg Ala Ala	Phe Glu Gly Val Val	Arg Ala Leu Leu Gln Lys	
265	270	275	280
Pro Ser Ala Arg Val Ala Val Leu Phe	Thr Arg Ser Glu Asp Ala Arg	Glu Leu Leu Ala	
285	290	295	300
Ala Ser Gln Arg Leu Asn Ala Ser Phe	Thr Trp Val Ala Ser Asp Gly Trp	Gly Ala Leu	
305	310	315	320
Glu Ser Val Val Ala Gly Ser Glu Gly	Ala Ala Glu Gly Ala Ile Thr	Ile Glu Leu Ala	
325	330	335	340
Ser Tyr Pro Ile Ser Asp Phe Ala Ser	Tyr Phe Gln Ser Leu Asp Pro Trp	Asn Asn Ser	
345	350	355	360
Arg Asn Pro Trp Phe Arg Glu Phe Trp	Glu Gln Arg Phe Arg Cys Ser Phe	Arg Gln Arg	
365	370	375	380
Asp Cys Ala Ala His Ser Leu Arg Ala	Val Pro Phe Glu Gln Glu Ser Lys	Ile Met Phe	
385	390	395	400
Val Val Asn Ala Val Tyr Ala Met Ala	His Ala Leu His Asn Met His Arg	Ala Leu Cys	
405	410	415	420
Pro Asn Thr Thr Arg Leu Cys Asp Ala	Met Arg Pro Val Asn Gly Arg Arg	Leu Tyr Lys	
425	430	435	440
Asp Phe Val Leu Asn Val Lys Phe Asp	Ala Pro Phe Arg Pro Ala Asp Thr	His Asn Glu	
445	450	455	460
Val Arg Phe Asp Arg Phe Gly Asp Gly	Ile Gly Arg Tyr Asn Ile Phe Thr	Tyr Leu Arg	
465	470	475	480
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln	Lys Val Gly Tyr Trp Ala Glu Gly	Leu Thr Leu	
485	490	495	500
Asp Thr Ser Leu Ile Pro Trp Ala Ser	Pro Ser Ala Gly Pro Leu Pro Ala	Ser Arg Cys	
505	510	515	520
Ser Glu Pro Cys Leu Gln Asn Glu Val	Lys Ser Val Gln Pro Gly Glu Val	Cys Cys Trp	
525	530	535	540
Leu Cys Ile Pro Cys Gln Pro Tyr Glu	Tyr Arg Leu Asp Glu Phe Thr Cys	Ala Asp Cys	
545	550	555	560
Gly Leu Gly Tyr Trp Pro Asn Ala Ser	Leu Thr Gly Cys Phe Glu Leu Pro	Gln Glu Tyr	
565	570	575	580
Ile Arg Trp Gly Asp Ala Trp Ala Val	Gly Pro Val Thr Ile Ala Cys Leu	Gly Ala Leu	
585	590	595	600
Ala Thr Leu Phe Val Leu Gly Val Phe	Val Arg His Asn Ala Thr Pro Val	Val Lys Ala	
605	610	615	620
Ser Gly Arg Glu Leu Cys Tyr Ile Leu	Leu Gly Gly Val Phe Leu Cys Tyr	Cys Met Thr	
625	630	635	640
Phe Ile Phe Ile Ala Lys Pro Ser Thr	Ala Val Cys Thr Leu Arg Arg Leu	Gly Leu Gly	

645	650	655	660
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile			
665	670	675	680
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val			
685	690	695	700
Ala Ile Cys Leu Ala Leu Ile Leu Val Gln Leu Leu Ile Val Val Ala Trp Leu Val Val			
705	710	715	720
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg			
725	730	735	740
Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asp Val Leu Leu Ile Ala			
745	750	755	760
Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys			
765	770	775	780
Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe			
785	790	795	800
Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu			
805	810	815	820
Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln			
825	830	835	840
Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Ala			
845	850	855	860
Arg Ala Ser Ser Ser Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn			
865	870		
Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter			

Seq ID 4: S688L

5	10	15	20
Met Gly Ser Leu Leu Ala Leu Leu Ala Leu Leu Pro Leu Trp Gly Ala Val Ala Glu Gly			
25	30	35	40
Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val			
45	50	55	60
His Gln Lys Gly Gly Pro Ala Glu Asp Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln			
65	70	75	80
Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro			
85	90	95	100
Gly Val Arg Leu Gly Ala His Ile Leu Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu			
105	110	115	120
Gln Ala Leu Asp Phe Val Arg Ala Ser Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile			
125	130	135	140
Cys Pro Asp Gly Ser Tyr Ala Thr His Gly Asp Ala Pro Thr Ala Ile Thr Gly Val Ile			
145	150	155	160
Gly Gly Ser Tyr Ser Asp Val Ser Ile Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile			

Pro Gln Ile Ser Tyr	165	Ala Ser Thr Ser	170	Lys Leu Ser Asp	175	Lys Ser Arg Tyr Asp	180
Phe Ala Arg Thr	185	Val Pro Pro Asp	190	Phe Phe Gln Ala	195	Lys Ala Met Ala	200
Phe Phe Asn Trp	205	Thr Tyr Val Ser	210	Thr Val Ala Ser	215	Glu Gly Asp Tyr	220
Ile Glu Ala Phe	225	Glu Leu Glu Ala	230	Arg Ala Arg Asn	235	Ile Cys Val Ala	240
Val Gly Arg Ala	245	Met Ser Arg Ala	250	Ala Phe Glu Gly	255	Val Val Arg Ala	260
Pro Ser Ala Arg	265	Val Ala Val Leu	270	Phe Thr Arg Ser	275	Glu Asp Ala Arg	280
Ala Ser Gln Arg	285	Leu Asn Ala Ser	290	Phe Thr Trp Val	295	Ala Ser Asp Gly	300
Glu Ser Val Val	305	Ala Gly Ser Glu	310	Gly Ala Ala Glu	315	Gly Ala Ile Thr	320
Ser Tyr Pro Ile	325	Ser Asp Phe Ala	330	Ser Tyr Phe Gln	335	Ser Leu Asp Pro	340
Arg Asn Pro Trp	345	Phe Arg Glu Phe	350	Trp Glu Gln Arg	355	Phe Arg Cys Ser	360
Asp Cys Ala Ala	365	His Ser Leu Arg	370	Ala Val Pro Phe	375	Glu Gln Glu Ser	380
Val Val Asn Ala	385	Val Tyr Ala Met	390	Ala His Ala Leu	395	His Asn Met His	400
Pro Asn Thr Thr	405	Arg Leu Cys Asp	410	Ala Met Arg Pro	415	Val Asn Gly Arg	420
Asp Phe Val Leu	425	Asn Val Lys Phe	430	Asp Ala Pro Phe	435	Arg Pro Ala Asp	440
Val Arg Phe Asp	445	Arg Phe Gly Asp	450	Gly Ile Gly Arg	455	Tyr Asn Ile Phe	460
Ala Gly Ser Gly	465	Arg Tyr Arg Tyr	470	Gln Lys Val Gly	475	Tyr Trp Ala Glu	480
Asp Thr Ser Leu	485	Ile Pro Trp Ala	490	Ser Pro Ser Ala	495	Gly Pro Leu Pro	500
Ser Glu Pro Cys	505	Leu Gln Asn Glu	510	Val Lys Ser Val	515	Gln Pro Gly Glu	520
Leu Cys Ile Pro	525	Cys Gln Pro Tyr	530	Glu Tyr Arg Leu	535	Asp Glu Phe Thr	540
Gly Leu Gly Tyr	545	Trp Pro Asn Ala	550	Ser Leu Thr Gly	555	Cys Phe Glu Leu	560

565	570	575	580
Ile Arg Trp Gly Asp Ala Trp Ala Val Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu			
585	590	595	600
Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala			
605	610	615	620
Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr			
625	630	635	640
Phe Ile Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly			
645	650	655	660
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile			
665	670	675	680
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val			
685	690	695	700
Ala Ile Cys Leu Ala Leu Ile Leu Gly Gln Leu Leu Ile Val Val Ala Trp Leu Val Val			
705	710	715	720
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg			
725	730	735	740
Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asn Val Leu Leu Ile Ala			
745	750	755	760
Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys			
765	770	775	780
Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe			
785	790	795	800
Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu			
805	810	815	820
Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln			
825	830	835	840
Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Ala			
845	850	855	860
Arg Ala Ser Ser Ser Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn			
865	870		
Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter			

Seq ID 5: S688L G689V

5	10	15	20
Met Gly Ser Leu Leu Ala Leu Leu Ala Leu Leu Pro Leu Trp Gly Ala Val Ala Glu Gly			
25	30	35	40
Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val			
45	50	55	60
His Gln Lys Gly Gly Pro Ala Glu Asp Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln			
65	70	75	80
Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro			

	85		90		95		100
Gly Val Arg Leu	Gly Ala His Ile Leu	Asp Ser Cys Ser Lys	Asp Thr His Ala Leu	Glu			
	105		110		115		120
Gln Ala Leu Asp	Phe Val Arg Ala Ser	Leu Ser Arg Gly Ala	Asp Gly Ser Arg His	Ile			
	125		130		135		140
Cys Pro Asp Gly	Ser Tyr Ala Thr His	Gly Asp Ala Pro Thr	Ala Ile Thr Gly Val	Ile			
	145		150		155		160
Gly Gly Ser Tyr	Ser Asp Val Ser Ile	Gln Val Ala Asn Leu	Leu Arg Leu Phe Gln	Ile			
	165		170		175		180
Pro Gln Ile Ser	Tyr Ala Ser Thr Ser	Ala Lys Leu Ser Asp	Lys Ser Arg Tyr Asp	Tyr			
	185		190		195		200
Phe Ala Arg Thr	Val Pro Pro Asp Phe	Phe Gln Ala Lys Ala	Met Ala Glu Ile Leu	Arg			
	205		210		215		220
Phe Phe Asn Trp	Thr Tyr Val Ser Thr	Val Ala Ser Glu Gly	Asp Tyr Gly Glu Thr	Gly			
	225		230		235		240
Ile Glu Ala Phe	Glu Leu Glu Ala Arg	Ala Arg Asn Ile Cys	Val Ala Thr Ser Glu	Lys			
	245		250		255		260
Val Gly Arg Ala	Met Ser Arg Ala Ala	Phe Glu Gly Val Val	Arg Ala Leu Leu Gln	Lys			
	265		270		275		280
Pro Ser Ala Arg	Val Ala Val Leu Phe	Thr Arg Ser Glu Asp	Ala Arg Glu Leu Leu	Ala			
	285		290		295		300
Ala Ser Gln Arg	Leu Asn Ala Ser Phe	Thr Trp Val Ala Ser	Asp Gly Trp Gly Ala	Leu			
	305		310		315		320
Glu Ser Val Val	Ala Gly Ser Glu Gly	Ala Ala Glu Gly Ala	Ile Thr Ile Glu Leu	Ala			
	325		330		335		340
Ser Tyr Pro Ile	Ser Asp Phe Ala Ser	Tyr Phe Gln Ser Leu	Asp Pro Trp Asn Asn	Ser			
	345		350		355		360
Arg Asn Pro Trp	Phe Arg Glu Phe Trp	Glu Gln Arg Phe Arg	Cys Ser Phe Arg Gln	Arg			
	365		370		375		380
Asp Cys Ala Ala	His Ser Leu Arg Ala	Val Pro Phe Glu Gln	Glu Ser Lys Ile Met	Phe			
	385		390		395		400
Val Val Asn Ala	Val Tyr Ala Met Ala	His Ala Leu His Asn	Met His Arg Ala Leu	Cys			
	405		410		415		420
Pro Asn Thr Thr	Arg Leu Cys Asp Ala	Met Arg Pro Val Asn	Gly Arg Arg Leu Tyr	Lys			
	425		430		435		440
Asp Phe Val Leu	Asn Val Lys Phe Asp	Ala Pro Phe Arg Pro	Ala Asp Thr His Asn	Glu			
	445		450		455		460
Val Arg Phe Asp	Arg Phe Gly Asp Gly	Ile Gly Arg Tyr Asn	Ile Phe Thr Tyr Leu	Arg			
	465		470		475		480
Ala Gly Ser Gly	Arg Tyr Arg Tyr Gln	Lys Val Gly Tyr Trp	Ala Glu Gly Leu Thr	Leu			

485	490	495	500
Asp Thr Ser Leu Ile Pro Trp Ala Ser	Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys		
505	510	515	520
Ser Glu Pro Cys Leu Gln Asn Glu Val Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp			
525	530	535	540
Leu Cys Ile Pro Cys Gln Pro Tyr Glu Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys			
545	550	555	560
Gly Leu Gly Tyr Trp Pro Asn Ala Ser Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr			
565	570	575	580
Ile Arg Trp Gly Asp Ala Trp Ala Val Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu			
585	590	595	600
Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala			
605	610	615	620
Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr			
625	630	635	640
Phe Ile Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly			
645	650	655	660
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile			
665	670	675	680
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val			
685	690	695	700
Ala Ile Cys Leu Ala Leu Ile Leu Val Gln Leu Leu Ile Val Val Ala Trp Leu Val Val			
705	710	715	720
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg			
725	730	735	740
Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asn Val Leu Leu Ile Ala			
745	750	755	760
Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys			
765	770	775	780
Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe			
785	790	795	800
Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu			
805	810	815	820
Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln			
825	830	835	840
Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Ala			
845	850	855	860
Arg Ala Ser Ser Ser Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn			
865	870		
Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter			

Seq ID 6: S688L N735D

	5		10		15		20												
Met	Gly	Ser	Leu	Leu	Ala	Leu	Leu	Ala	Leu	Pro	Leu	Trp	Gly	Ala	Val	Ala	Glu	Gly	
	25		30		35		40												
Pro	Ala	Lys	Lys	Val	Leu	Thr	Leu	Glu	Gly	Asp	Leu	Val	Leu	Gly	Gly	Leu	Phe	Pro	Val
	45		50		55		60												
His	Gln	Lys	Gly	Gly	Pro	Ala	Glu	Asp	Cys	Gly	Pro	Val	Asn	Glu	His	Arg	Gly	Ile	Gln
	65		70		75		80												
Arg	Leu	Glu	Ala	Met	Leu	Phe	Ala	Leu	Asp	Arg	Ile	Asn	Arg	Asp	Pro	His	Leu	Leu	Pro
	85		90		95		100												
Gly	Val	Arg	Leu	Gly	Ala	His	Ile	Leu	Asp	Ser	Cys	Ser	Lys	Asp	Thr	His	Ala	Leu	Glu
	105		110		115		120												
Gln	Ala	Leu	Asp	Phe	Val	Arg	Ala	Ser	Leu	Ser	Arg	Gly	Ala	Asp	Gly	Ser	Arg	His	Ile
	125		130		135		140												
Cys	Pro	Asp	Gly	Ser	Tyr	Ala	Thr	His	Gly	Asp	Ala	Pro	Thr	Ala	Ile	Thr	Gly	Val	Ile
	145		150		155		160												
Gly	Gly	Ser	Tyr	Ser	Asp	Val	Ser	Ile	Gln	Val	Ala	Asn	Leu	Leu	Arg	Leu	Phe	Gln	Ile
	165		170		175		180												
Pro	Gln	Ile	Ser	Tyr	Ala	Ser	Thr	Ser	Ala	Lys	Leu	Ser	Asp	Lys	Ser	Arg	Tyr	Asp	Tyr
	185		190		195		200												
Phe	Ala	Arg	Thr	Val	Pro	Pro	Asp	Phe	Phe	Gln	Ala	Lys	Ala	Met	Ala	Glu	Ile	Leu	Arg
	205		210		215		220												
Phe	Phe	Asn	Trp	Thr	Tyr	Val	Ser	Thr	Val	Ala	Ser	Glu	Gly	Asp	Tyr	Gly	Glu	Thr	Gly
	225		230		235		240												
Ile	Glu	Ala	Phe	Glu	Leu	Glu	Ala	Arg	Ala	Arg	Asn	Ile	Cys	Val	Ala	Thr	Ser	Glu	Lys
	245		250		255		260												
Val	Gly	Arg	Ala	Met	Ser	Arg	Ala	Ala	Phe	Glu	Gly	Val	Val	Arg	Ala	Leu	Leu	Gln	Lys
	265		270		275		280												
Pro	Ser	Ala	Arg	Val	Ala	Val	Leu	Phe	Thr	Arg	Ser	Glu	Asp	Ala	Arg	Glu	Leu	Leu	Ala
	285		290		295		300												
Ala	Ser	Gln	Arg	Leu	Asn	Ala	Ser	Phe	Thr	Trp	Val	Ala	Ser	Asp	Gly	Trp	Gly	Ala	Leu
	305		310		315		320												
Glu	Ser	Val	Val	Ala	Gly	Ser	Glu	Gly	Ala	Ala	Glu	Gly	Ala	Ile	Thr	Ile	Glu	Leu	Ala
	325		330		335		340												
Ser	Tyr	Pro	Ile	Ser	Asp	Phe	Ala	Ser	Tyr	Phe	Gln	Ser	Leu	Asp	Pro	Trp	Asn	Asn	Ser
	345		350		355		360												
Arg	Asn	Pro	Trp	Phe	Arg	Glu	Phe	Trp	Glu	Gln	Arg	Phe	Arg	Cys	Ser	Phe	Arg	Gln	Arg
	365		370		375		380												
Asp	Cys	Ala	Ala	His	Ser	Leu	Arg	Ala	Val	Pro	Phe	Glu	Gln	Glu	Ser	Lys	Ile	Met	Phe
	385		390		395		400												
Val	Val	Asn	Ala	Val	Tyr	Ala	Met	Ala	His	Ala	Leu	His	Asn	Met	His	Arg	Ala	Leu	Cys

Pro Asn Thr Thr Arg Leu Cys Asp Ala Met Arg Pro Val Asn Gly Arg Arg Leu Tyr Lys	405	410	415	420
Asp Phe Val Leu Asn Val Lys Phe Asp Ala Pro Phe Arg Pro Ala Asp Thr His Asn Glu	425	430	435	440
Val Arg Phe Asp Arg Phe Gly Asp Gly Ile Gly Arg Tyr Asn Ile Phe Thr Tyr Leu Arg	445	450	455	460
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln Lys Val Gly Tyr Trp Ala Glu Gly Leu Thr Leu	465	470	475	480
Asp Thr Ser Leu Ile Pro Trp Ala Ser Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys	485	490	495	500
Ser Glu Pro Cys Leu Gln Asn Glu Val Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp	505	510	515	520
Leu Cys Ile Pro Cys Gln Pro Tyr Glu Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys	525	530	535	540
Gly Leu Gly Tyr Trp Pro Asn Ala Ser Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr	545	550	555	560
Ile Arg Trp Gly Asp Ala Trp Ala Val Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu	565	570	575	580
Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala	585	590	595	600
Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr	605	610	615	620
Phe Ile Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly	625	630	635	640
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile	645	650	655	660
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val	665	670	675	680
Ala Ile Cys Leu Ala Leu Ile Leu Gly Gln Leu Leu Ile Val Val Ala Trp Leu Val Val	685	690	695	700
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg	705	710	715	720
Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asp Val Leu Leu Ile Ala	725	730	735	740
Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys	745	750	755	760
Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe	765	770	775	780
Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu	785	790	795	800
	805	810	815	820

Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln
 825 830 835 840
 Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Ala
 845 850 855 860
 Arg Ala Ser Ser Ser Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn
 865 870
 Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter

Seq ID 7: G689V

5 10 15 20
 Met Gly Ser Leu Leu Ala Leu Leu Ala Leu Leu Pro Leu Trp Gly Ala Val Ala Glu Gly
 25 30 35 40
 Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val
 45 50 55 60
 His Gln Lys Gly Gly Pro Ala Glu Asp Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln
 65 70 75 80
 Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro
 85 90 95 100
 Gly Val Arg Leu Gly Ala His Ile Leu Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu
 105 110 115 120
 Gln Ala Leu Asp Phe Val Arg Ala Ser Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile
 125 130 135 140
 Cys Pro Asp Gly Ser Tyr Ala Thr His Gly Asp Ala Pro Thr Ala Ile Thr Gly Val Ile
 145 150 155 160
 Gly Gly Ser Tyr Ser Asp Val Ser Ile Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile
 165 170 175 180
 Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp Lys Ser Arg Tyr Asp Tyr
 185 190 195 200
 Phe Ala Arg Thr Val Pro Pro Asp Phe Phe Gln Ala Lys Ala Met Ala Glu Ile Leu Arg
 205 210 215 220
 Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly Asp Tyr Gly Glu Thr Gly
 225 230 235 240
 Ile Glu Ala Phe Glu Leu Glu Ala Arg Ala Arg Asn Ile Cys Val Ala Thr Ser Glu Lys
 245 250 255 260
 Val Gly Arg Ala Met Ser Arg Ala Ala Phe Glu Gly Val Val Arg Ala Leu Leu Gln Lys
 265 270 275 280
 Pro Ser Ala Arg Val Ala Val Leu Phe Thr Arg Ser Glu Asp Ala Arg Glu Leu Leu Ala
 285 290 295 300
 Ala Ser Gln Arg Leu Asn Ala Ser Phe Thr Trp Val Ala Ser Asp Gly Trp Gly Ala Leu
 305 310 315 320
 Glu Ser Val Val Ala Gly Ser Glu Gly Ala Ala Glu Gly Ala Ile Thr Ile Glu Leu Ala

Ser Tyr Pro Ile	325	Ser Asp Phe Ala	330	Phe Gln Ser Leu	335	Asp Pro Trp Asn Asn	340
Arg Asn Pro Trp	345	Phe Arg Glu Phe	350	Glu Gln Arg Phe	355	Cys Ser Phe Arg Gln	360
Asp Cys Ala Ala	365	His Ser Leu Arg	370	Ala Val Pro Phe	375	Glu Gln Glu Ser Lys	380
Val Val Asn Ala	385	Val Tyr Ala Met	390	Ala His Ala Leu	395	His Asn Met His Arg	400
Pro Asn Thr Thr	405	Arg Leu Cys Asp	410	Ala Met Arg Pro	415	Val Asn Gly Arg Arg	420
Asp Phe Val Leu	425	Asn Val Lys Phe	430	Ala Pro Phe Arg	435	Pro Ala Asp Thr His	440
Val Arg Phe Asp	445	Arg Phe Gly Asp	450	Gly Ile Gly Arg	455	Tyr Asn Ile Phe Thr	460
Ala Gly Ser Gly	465	Arg Tyr Arg Tyr	470	Gln Lys Val Gly	475	Tyr Trp Ala Glu Gly	480
Asp Thr Ser Leu	485	Ile Pro Trp Ala	490	Ser Pro Ser Ala	495	Gly Pro Leu Pro Ala	500
Ser Glu Pro Cys	505	Leu Gln Asn Glu	510	Val Lys Ser Val	515	Gln Pro Gly Glu Val	520
Leu Cys Ile Pro	525	Cys Gln Pro Tyr	530	Glu Tyr Arg Leu	535	Asp Glu Phe Thr Cys	540
Gly Leu Gly Tyr	545	Trp Pro Asn Ala	550	Ser Leu Thr Gly	555	Cys Phe Glu Leu Pro	560
Ile Arg Trp Gly	565	Asp Ala Trp Ala	570	Val Gly Pro Val	575	Thr Ile Ala Cys Leu	580
Ala Thr Leu Phe	585	Val Leu Gly Val	590	Phe Val Arg His	595	Asn Ala Thr Pro Val	600
Ser Gly Arg Glu	605	Leu Cys Tyr Ile	610	Leu Leu Gly Gly	615	Val Phe Leu Cys Tyr	620
Phe Ile Phe Ile	625	Ala Lys Pro Ser	630	Thr Ala Val Cys	635	Thr Leu Arg Arg Leu	640
Thr Ala Phe Ser	645	Val Cys Tyr Ser	650	Ala Leu Leu Thr	655	Lys Thr Asn Arg Ile	660
Phe Gly Gly Ala	665	Arg Glu Gly Ala	670	Gln Arg Pro Arg	675	Phe Ile Ser Pro Ala	680
Ala Ile Cys Leu	685	Ala Leu Ile Ser	690	Val Gln Leu Leu	695	Ile Val Val Ala Trp	700
Glu Ala Pro Gly	705	Thr Gly Lys Glu	710	Thr Ala Pro Glu	715	Arg Arg Glu Val Val	720
	725		730		735		740

Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asn Val Leu Leu Ile Ala
 745 750 755 760
 Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys
 765 770 775 780
 Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe
 785 790 795 800
 Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu
 805 810 815 820
 Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln
 825 830 835 840
 Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Ala
 845 850 855 860
 Arg Ala Ser Ser Ser Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn
 865 870
 Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter

Seq ID 8: S688L, G689V, A733T, N735D

5 10 15 20
 Met Gly Ser Leu Leu Ala Leu Leu Ala Leu Leu Pro Leu Trp Gly Ala Val Ala Glu Gly
 25 30 35 40
 Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val
 45 50 55 60
 His Gln Lys Gly Gly Pro Ala Glu Asp Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln
 65 70 75 80
 Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro
 85 90 95 100
 Gly Val Arg Leu Gly Ala His Ile Leu Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu
 105 110 115 120
 Gln Ala Leu Asp Phe Val Arg Ala Ser Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile
 125 130 135 140
 Cys Pro Asp Gly Ser Tyr Ala Thr His Gly Asp Ala Pro Thr Ala Ile Thr Gly Val Ile
 145 150 155 160
 Gly Gly Ser Tyr Ser Asp Val Ser Ile Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile
 165 170 175 180
 Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp Lys Ser Arg Tyr Asp Tyr
 185 190 195 200
 Phe Ala Arg Thr Val Pro Pro Asp Phe Phe Gln Ala Lys Ala Met Ala Glu Ile Leu Arg
 205 210 215 220
 Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly Asp Tyr Gly Glu Thr Gly
 225 230 235 240
 Ile Glu Ala Phe Glu Leu Glu Ala Arg Ala Arg Asn Ile Cys Val Ala Thr Ser Glu Lys

245 250 255 260
 Val Gly Arg Ala Met Ser Arg Ala Ala Phe Glu Gly Val Val Arg Ala Leu Leu Gln Lys
 270 275 280
 Pro Ser Ala Arg Val Ala Val Leu Phe Thr Arg Ser Glu Asp Ala Arg Glu Leu Leu Ala
 285 290 295 300
 Ala Ser Gln Arg Leu Asn Ala Ser Phe Thr Trp Val Ala Ser Asp Gly Trp Gly Ala Leu
 305 310 315 320
 Glu Ser Val Val Ala Gly Ser Glu Gly Ala Ala Glu Gly Ala Ile Thr Ile Glu Leu Ala
 325 330 335 340
 Ser Tyr Pro Ile Ser Asp Phe Ala Ser Tyr Phe Gln Ser Leu Asp Pro Trp Asn Asn Ser
 345 350 355 360
 Arg Asn Pro Trp Phe Arg Glu Phe Trp Glu Gln Arg Phe Arg Cys Ser Phe Arg Gln Arg
 365 370 375 380
 Asp Cys Ala Ala His Ser Leu Arg Ala Val Pro Phe Glu Gln Glu Ser Lys Ile Met Phe
 385 390 395 400
 Val Val Asn Ala Val Tyr Ala Met Ala His Ala Leu His Asn Met His Arg Ala Leu Cys
 405 410 415 420
 Pro Asn Thr Thr Arg Leu Cys Asp Ala Met Arg Pro Val Asn Gly Arg Arg Leu Tyr Lys
 425 430 435 440
 Asp Phe Val Leu Asn Val Lys Phe Asp Ala Pro Phe Arg Pro Ala Asp Thr His Asn Glu
 445 450 455 460
 Val Arg Phe Asp Arg Phe Gly Asp Gly Ile Gly Arg Tyr Asn Ile Phe Thr Tyr Leu Arg
 465 470 475 480
 Ala Gly Ser Gly Arg Tyr Arg Tyr Gln Lys Val Gly Tyr Trp Ala Glu Gly Leu Thr Leu
 485 490 495 500
 Asp Thr Ser Leu Ile Pro Trp Ala Ser Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys
 505 510 515 520
 Ser Glu Pro Cys Leu Gln Asn Glu Val Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp
 525 530 535 540
 Leu Cys Ile Pro Cys Gln Pro Tyr Glu Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys
 545 550 555 560
 Gly Leu Gly Tyr Trp Pro Asn Ala Ser Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr
 565 570 575 580
 Ile Arg Trp Gly Asp Ala Trp Ala Val Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu
 585 590 595 600
 Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala
 605 610 615 620
 Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr
 625 630 635 640
 Phe Ile Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly
 645 650 655 660

Thr	Ala	Phe	Ser	Val	Cys	Tyr	Ser	Ala	Leu	Leu	Thr	Lys	Thr	Asn	Arg	Ile	Ala	Arg	Ile
				665					670					675					680
Phe	Gly	Gly	Ala	Arg	Glu	Gly	Ala	Gln	Arg	Pro	Arg	Phe	Ile	Ser	Pro	Ala	Ser	Gln	Val
				685					690					695					700
Ala	Ile	Cys	Leu	Ala	Leu	Ile	Leu	Val	Gln	Leu	Leu	Ile	Val	Val	Ala	Trp	Leu	Val	Val
				705					710					715					720
Glu	Ala	Pro	Gly	Thr	Gly	Lys	Glu	Thr	Ala	Pro	Glu	Arg	Arg	Glu	Val	Val	Thr	Leu	Arg
				725					730					735					740
Cys	Asn	His	Arg	Asp	Ala	Ser	Met	Leu	Gly	Ser	Leu	Thr	Tyr	Asp	Val	Leu	Leu	Ile	Ala
				745					750					755					760
Leu	Cys	Thr	Leu	Tyr	Ala	Phe	Lys	Thr	Arg	Lys	Cys	Pro	Glu	Asn	Phe	Asn	Glu	Ala	Lys
				765					770					775					780
Phe	Ile	Gly	Phe	Thr	Met	Tyr	Thr	Thr	Cys	Ile	Ile	Trp	Leu	Ala	Phe	Leu	Pro	Ile	Phe
				785					790					795					800
Tyr	Val	Thr	Ser	Ser	Asp	Tyr	Arg	Val	Gln	Thr	Thr	Thr	Met	Cys	Val	Ser	Val	Ser	Leu
				805					810					815					820
Ser	Gly	Ser	Val	Val	Leu	Gly	Cys	Leu	Phe	Ala	Pro	Lys	Leu	His	Ile	Ile	Leu	Phe	Gln
				825					830					835					840
Pro	Gln	Lys	Asn	Val	Val	Ser	His	Arg	Ala	Pro	Thr	Ser	Arg	Phe	Gly	Ser	Ala	Ala	Ala
				845					850					855					860
Arg	Ala	Ser	Ser	Ser	Leu	Gly	Gln	Gly	Ser	Gly	Ser	Gln	Phe	Val	Pro	Thr	Val	Cys	Asn
				865					870										
Gly	Arg	Glu	Val	Val	Asp	Ser	Thr	Thr	Ser	Ser	Leu	Ter							

Seq ID 9: N735D

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1  atgggatcgc tgcttgcgct cctggcactg ctgccgctgt ggggtgctgt ggctgagggc
61  ccagccaaga aggtgctgac cctggaggga gacttggtgc tgggtgggct gttcccagtg
121 caccagaagg gcgggccagc agaggactgt ggtcctgtca atgagcaccg tggcatccag
181 cgcctggagg ccatgctttt tgcactggac cgcacaaacc gtgacccgca cctgctgcct
241 ggcgtgcgcc tgggtgcaca catcctcgac agttgctcca aggacacaca tgcgctggag
301 caggcactgg actttgtgcg tgcctcactc agccgtggtg ctgatggctc acgccacatc
361 tgccccgacg gctcttatgc gacccatggt gatgctccca ctgccatcac tgggtgttatt
421 ggcggttcct acagtgatgt ctccatccag gtggccaacc tcttgaggct atttcagatc
481 ccacagatta gctacgcctc taccagtgcc aagctgagtg acaagtcccg ctatgactac
541 tttgcccgca cagtgcctcc tgacttcttc caagccaagg ccatggctga gattctccgc
601 ttcttcaact ggacctatgt gtccactgtg gcgtctgagg gcgactatgg cgagacaggg
661 attgaggcct ttgagctaga ggctcgtgcc cgcaacatct gtgtggccac ctcgagaaaa
721 gtggggcgtg ccatgagccg cgcggccttt gaggggtgtg tgcgagccct gctgcagaag
781 cccagtgcgc gcgtggctgt cctgttcacc cgttctgagg atgccgggga gctgcttgct
841 gccagccagc gcctcaatgc cagcttcacc tgggtggcca gtgatgggtg gggggccctg
901 gagagtgtgg tggcaggcag tgagggggct gctgaggggt ctatcaccat cgagctggcc
961 tcctacccca tcagtgaact tgccctctac ttccagagcc tggacccttg gaacaacagc
1021 cggaaccctt ggttccgtga attctgggag cagaggttcc gctgcagctt ccggcagcga
1081 gactgcgcag cccactctct ccgggctgtg ccctttgagc aggagtccaa gatctgttt
1141 gtggtcaatg cagtgtacgc catggcccat gcgtccaca acatgcaccg tgccctctgc
1201 cccaacacca ccggctcttg tgacgcgatg cggccagtta acgggcgccc cctctacaag
1261 gactttgtgc tcaacgtcaa gtttgatgcc ccctttcgcc cagctgacac ccacaatgag
1321 gtccgctttg accgcttttg tgatgggtat ggccgctaca acatcttcac ctatctgcgt
1381 gcaggcagtg ggcgctatcg ctaccagaag gtgggctact gggcagaagg cttgactctg
1441 gacaccagcc tcatcccatg ggcctcacc tcagccggcc ccctgcccgc ctctcgctgc
1501 agtgagccct gcctccagaa tgaggtgaag agtgtgcagc cgggcgaagt ctgctgctgg
1561 ctctgcattc cgtgccagcc ctatgagtac cgattggacg aattcacttg cgctgattgt
1621 ggctgggct actggcccaa tgccagcctg actggctgct tcgaactgcc ccaggagtac
1681 atccgctggg gcgatgcctg ggctgtggga cctgtcacca tcgcctgcct cggtgccctg
1741 gccaccctct ttgtgctggg tgtctttgtg cggcacaatg ccacaccagt ggtcaaggcc
1801 tcaggctcgg agctctgcta catcctgctg ggtggtgtct tcctctgcta ctgcatgacc
1861 ttcattctca ttgccaagcc atccacggca gtgtgtacct tacggcgtct tggtttgggc
1921 actgccttct ctgtctgcta ctcagccctg ctcaccaaga ccaaccgcat tgcacgcata
1981 ttcggtgggg ccggggaggg tgcccagcgg ccacgcttca tcagtccctg ctcacagggtg
2041 gccatctgcc tggcacttat ctcgggccag ctgctcatcg tggctgcctg gctggtggtg
2101 gaggcaccgg gcacaggcaa ggagacagcc ccggaacggc gggagggtgg gacactgcgc
2161 tgcaaccacc gcgatgcaag tatgttgggc tcgctggcct acgatgtgct cctcatcgcg
2221 ctctgcacgc tttatgcctt caagactcgc aagtgcctcg aaaacttcaa cgaggccaag
2281 ttcatgtggt tcaccatgta caccacctgc atcatctggc tggcattctt gcccatcttc
2341 tatgtcacct ccagtgacta ccgggtacag accaccacca tgtgcgtgtc agtcagcctc
2401 agcggctccg tgggtgcttg ctgcctcttt gcgcccaagc tgcacatcat cctcttcag
2461 ccgcagaaga acgtggttag ccaccgggca cccaccagcc gctttggcag tgctgctgcc
2521 agggccagct ccagccttgg ccaagggtct ggctccagct ttgtcccccac tgtttgcaat
2581 ggccgtgagg tgggtgactc gacaacgtca tcgctttga

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Seq ID 10: G689V N735D

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1 atgggatcgc tgcttgcgct cctggcactg ctgccgctgt ggggtgctgt ggctgagggc
61 ccagccaaga aggtgctgac cctggaggga gacttggtgc tgggtgggct gttcccagtg
121 caccagaagg gcggcccagc agaggactgt ggtcctgtca atgagcaccg tggcatccag
181 cgcctggagg ccatgctttt tgcactggac cgcatacaacc gtgaccgcga cctgctgcct
241 ggcgtgcgcc tgggtgcaca catcctcgac agttgctcca aggacacaca tgcgctggag
301 caggcactgg actttgtgcg tgcctcactc agccgtggtg ctgatggctc acgccacatc
361 tgccccgacg gctcttatgc gacccatggg gatgctccca ctgccatcac tgggtgttatt
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361 tgccccgacg  gctcttatgc  gacctatggt  gatgctccca  ctgccatcac  tgggtgttatt
421 ggcggttcct  acagtgatgt  ctccatccag  gtggccaacc  tcttgaggct  atttcagatc
481 ccacagatta  gctacgcctc  taccagtgcc  aagctgagtg  acaagtcccg  ctatgactac
541 tttgcccgca  cagtgcctcc  tgacttcttc  caagccaagg  ccatggctga  gattctccgc
601 ttcttcaact  ggacctatgt  gtccactgtg  gcgtctgagg  gcgactatgg  cgagacaggc
661 attgaggcct  ttgagctaga  ggctcgtgcc  cgcaacatct  gtgtggccac  ctcgagaaaa
721 gtggggccgtg  ccatgagccg  cgcgcccttt  gagggtgtgg  tgcgagccct  gctgcagaag
781 cccagtgcctc  gcgtggctgt  cctgttcacc  cgttctgagg  atgcccgga  gctgcttgct
841 gccagccagc  gcctcaatgc  cagcttcacc  tgggtggcca  gtgatgggtg  gggggccctg
901 gagagtgtgg  tggcaggcag  tgagggggct  gctgaggggtg  ctatcaccat  cgagctggcc
961 tcctacccca  tcagtgaact  tgcctcctac  ttccagagcc  tggacccttg  gaacaacagc
1021 cggaaccctt  gggtccgtga  attctgggag  cagagggttc  gctgcagctt  ccggcagcga
1081 gactgcgcag  cccactctct  ccgggctgtg  ccctttgagc  aggagtccaa  gatcatgttt
1141 gtggtcaatg  cagtgtacgc  catggcccat  gcgtccaca  acatgcaccg  tgccctctgc
1201 cccaacacca  cccggctctg  tgacgcgatg  cggccagtta  acgggcgcgc  cctctacaag
1261 gactttgtgc  tcaacgtcaa  gtttgatgcc  ccctttcgcc  cagctgacac  ccacaatgag
1321 gtccgctttg  accgcttttg  tgatggtatt  ggccgctaca  acatcttcac  ctatctgcgt
1381 gcaggcagtg  ggcgctatcg  ctaccagaag  gtgggctact  gggcagaagg  cttgactctg
1441 gacaccagcc  tcatcccatg  ggctcacc  tcagccggcc  ccctgcccgc  ctctcgctgc
1501 agtgagccct  gcctccagaa  tgaggtgaag  agtgtgcagc  cgggcgaagt  ctgctgctgg
1561 ctctgcattc  cgtgccagcc  ctatgagtac  cgattggacg  aattcacttg  cgctgattgt
1621 ggcctgggct  actggcccaa  tgccagcctg  actggctgct  tcgaactgcc  ccaggagtac
1681 atccgctggg  gcgatgcctg  ggctgtggga  cctgtcacca  tcgcctgcct  cgggtgccctg
1741 gccaccctct  ttgtgctggg  tgtctttgtg  cggcacaatg  ccacaccagt  ggtcaaggcc
1801 tcaggctcggg  agctctgcta  catcctgctg  ggtgggtgtct  tcctctgcta  ctgcatgacc
1861 ttcattcttca  ttgccaagcc  atccacggca  gtgtgtacct  tacggcgtct  tgggttgggc
1921 actgccttct  ctgtctgcta  ctacagccctg  ctacaccaaga  ccaaccgcat  tgcacgcac
1981 ttccggtggg  cccgggaggg  tgcccagcgg  ccacgcttca  tcagtccctg  ctacagggtg
2041 gccatctgcc  tggcacttat  ctccgtccag  ctgctcatcg  tggtcgcctg  gctgggtggg
2101 gaggcaccgg  gcacaggcaa  ggagacagcc  ccgaacggc  gggagggtgg  gacactgcgc
2161 tgcaaccacc  gcgatgcaag  tatgttgggc  tcgctggcct  acaatgtgct  cctcatcgcg
2221 ctctgcacgc  tttatgcctt  caagactcgc  aagtgcctcg  aaaacttcaa  cgaggccaag
2281 ttcattggct  tcaccatgta  caccacctgc  atcatctggc  tggcattctt  gcccatcttc
2341 tatgtcacct  ccagtgacta  ccgggtacag  accaccacca  tgtgcgtgtc  agtcagcctc
2401 agcggctccg  tgggtgcttg  ctgcctcttt  gcgcccaagc  tgcacatcat  cctcttccag
2461 ccgcagaaga  acgtgggttag  ccaccgggca  cccaccagcc  gctttggcag  tgctgctgcc
2521 agggccagct  ccagccttgg  ccaagggtct  ggctcccagt  ttgtcccccac  tgtttgcaat
2581 ggcctgagg  tgggtgactc  gacaacgtca  tcgctttga
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Seq ID 16: S688L, G689V, A733T, N735D

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1 atgggatcgc tgcttgcgct cctggcactg ctgccgctgt ggggtgctgt ggctgagggc
61 ccagccaaga aggtgctgac cctggagggg gacttggtgc tgggtgggct gttcccagtg
121 caccagaagg gcggcccagc agaggactgt ggtcctgtca atgagcaccg tggcatccag
181 cgcctggagg ccatgctttt tgcactggac cgcatacaacc gtgaccgcga cctgctgcct
241 ggcgtgcgcc tgggtgcaca catcctcgac agttgctcca aggacacaca tgcgctggag
301 caggcactgg actttgtgcg tgcctcactc agccgtggtg ctgatggctc acgccacatc
361 tgccccgacg gctcttatgc gacctatggt gatgtcccca ctgccatcac tgggtgttatt
421 ggcggttcct acagtgatgt ctccatccag gtggccaacc tcttgaggct atttcagatc
481 ccacagatta gctacgcctc taccagtgcc aaagtgagtg acaagtcctc gatgactac
541 tttgcccgca cagtgcctcc tgacttcttc caagccaagg ccatggctga gattctccgc
601 ttcttcaact ggacctatgt gtccactgtg gcgtctgagg gcgactatgg cgagacaggc
661 attgaggcct ttgagctaga ggctcgtgcc cgcaacatct gtgtggccac ctcgagagaa
721 gtgggcccgt ccatgagccg cgcggccttt gaggggtgtg tgcgagccct gctgcagaag
781 cccagtgcgc gcgtggctgt cctgttcacc cgttctgagg atgcccgga gctgcttget
841 gccagccagc gcctcaatgc cagcttcacc tgggtggcca gtgatggtg gggggccctg
901 gagagtgtgg tggcaggcag tgagggggct gctgaggggt ctatcaccat cgagctggcc
961 tcctacccca tcagtactt tgcctcctac ttccagagcc tggacccttg gaacaacagc
1021 cggaaccctt ggttccgtga attctgggag cagaggttcc gctgcagctt ccggcagcga
1081 gactgcgcag cccactctct ccgggctgtg ccctttgagc aggagtccaa gatcatgttt
1141 gtggtcaatg cagtgtacgc catggcccat gcgtccaca acatgcaccg tgccctctgc
1201 cccaacacca ccggctctg tgacgcgatg cggccagtta acgggcgcgc cctctacaag
1261 gactttgtgc tcaacgtcaa gtttgatgcc ccctttcgcc cagctgacac ccacaatgag
1321 gtccgctttg accgcttttg tgatggtatt ggccgctaca acatcttcac ctatctgcgt
1381 gcaggcagtg ggcgctatcg ctaccagaag gtgggctact gggcagaagg cttgactctg
1441 gacaccagcc tcatcccatg ggcctcacc tcagccggcc ccctgcccgc ctctcgctgc
1501 agtgagccct gcctccagaa tgaggtgaag agtggtgcagc cgggcgaagt ctgctgctgg
1561 ctctgcattc cgtgccagcc ctatgagtac cgattggacg aattcacttg cgctgattgt
1621 ggcctgggct actggcccaa tgccagcctg actggctgct tcgaactgcc ccaggagtac
1681 atccgctggg gcgatgcctg ggctgtggga cctgtcacca tcgcctgcct cgggtgccctg
1741 gccaccctct ttgtgctggg tgtctttgtg cggcacaatg ccacaccagt ggtcaaggcc
1801 tcaggtcggg agctctgcta catcctgctg ggtggtgtct tcctctgcta ctgcatgacc
1861 ttcatcttca ttgccaagcc atccacggca gtgtgtacct tacggcgtct tggtttgggc
1921 actgccttct ctgtctgcta ctcagccctg ctcaccaaga ccaaccgcat tgcacgcatac
1981 ttcggtgggg ccggggaggg tgcccagcgg ccacgcttca tcagtcctgc ctcacagggtg
2041 gccatctgcc tggcacttat cttggtccag ctgctcatcg tggtcgcctg gctggtggtg
2101 gaggcaccgg gcacaggcaa ggagacagcc ccgaacggc gggaggtggt gacactgcgc
2161 tgcaaccacc gcgatgcaag tatgttgggc tcgctgacct acgatgtgct cctcatcgcg
2221 ctctgcacgc tttatgcctt caagactcgc aagtgcctcg aaaacttcaa cgaggccaag
2281 ttcatgtggt tcacatgta caccacctgc atcatctggc tggcattctt gcccatcttc
2341 tatgtcacct ccagtgacta ccgggtacag accaccacca tgtgcgtgtc agtcagcctc
2401 agcggctcgc tgggtgcttg ctgcctcttt gcgccaagc tgcacatcat cctcttcag
2461 ccgcagaaga acgtggttag ccaccgggca cccaccagcc gctttggcag tgctgctgcc
2521 agggccagct ccagccttgg ccaaggtctt ggctcccagt ttgtcccccac tgtttgcaat
2581 ggccgtgagg tgggtggactc gacaacgtca tcgctttga
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Rat Sequences

Seq ID 17: N735D

5	10	15	20
Met Glu Ser Leu Leu Gly Phe Leu Ala Leu Leu Leu Leu Trp Gly Ala Val Ala Glu Gly			
25	30	35	40
Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val			
45	50	55	60
His Gln Lys Gly Gly Pro Ala Glu Glu Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln			
65	70	75	80
Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro			
85	90	95	100
Gly Val Arg Leu Gly Ala His Ile Leu Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu			
105	110	115	120
Gln Ala Leu Asp Phe Val Arg Ala Ser Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile			
125	130	135	140
Cys Pro Asp Gly Ser Tyr Ala Thr His Ser Asp Ala Pro Thr Ala Val Thr Gly Val Ile			
145	150	155	160
Gly Gly Ser Tyr Ser Asp Val Ser Ile Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile			
165	170	175	180
Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp Lys Ser Arg Tyr Asp Tyr			
185	190	195	200
Phe Ala Arg Thr Val Pro Pro Asp Phe Phe Gln Ala Lys Ala Met Ala Glu Ile Leu Arg			
205	210	215	220
Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly Asp Tyr Gly Glu Thr Gly			
225	230	235	240
Ile Glu Ala Phe Glu Leu Glu Ala Arg Ala Arg Asn Ile Cys Val Ala Thr Ser Glu Lys			
245	250	255	260
Val Gly Arg Ala Met Ser Arg Ala Ala Phe Glu Gly Val Val Arg Ala Leu Leu Gln Lys			
265	270	275	280
Pro Ser Ala Arg Val Ala Val Leu Phe Thr Arg Ser Glu Asp Ala Arg Glu Leu Leu Ala			
285	290	295	300
Ala Thr Gln Arg Leu Asn Ala Ser Phe Thr Trp Val Ala Ser Asp Gly Trp Gly Ala Leu			
305	310	315	320
Glu Ser Val Val Ala Gly Ser Glu Arg Ala Ala Glu Gly Ala Ile Thr Ile Glu Leu Ala			
325	330	335	340
Ser Tyr Pro Ile Ser Asp Phe Ala Ser Tyr Phe Gln Ser Leu Asp Pro Trp Asn Asn Ser			
345	350	355	360
Arg Asn Pro Trp Phe Arg Glu Phe Trp Glu Glu Arg Phe His Cys Ser Phe Arg Gln Arg			
365	370	375	380
Asp Cys Ala Ala His Ser Leu Arg Ala Val Pro Phe Glu Gln Glu Ser Lys Ile Met Phe			
385	390	395	400
Val Val Asn Ala Val Tyr Ala Met Ala His Ala Leu His Asn Met His Arg Ala Leu Cys			

405	410	415	420
Pro Asn Thr Thr His Leu Cys Asp Ala Met Arg Pro Val Asn Gly Arg Arg Leu Tyr Lys			
425	430	435	440
Asp Phe Val Leu Asn Val Lys Phe Asp Ala Pro Phe Arg Pro Ala Asp Thr Asp Asp Glu			
445	450	455	460
Val Arg Phe Asp Arg Phe Gly Asp Gly Ile Gly Arg Tyr Asn Ile Phe Thr Tyr Leu Arg			
465	470	475	480
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln Lys Val Gly Tyr Trp Ala Glu Gly Leu Thr Leu			
485	490	495	500
Asp Thr Ser Phe Ile Pro Trp Ala Ser Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys			
505	510	515	520
Ser Glu Pro Cys Leu Gln Asn Glu Val Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp			
525	530	535	540
Leu Cys Ile Pro Cys Gln Pro Tyr Glu Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys			
545	550	555	560
Gly Leu Gly Tyr Trp Pro Asn Ala Ser Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr			
565	570	575	580
Ile Arg Trp Gly Asp Ala Trp Ala Val Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu			
585	590	595	600
Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala			
605	610	615	620
Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr			
625	630	635	640
Phe Val Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly			
645	650	655	660
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile			
665	670	675	680
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val			
685	690	695	700
Ala Ile Cys Leu Ala Leu Ile Ser Gly Gln Leu Leu Ile Val Ala Ala Trp Leu Val Val			
705	710	715	720
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg			
725	730	735	740
Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asp Val Leu Leu Ile Ala			
745	750	755	760
Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys			
765	770	775	780
Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe			
785	790	795	800
Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu			

805	810	815	820
Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln			
825	830	835	840
Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Pro			
845	850	855	860
Arg Ala Ser Ala Asn Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn			
865	870		
Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter			

Seq ID 18: G689V, N735D

5	10	15	20
Met Glu Ser Leu Leu Gly Phe Leu Ala Leu Leu Leu Leu Trp Gly Ala Val Ala Glu Gly			
25	30	35	40
Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val			
45	50	55	60
His Gln Lys Gly Gly Pro Ala Glu Glu Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln			
65	70	75	80
Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro			
85	90	95	100
Gly Val Arg Leu Gly Ala His Ile Leu Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu			
105	110	115	120
Gln Ala Leu Asp Phe Val Arg Ala Ser Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile			
125	130	135	140
Cys Pro Asp Gly Ser Tyr Ala Thr His Ser Asp Ala Pro Thr Ala Val Thr Gly Val Ile			
145	150	155	160
Gly Gly Ser Tyr Ser Asp Val Ser Ile Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile			
165	170	175	180
Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp Lys Ser Arg Tyr Asp Tyr			
185	190	195	200
Phe Ala Arg Thr Val Pro Pro Asp Phe Phe Gln Ala Lys Ala Met Ala Glu Ile Leu Arg			
205	210	215	220
Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly Asp Tyr Gly Glu Thr Gly			
225	230	235	240
Ile Glu Ala Phe Glu Leu Glu Ala Arg Ala Arg Asn Ile Cys Val Ala Thr Ser Glu Lys			
245	250	255	260
Val Gly Arg Ala Met Ser Arg Ala Ala Phe Glu Gly Val Val Arg Ala Leu Leu Gln Lys			
265	270	275	280
Pro Ser Ala Arg Val Ala Val Leu Phe Thr Arg Ser Glu Asp Ala Arg Glu Leu Leu Ala			
285	290	295	300
Ala Thr Gln Arg Leu Asn Ala Ser Phe Thr Trp Val Ala Ser Asp Gly Trp Gly Ala Leu			
305	310	315	320
Glu Ser Val Val Ala Gly Ser Glu Arg Ala Ala Glu Gly Ala Ile Thr Ile Glu Leu Ala			

325	330	335	340
Ser Tyr Pro Ile Ser Asp Phe Ala Ser Tyr Phe Gln Ser Leu Asp Pro Trp Asn Asn Ser			
345	350	355	360
Arg Asn Pro Trp Phe Arg Glu Phe Trp Glu Glu Arg Phe His Cys Ser Phe Arg Gln Arg			
365	370	375	380
Asp Cys Ala Ala His Ser Leu Arg Ala Val Pro Phe Glu Gln Glu Ser Lys Ile Met Phe			
385	390	395	400
Val Val Asn Ala Val Tyr Ala Met Ala His Ala Leu His Asn Met His Arg Ala Leu Cys			
405	410	415	420
Pro Asn Thr Thr His Leu Cys Asp Ala Met Arg Pro Val Asn Gly Arg Arg Leu Tyr Lys			
425	430	435	440
Asp Phe Val Leu Asn Val Lys Phe Asp Ala Pro Phe Arg Pro Ala Asp Thr Asp Asp Glu			
445	450	455	460
Val Arg Phe Asp Arg Phe Gly Asp Gly Ile Gly Arg Tyr Asn Ile Phe Thr Tyr Leu Arg			
465	470	475	480
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln Lys Val Gly Tyr Trp Ala Glu Gly Leu Thr Leu			
485	490	495	500
Asp Thr Ser Phe Ile Pro Trp Ala Ser Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys			
505	510	515	520
Ser Glu Pro Cys Leu Gln Asn Glu Val Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp			
525	530	535	540
Leu Cys Ile Pro Cys Gln Pro Tyr Glu Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys			
545	550	555	560
Gly Leu Gly Tyr Trp Pro Asn Ala Ser Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr			
565	570	575	580
Ile Arg Trp Gly Asp Ala Trp Ala Val Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu			
585	590	595	600
Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala			
605	610	615	620
Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr			
625	630	635	640
Phe Val Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly			
645	650	655	660
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile			
665	670	675	680
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val			
685	690	695	700
Ala Ile Cys Leu Ala Leu Ile Ser Val Gln Leu Leu Ile Val Ala Ala Trp Leu Val Val			
705	710	715	720
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg			

725	730	735	740
Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asp Val Leu Leu Ile Ala			
745	750	755	760
Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys			
765	770	775	780
Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe			
785	790	795	800
Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu			
805	810	815	820
Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln			
825	830	835	840
Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Pro			
845	850	855	860
Arg Ala Ser Ala Asn Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn			
865	870		
Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter			

Seq ID 19: S688L, G689V, N735D

5	10	15	20
Met Glu Ser Leu Leu Gly Phe Leu Ala Leu Leu Leu Leu Trp Gly Ala Val Ala Glu Gly			
25	30	35	40
Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val			
45	50	55	60
His Gln Lys Gly Gly Pro Ala Glu Glu Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln			
65	70	75	80
Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro			
85	90	95	100
Gly Val Arg Leu Gly Ala His Ile Leu Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu			
105	110	115	120
Gln Ala Leu Asp Phe Val Arg Ala Ser Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile			
125	130	135	140
Cys Pro Asp Gly Ser Tyr Ala Thr His Ser Asp Ala Pro Thr Ala Val Thr Gly Val Ile			
145	150	155	160
Gly Gly Ser Tyr Ser Asp Val Ser Ile Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile			
165	170	175	180
Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp Lys Ser Arg Tyr Asp Tyr			
185	190	195	200
Phe Ala Arg Thr Val Pro Pro Asp Phe Phe Gln Ala Lys Ala Met Ala Glu Ile Leu Arg			
205	210	215	220
Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly Asp Tyr Gly Glu Thr Gly			
225	230	235	240
Ile Glu Ala Phe Glu Leu Glu Ala Arg Ala Arg Asn Ile Cys Val Ala Thr Ser Glu Lys			

245	250	255	260
Val Gly Arg Ala Met Ser Arg Ala Ala	Phe Glu Gly Val Val	Arg Ala Leu Leu	Gln Lys
265	270	275	280
Pro Ser Ala Arg Val Ala Val Leu Phe	Thr Arg Ser Glu Asp	Ala Arg Glu Leu Leu	Ala
285	290	295	300
Ala Thr Gln Arg Leu Asn Ala Ser Phe	Thr Trp Val Ala Ser	Asp Gly Trp Gly Ala	Leu
305	310	315	320
Glu Ser Val Val Ala Gly Ser Glu Arg	Ala Ala Glu Gly Ala	Ile Thr Ile Glu Leu	Ala
325	330	335	340
Ser Tyr Pro Ile Ser Asp Phe Ala Ser	Tyr Phe Gln Ser Leu Asp	Pro Trp Asn Asn	Ser
345	350	355	360
Arg Asn Pro Trp Phe Arg Glu Phe Trp	Glu Glu Arg Phe His Cys	Ser Phe Arg Gln	Arg
365	370	375	380
Asp Cys Ala Ala His Ser Leu Arg Ala	Val Pro Phe Glu Gln Glu	Ser Lys Ile Met	Phe
385	390	395	400
Val Val Asn Ala Val Tyr Ala Met Ala	His Ala Leu His Asn Met	His Arg Ala Leu	Cys
405	410	415	420
Pro Asn Thr Thr His Leu Cys Asp Ala	Met Arg Pro Val Asn Gly	Arg Arg Leu Tyr	Lys
425	430	435	440
Asp Phe Val Leu Asn Val Lys Phe Asp	Ala Pro Phe Arg Pro Ala	Asp Thr Asp Asp	Glu
445	450	455	460
Val Arg Phe Asp Arg Phe Gly Asp Gly	Ile Gly Arg Tyr Asn Ile	Phe Thr Tyr Leu	Arg
465	470	475	480
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln	Lys Val Gly Tyr Trp Ala	Glu Gly Leu Thr	Leu
485	490	495	500
Asp Thr Ser Phe Ile Pro Trp Ala Ser	Pro Ser Ala Gly Pro Leu	Pro Ala Ser Arg	Cys
505	510	515	520
Ser Glu Pro Cys Leu Gln Asn Glu Val	Lys Ser Val Gln Pro Gly	Glu Val Cys Cys	Trp
525	530	535	540
Leu Cys Ile Pro Cys Gln Pro Tyr Glu	Tyr Arg Leu Asp Glu	Phe Thr Cys Ala	Asp Cys
545	550	555	560
Gly Leu Gly Tyr Trp Pro Asn Ala Ser	Leu Thr Gly Cys Phe	Glu Leu Pro Gln	Glu Tyr
565	570	575	580
Ile Arg Trp Gly Asp Ala Trp Ala Val	Gly Pro Val Thr Ile Ala	Cys Leu Gly Ala	Leu
585	590	595	600
Ala Thr Leu Phe Val Leu Gly Val Phe	Val Arg His Asn Ala Thr	Pro Val Val Lys	Ala
605	610	615	620
Ser Gly Arg Glu Leu Cys Tyr Ile Leu	Leu Gly Gly Val Phe	Leu Cys Tyr Cys	Met Thr
625	630	635	640
Phe Val Phe Ile Ala Lys Pro Ser Thr	Ala Val Cys Thr Leu	Arg Arg Leu Gly	Leu Gly

645	650	655	660
Thr Ala Phe Ser Val Cys Tyr Ser Ala	Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile		
665	670	675	680
Phe Gly Gly Ala Arg Glu Gly Ala Gln	Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val		
685	690	695	700
Ala Ile Cys Leu Ala Leu Ile Leu Val	Gln Leu Leu Ile Val Ala Ala Trp Leu Val Val		
705	710	715	720
Glu Ala Pro Gly Thr Gly Lys Glu Thr	Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg		
725	730	735	740
Cys Asn His Arg Asp Ala Ser Met Leu	Gly Ser Leu Ala Tyr Asp Val Leu Leu Ile Ala		
745	750	755	760
Leu Cys Thr Leu Tyr Ala Phe Lys Thr	Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys		
765	770	775	780
Phe Ile Gly Phe Thr Met Tyr Thr Thr	Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe		
785	790	795	800
Tyr Val Thr Ser Ser Asp Tyr Arg Val	Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu		
805	810	815	820
Ser Gly Ser Val Val Leu Gly Cys Leu	Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln		
825	830	835	840
Pro Gln Lys Asn Val Val Ser His Arg	Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Pro		
845	850	855	860
Arg Ala Ser Ala Asn Leu Gly Gln Gly	Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn		
865	870		
Gly Arg Glu Val Val Asp Ser Thr Thr	Ser Ser Leu Ter		

Seq ID 20: S688L

5	10	15	20
Met Glu Ser Leu Leu Gly Phe Leu Ala	Leu Leu Leu Leu Trp Gly Ala Val Ala Glu Gly		
25	30	35	40
Pro Ala Lys Lys Val Leu Thr Leu Glu	Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val		
45	50	55	60
His Gln Lys Gly Gly Pro Ala Glu Glu	Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln		
65	70	75	80
Arg Leu Glu Ala Met Leu Phe Ala Leu	Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro		
85	90	95	100
Gly Val Arg Leu Gly Ala His Ile Leu	Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu		
105	110	115	120
Gln Ala Leu Asp Phe Val Arg Ala Ser	Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile		
125	130	135	140
Cys Pro Asp Gly Ser Tyr Ala Thr His	Ser Asp Ala Pro Thr Ala Val Thr Gly Val Ile		
145	150	155	160
Gly Gly Ser Tyr Ser Asp Val Ser Ile	Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile		

Pro Gln Ile Ser Tyr Ala Ser Thr Ser	165	170	175	180
Ala Lys Leu Ser Asp Lys Ser Arg Tyr Asp Tyr				
Phe Ala Arg Thr Val Pro Pro Asp Phe	185	190	195	200
Phe Gln Ala Lys Ala Met Ala Glu Ile Leu Arg				
Phe Phe Asn Trp Thr Tyr Val Ser Thr	205	210	215	220
Val Ala Ser Glu Gly Asp Tyr Gly Glu Thr Gly				
Ile Glu Ala Phe Glu Leu Glu Ala Arg	225	230	235	240
Ala Arg Asn Ile Cys Val Ala Thr Ser Glu Lys				
Val Gly Arg Ala Met Ser Arg Ala Ala	245	250	255	260
Phe Glu Gly Val Val Arg Ala Leu Leu Gln Lys				
Pro Ser Ala Arg Val Ala Val Leu Phe	265	270	275	280
Thr Arg Ser Glu Asp Ala Arg Glu Leu Leu Ala				
Ala Thr Gln Arg Leu Asn Ala Ser Phe	285	290	295	300
Thr Trp Val Ala Ser Asp Gly Trp Gly Ala Leu				
Glu Ser Val Val Ala Gly Ser Glu Arg	305	310	315	320
Ala Ala Glu Gly Ala Ile Thr Ile Glu Leu Ala				
Ser Tyr Pro Ile Ser Asp Phe Ala Ser	325	330	335	340
Tyr Phe Gln Ser Leu Asp Pro Trp Asn Asn Ser				
Arg Asn Pro Trp Phe Arg Glu Phe Trp	345	350	355	360
Glu Glu Arg Phe His Cys Ser Phe Arg Gln Arg				
Asp Cys Ala Ala His Ser Leu Arg Ala	365	370	375	380
Val Pro Phe Glu Gln Glu Ser Lys Ile Met Phe				
Val Val Asn Ala Val Tyr Ala Met Ala	385	390	395	400
His Ala Leu His Asn Met His Arg Ala Leu Cys				
Pro Asn Thr Thr His Leu Cys Asp Ala	405	410	415	420
Met Arg Pro Val Asn Gly Arg Arg Leu Tyr Lys				
Asp Phe Val Leu Asn Val Lys Phe Asp	425	430	435	440
Ala Pro Phe Arg Pro Ala Asp Thr Asp Asp Glu				
Val Arg Phe Asp Arg Phe Gly Asp Gly	445	450	455	460
Ile Gly Arg Tyr Asn Ile Phe Thr Tyr Leu Arg				
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln	465	470	475	480
Lys Val Gly Tyr Trp Ala Glu Gly Leu Thr Leu				
Asp Thr Ser Phe Ile Pro Trp Ala Ser	485	490	495	500
Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys				
Ser Glu Pro Cys Leu Gln Asn Glu Val	505	510	515	520
Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp				
Leu Cys Ile Pro Cys Gln Pro Tyr Glu	525	530	535	540
Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys				
Gly Leu Gly Tyr Trp Pro Asn Ala Ser	545	550	555	560
Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr				

565	570	575	580
Ile Arg Trp Gly Asp Ala Trp Ala Val Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu			
585	590	595	600
Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala			
605	610	615	620
Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr			
625	630	635	640
Phe Val Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly			
645	650	655	660
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile			
665	670	675	680
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val			
685	690	695	700
Ala Ile Cys Leu Ala Leu Ile Leu Gly Gln Leu Leu Ile Val Ala Ala Trp Leu Val Val			
705	710	715	720
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg			
725	730	735	740
Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asn Val Leu Leu Ile Ala			
745	750	755	760
Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys			
765	770	775	780
Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe			
785	790	795	800
Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu			
805	810	815	820
Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln			
825	830	835	840
Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Pro			
845	850	855	860
Arg Ala Ser Ala Asn Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn			
865	870		
Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter			

Seq ID 21: S688L, G689V

5	10	15	20
Met Glu Ser Leu Leu Gly Phe Leu Ala Leu Leu Leu Leu Trp Gly Ala Val Ala Glu Gly			
25	30	35	40
Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val			
45	50	55	60
His Gln Lys Gly Gly Pro Ala Glu Glu Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln			
65	70	75	80
Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro			

85	90	95	100
Gly Val Arg Leu Gly Ala His Ile Leu Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu			
105	110	115	120
Gln Ala Leu Asp Phe Val Arg Ala Ser Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile			
125	130	135	140
Cys Pro Asp Gly Ser Tyr Ala Thr His Ser Asp Ala Pro Thr Ala Val Thr Gly Val Ile			
145	150	155	160
Gly Gly Ser Tyr Ser Asp Val Ser Ile Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile			
165	170	175	180
Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp Lys Ser Arg Tyr Asp Tyr			
185	190	195	200
Phe Ala Arg Thr Val Pro Pro Asp Phe Phe Gln Ala Lys Ala Met Ala Glu Ile Leu Arg			
205	210	215	220
Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly Asp Tyr Gly Glu Thr Gly			
225	230	235	240
Ile Glu Ala Phe Glu Leu Glu Ala Arg Ala Arg Asn Ile Cys Val Ala Thr Ser Glu Lys			
245	250	255	260
Val Gly Arg Ala Met Ser Arg Ala Ala Phe Glu Gly Val Val Arg Ala Leu Leu Gln Lys			
265	270	275	280
Pro Ser Ala Arg Val Ala Val Leu Phe Thr Arg Ser Glu Asp Ala Arg Glu Leu Leu Ala			
285	290	295	300
Ala Thr Gln Arg Leu Asn Ala Ser Phe Thr Trp Val Ala Ser Asp Gly Trp Gly Ala Leu			
305	310	315	320
Glu Ser Val Val Ala Gly Ser Glu Arg Ala Ala Glu Gly Ala Ile Thr Ile Glu Leu Ala			
325	330	335	340
Ser Tyr Pro Ile Ser Asp Phe Ala Ser Tyr Phe Gln Ser Leu Asp Pro Trp Asn Asn Ser			
345	350	355	360
Arg Asn Pro Trp Phe Arg Glu Phe Trp Glu Glu Arg Phe His Cys Ser Phe Arg Gln Arg			
365	370	375	380
Asp Cys Ala Ala His Ser Leu Arg Ala Val Pro Phe Glu Gln Glu Ser Lys Ile Met Phe			
385	390	395	400
Val Val Asn Ala Val Tyr Ala Met Ala His Ala Leu His Asn Met His Arg Ala Leu Cys			
405	410	415	420
Pro Asn Thr Thr His Leu Cys Asp Ala Met Arg Pro Val Asn Gly Arg Arg Leu Tyr Lys			
425	430	435	440
Asp Phe Val Leu Asn Val Lys Phe Asp Ala Pro Phe Arg Pro Ala Asp Thr Asp Asp Glu			
445	450	455	460
Val Arg Phe Asp Arg Phe Gly Asp Gly Ile Gly Arg Tyr Asn Ile Phe Thr Tyr Leu Arg			
465	470	475	480
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln Lys Val Gly Tyr Trp Ala Glu Gly Leu Thr Leu			

485	490	495	500
Asp Thr Ser Phe Ile Pro Trp Ala Ser	Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys		
505	510	515	520
Ser Glu Pro Cys Leu Gln Asn Glu Val	Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp		
525	530	535	540
Leu Cys Ile Pro Cys Gln Pro Tyr Glu Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys			
545	550	555	560
Gly Leu Gly Tyr Trp Pro Asn Ala Ser	Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr		
565	570	575	580
Ile Arg Trp Gly Asp Ala Trp Ala Val	Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu		
585	590	595	600
Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala			
605	610	615	620
Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr			
625	630	635	640
Phe Val Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly			
645	650	655	660
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile			
665	670	675	680
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val			
685	690	695	700
Ala Ile Cys Leu Ala Leu Ile Leu Val Gln Leu Leu Ile Val Ala Ala Trp Leu Val Val			
705	710	715	720
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg			
725	730	735	740
Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asn Val Leu Leu Ile Ala			
745	750	755	760
Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys			
765	770	775	780
Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe			
785	790	795	800
Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu			
805	810	815	820
Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln			
825	830	835	840
Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Pro			
845	850	855	860
Arg Ala Ser Ala Asn Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn			
865	870		
Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter			

Seq ID 22: S688L, N735D

	5		10		15		20												
Met	Glu	Ser	Leu	Leu	Gly	Phe	Leu	Ala	Leu	Leu	Leu	Trp	Gly	Ala	Val	Ala	Glu	Gly	
	25		30		35		40												
Pro	Ala	Lys	Lys	Val	Leu	Thr	Leu	Glu	Gly	Asp	Leu	Val	Leu	Gly	Gly	Leu	Phe	Pro	Val
	45		50		55		60												
His	Gln	Lys	Gly	Gly	Pro	Ala	Glu	Glu	Cys	Gly	Pro	Val	Asn	Glu	His	Arg	Gly	Ile	Gln
	65		70		75		80												
Arg	Leu	Glu	Ala	Met	Leu	Phe	Ala	Leu	Asp	Arg	Ile	Asn	Arg	Asp	Pro	His	Leu	Leu	Pro
	85		90		95		100												
Gly	Val	Arg	Leu	Gly	Ala	His	Ile	Leu	Asp	Ser	Cys	Ser	Lys	Asp	Thr	His	Ala	Leu	Glu
	105		110		115		120												
Gln	Ala	Leu	Asp	Phe	Val	Arg	Ala	Ser	Leu	Ser	Arg	Gly	Ala	Asp	Gly	Ser	Arg	His	Ile
	125		130		135		140												
Cys	Pro	Asp	Gly	Ser	Tyr	Ala	Thr	His	Ser	Asp	Ala	Pro	Thr	Ala	Val	Thr	Gly	Val	Ile
	145		150		155		160												
Gly	Gly	Ser	Tyr	Ser	Asp	Val	Ser	Ile	Gln	Val	Ala	Asn	Leu	Leu	Arg	Leu	Phe	Gln	Ile
	165		170		175		180												
Pro	Gln	Ile	Ser	Tyr	Ala	Ser	Thr	Ser	Ala	Lys	Leu	Ser	Asp	Lys	Ser	Arg	Tyr	Asp	Tyr
	185		190		195		200												
Phe	Ala	Arg	Thr	Val	Pro	Pro	Asp	Phe	Phe	Gln	Ala	Lys	Ala	Met	Ala	Glu	Ile	Leu	Arg
	205		210		215		220												
Phe	Phe	Asn	Trp	Thr	Tyr	Val	Ser	Thr	Val	Ala	Ser	Glu	Gly	Asp	Tyr	Gly	Glu	Thr	Gly
	225		230		235		240												
Ile	Glu	Ala	Phe	Glu	Leu	Glu	Ala	Arg	Ala	Arg	Asn	Ile	Cys	Val	Ala	Thr	Ser	Glu	Lys
	245		250		255		260												
Val	Gly	Arg	Ala	Met	Ser	Arg	Ala	Ala	Phe	Glu	Gly	Val	Val	Arg	Ala	Leu	Leu	Gln	Lys
	265		270		275		280												
Pro	Ser	Ala	Arg	Val	Ala	Val	Leu	Phe	Thr	Arg	Ser	Glu	Asp	Ala	Arg	Glu	Leu	Leu	Ala
	285		290		295		300												
Ala	Thr	Gln	Arg	Leu	Asn	Ala	Ser	Phe	Thr	Trp	Val	Ala	Ser	Asp	Gly	Trp	Gly	Ala	Leu
	305		310		315		320												
Glu	Ser	Val	Val	Ala	Gly	Ser	Glu	Arg	Ala	Ala	Glu	Gly	Ala	Ile	Thr	Ile	Glu	Leu	Ala
	325		330		335		340												
Ser	Tyr	Pro	Ile	Ser	Asp	Phe	Ala	Ser	Tyr	Phe	Gln	Ser	Leu	Asp	Pro	Trp	Asn	Asn	Ser
	345		350		355		360												
Arg	Asn	Pro	Trp	Phe	Arg	Glu	Phe	Trp	Glu	Glu	Arg	Phe	His	Cys	Ser	Phe	Arg	Gln	Arg
	365		370		375		380												
Asp	Cys	Ala	Ala	His	Ser	Leu	Arg	Ala	Val	Pro	Phe	Glu	Gln	Glu	Ser	Lys	Ile	Met	Phe
	385		390		395		400												
Val	Val	Asn	Ala	Val	Tyr	Ala	Met	Ala	His	Ala	Leu	His	Asn	Met	His	Arg	Ala	Leu	Cys

Pro Asn Thr Thr His Leu Cys Asp Ala Met Arg Pro Val Asn Gly Arg Arg Leu Tyr Lys	405	410	415	420
Asp Phe Val Leu Asn Val Lys Phe Asp Ala Pro Phe Arg Pro Ala Asp Thr Asp Asp Glu	425	430	435	440
Val Arg Phe Asp Arg Phe Gly Asp Gly Ile Gly Arg Tyr Asn Ile Phe Thr Tyr Leu Arg	445	450	455	460
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln Lys Val Gly Tyr Trp Ala Glu Gly Leu Thr Leu	465	470	475	480
Asp Thr Ser Phe Ile Pro Trp Ala Ser Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys	485	490	495	500
Ser Glu Pro Cys Leu Gln Asn Glu Val Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp	505	510	515	520
Leu Cys Ile Pro Cys Gln Pro Tyr Glu Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys	525	530	535	540
Gly Leu Gly Tyr Trp Pro Asn Ala Ser Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr	545	550	555	560
Ile Arg Trp Gly Asp Ala Trp Ala Val Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu	565	570	575	580
Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala	585	590	595	600
Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr	605	610	615	620
Phe Val Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly	625	630	635	640
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile	645	650	655	660
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val	665	670	675	680
Ala Ile Cys Leu Ala Leu Ile Leu Gly Gln Leu Leu Ile Val Ala Ala Trp Leu Val Val	685	690	695	700
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg	705	710	715	720
Cys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asp Val Leu Leu Ile Ala	725	730	735	740
Leu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys	745	750	755	760
Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe	765	770	775	780
Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu	785	790	795	800
	805	810	815	820

Ser Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln
 825 830 835 840
 Pro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Pro
 845 850 855 860
 Arg Ala Ser Ala Asn Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn
 865 870
 Gly Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter

Seq ID 23: G689V

5 10 15 20
 Met Glu Ser Leu Leu Gly Phe Leu Ala Leu Leu Leu Leu Trp Gly Ala Val Ala Glu Gly
 25 30 35 40
 Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val
 45 50 55 60
 His Gln Lys Gly Gly Pro Ala Glu Glu Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln
 65 70 75 80
 Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro
 85 90 95 100
 Gly Val Arg Leu Gly Ala His Ile Leu Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu
 105 110 115 120
 Gln Ala Leu Asp Phe Val Arg Ala Ser Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile
 125 130 135 140
 Cys Pro Asp Gly Ser Tyr Ala Thr His Ser Asp Ala Pro Thr Ala Val Thr Gly Val Ile
 145 150 155 160
 Gly Gly Ser Tyr Ser Asp Val Ser Ile Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile
 165 170 175 180
 Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp Lys Ser Arg Tyr Asp Tyr
 185 190 195 200
 Phe Ala Arg Thr Val Pro Pro Asp Phe Phe Gln Ala Lys Ala Met Ala Glu Ile Leu Arg
 205 210 215 220
 Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly Asp Tyr Gly Glu Thr Gly
 225 230 235 240
 Ile Glu Ala Phe Glu Leu Glu Ala Arg Ala Arg Asn Ile Cys Val Ala Thr Ser Glu Lys
 245 250 255 260
 Val Gly Arg Ala Met Ser Arg Ala Ala Phe Glu Gly Val Val Arg Ala Leu Leu Gln Lys
 265 270 275 280
 Pro Ser Ala Arg Val Ala Val Leu Phe Thr Arg Ser Glu Asp Ala Arg Glu Leu Leu Ala
 285 290 295 300
 Ala Thr Gln Arg Leu Asn Ala Ser Phe Thr Trp Val Ala Ser Asp Gly Trp Gly Ala Leu
 305 310 315 320
 Glu Ser Val Val Ala Gly Ser Glu Arg Ala Ala Glu Gly Ala Ile Thr Ile Glu Leu Ala

325	330	335	340
Ser Tyr Pro Ile Ser Asp Phe Ala Ser Tyr Phe Gln Ser Leu Asp Pro Trp Asn Asn Ser			
345	350	355	360
Arg Asn Pro Trp Phe Arg Glu Phe Trp Glu Glu Arg Phe His Cys Ser Phe Arg Gln Arg			
365	370	375	380
Asp Cys Ala Ala His Ser Leu Arg Ala Val Pro Phe Glu Gln Glu Ser Lys Ile Met Phe			
385	390	395	400
Val Val Asn Ala Val Tyr Ala Met Ala His Ala Leu His Asn Met His Arg Ala Leu Cys			
405	410	415	420
Pro Asn Thr Thr His Leu Cys Asp Ala Met Arg Pro Val Asn Gly Arg Arg Leu Tyr Lys			
425	430	435	440
Asp Phe Val Leu Asn Val Lys Phe Asp Ala Pro Phe Arg Pro Ala Asp Thr Asp Asp Glu			
445	450	455	460
Val Arg Phe Asp Arg Phe Gly Asp Gly Ile Gly Arg Tyr Asn Ile Phe Thr Tyr Leu Arg			
465	470	475	480
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln Lys Val Gly Tyr Trp Ala Glu Gly Leu Thr Leu			
485	490	495	500
Asp Thr Ser Phe Ile Pro Trp Ala Ser Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys			
505	510	515	520
Ser Glu Pro Cys Leu Gln Asn Glu Val Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp			
525	530	535	540
Leu Cys Ile Pro Cys Gln Pro Tyr Glu Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys			
545	550	555	560
Gly Leu Gly Tyr Trp Pro Asn Ala Ser Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr			
565	570	575	580
Ile Arg Trp Gly Asp Ala Trp Ala Val Gly Pro Val Thr Ile Ala Cys Leu Gly Ala Leu			
585	590	595	600
Ala Thr Leu Phe Val Leu Gly Val Phe Val Arg His Asn Ala Thr Pro Val Val Lys Ala			
605	610	615	620
Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu Gly Gly Val Phe Leu Cys Tyr Cys Met Thr			
625	630	635	640
Phe Val Phe Ile Ala Lys Pro Ser Thr Ala Val Cys Thr Leu Arg Arg Leu Gly Leu Gly			
645	650	655	660
Thr Ala Phe Ser Val Cys Tyr Ser Ala Leu Leu Thr Lys Thr Asn Arg Ile Ala Arg Ile			
665	670	675	680
Phe Gly Gly Ala Arg Glu Gly Ala Gln Arg Pro Arg Phe Ile Ser Pro Ala Ser Gln Val			
685	690	695	700
Ala Ile Cys Leu Ala Leu Ile Ser Val Gln Leu Leu Ile Val Ala Ala Trp Leu Val Val			
705	710	715	720
Glu Ala Pro Gly Thr Gly Lys Glu Thr Ala Pro Glu Arg Arg Glu Val Val Thr Leu Arg			
725	730	735	740

ys Asn His Arg Asp Ala Ser Met Leu Gly Ser Leu Ala Tyr Asn Val Leu Leu Ile Ala
 745 750 755 760
 eu Cys Thr Leu Tyr Ala Phe Lys Thr Arg Lys Cys Pro Glu Asn Phe Asn Glu Ala Lys
 765 770 775 780
 he Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile Ile Trp Leu Ala Phe Leu Pro Ile Phe
 785 790 795 800
 yr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr Thr Thr Met Cys Val Ser Val Ser Leu
 805 810 815 820
 er Gly Ser Val Val Leu Gly Cys Leu Phe Ala Pro Lys Leu His Ile Ile Leu Phe Gln
 825 830 835 840
 ro Gln Lys Asn Val Val Ser His Arg Ala Pro Thr Ser Arg Phe Gly Ser Ala Ala Pro
 845 850 855 860
 rg Ala Ser Ala Asn Leu Gly Gln Gly Ser Gly Ser Gln Phe Val Pro Thr Val Cys Asn
 865 870
 ily Arg Glu Val Val Asp Ser Thr Thr Ser Ser Leu Ter

Seq ID 24: S688L, G689V, A733T, N735D

5 10 15 20
 Met Glu Ser Leu Leu Gly Phe Leu Ala Leu Leu Leu Leu Trp Gly Ala Val Ala Glu Gly
 25 30 35 40
 Pro Ala Lys Lys Val Leu Thr Leu Glu Gly Asp Leu Val Leu Gly Gly Leu Phe Pro Val
 45 50 55 60
 His Gln Lys Gly Gly Pro Ala Glu Glu Cys Gly Pro Val Asn Glu His Arg Gly Ile Gln
 65 70 75 80
 Arg Leu Glu Ala Met Leu Phe Ala Leu Asp Arg Ile Asn Arg Asp Pro His Leu Leu Pro
 85 90 95 100
 Gly Val Arg Leu Gly Ala His Ile Leu Asp Ser Cys Ser Lys Asp Thr His Ala Leu Glu
 105 110 115 120
 Gln Ala Leu Asp Phe Val Arg Ala Ser Leu Ser Arg Gly Ala Asp Gly Ser Arg His Ile
 125 130 135 140
 Cys Pro Asp Gly Ser Tyr Ala Thr His Ser Asp Ala Pro Thr Ala Val Thr Gly Val Ile
 145 150 155 160
 Gly Gly Ser Tyr Ser Asp Val Ser Ile Gln Val Ala Asn Leu Leu Arg Leu Phe Gln Ile
 165 170 175 180
 Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp Lys Ser Arg Tyr Asp Tyr
 185 190 195 200
 Phe Ala Arg Thr Val Pro Pro Asp Phe Phe Gln Ala Lys Ala Met Ala Glu Ile Leu Arg
 205 210 215 220
 Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly Asp Tyr Gly Glu Thr Gly
 225 230 235 240
 Ile Glu Ala Phe Glu Leu Glu Ala Arg Ala Arg Asn Ile Cys Val Ala Thr Ser Glu Lys

245	250	255	260
Val Gly Arg Ala Met Ser Arg Ala Ala Phe Glu Gly Val Val Arg Ala Leu Leu Gln Lys			
265	270	275	280
Pro Ser Ala Arg Val Ala Val Leu Phe Thr Arg Ser Glu Asp Ala Arg Glu Leu Leu Ala			
285	290	295	300
Ala Thr Gln Arg Leu Asn Ala Ser Phe Thr Trp Val Ala Ser Asp Gly Trp Gly Ala Leu			
305	310	315	320
Glu Ser Val Val Ala Gly Ser Glu Arg Ala Ala Glu Gly Ala Ile Thr Ile Glu Leu Ala			
325	330	335	340
Ser Tyr Pro Ile Ser Asp Phe Ala Ser Tyr Phe Gln Ser Leu Asp Pro Trp Asn Asn Ser			
345	350	355	360
Arg Asn Pro Trp Phe Arg Glu Phe Trp Glu Glu Arg Phe His Cys Ser Phe Arg Gln Arg			
365	370	375	380
Asp Cys Ala Ala His Ser Leu Arg Ala Val Pro Phe Glu Gln Glu Ser Lys Ile Met Phe			
385	390	395	400
Val Val Asn Ala Val Tyr Ala Met Ala His Ala Leu His Asn Met His Arg Ala Leu Cys			
405	410	415	420
Pro Asn Thr Thr His Leu Cys Asp Ala Met Arg Pro Val Asn Gly Arg Arg Leu Tyr Lys			
425	430	435	440
Asp Phe Val Leu Asn Val Lys Phe Asp Ala Pro Phe Arg Pro Ala Asp Thr Asp Asp Glu			
445	450	455	460
Val Arg Phe Asp Arg Phe Gly Asp Gly Ile Gly Arg Tyr Asn Ile Phe Thr Tyr Leu Arg			
465	470	475	480
Ala Gly Ser Gly Arg Tyr Arg Tyr Gln Lys Val Gly Tyr Trp Ala Glu Gly Leu Thr Leu			
485	490	495	500
Asp Thr Ser Phe Ile Pro Trp Ala Ser Pro Ser Ala Gly Pro Leu Pro Ala Ser Arg Cys			
505	510	515	520
Ser Glu Pro Cys Leu Gln Asn Glu Val Lys Ser Val Gln Pro Gly Glu Val Cys Cys Trp			
525	530	535	540
Leu Cys Ile Pro Cys Gln Pro Tyr Glu Tyr Arg Leu Asp Glu Phe Thr Cys Ala Asp Cys			
545	550	555	560
Gly Leu Gly Tyr Trp Pro Asn Ala Ser Leu Thr Gly Cys Phe Glu Leu Pro Gln Glu Tyr			
565	570	575	580
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Seq ID 30: S688L, N735D

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Seq ID 31: G698V

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Seq ID 32: S688L, G689V, A733T, N735D

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Human mGluR3 sequences

Seq ID 33: D744N

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Leu

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Val

 45 50 55

60
Leu Gly Gly Leu Phe Pro Ile Asn Glu Lys Gly Thr Gly Thr Glu Glu Cys Gly Arg
Ile

 65 70 75

80
Asn Glu Asp Arg Gly Ile Gln Arg Leu Glu Ala Met Leu Phe Ala Ile Asp Glu Ile
Asn

 85 90 95

100
Lys Asp Asp Tyr Leu Leu Pro Gly Val Lys Leu Gly Val His Ile Leu Asp Thr Cys
Ser

 105 110 115

120
Arg Asp Thr Tyr Ala Leu Glu Gln Ser Leu Glu Phe Val Arg Ala Ser Leu Thr Lys
Val

 125 130 135

140
Asp Glu Ala Glu Tyr Met Cys Pro Asp Gly Ser Tyr Ala Ile Gln Glu Asn Ile Pro
Leu

 145 150 155

160
Leu Ile Ala Gly Val Ile Gly Gly Ser Tyr Ser Ser Val Ser Ile Gln Val Ala Asn
Leu

 165 170 175

180
Leu Arg Leu Phe Gln Ile Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser
Asp

 185 190 195

200
Lys Ser Arg Tyr Asp Tyr Phe Ala Arg Thr Val Pro Pro Asp Phe Tyr Gln Ala Lys
Ala

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Gly

 225 230 235

240
Asp Tyr Gly Glu Thr Gly Ile Glu Ala Phe Glu Gln Glu Ala Arg Leu Arg Asn Ile
Cys

245 250 255
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Ile Ala Thr Ala Glu Lys Val Gly Arg Ser Asn Ile Arg Lys Ser Tyr Asp Ser Val
Ile

265 270 275
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Arg Glu Leu Leu Gln Lys Pro Asn Ala Arg Val Val Val Leu Phe Met Arg Ser Asp
Asp

285 290 295
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Ser Arg Glu Leu Ile Ala Ala Ala Ser Arg Ala Asn Ala Ser Phe Thr Trp Val Ala
Ser

305 310 315
320
Asp Gly Trp Gly Ala Gln Glu Ser Ile Ile Lys Gly Ser Glu His Val Ala Tyr Gly
Asp

325 330 335
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Ile Thr Leu Glu Leu Ala Ser Gln Pro Val Arg Gln Phe Gly Arg Tyr Phe Gln Ser
Leu

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Asn Pro Tyr Asn Asn His Arg Asn Pro Trp Phe Arg Asp Phe Trp Glu Gln Lys Phe
Gln

365 370 375
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Cys Ser Leu Gln Asn Lys Arg Asn His Arg Arg Val Cys Glu Lys His Leu Ala Ile
Asp

385 390 395
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Ser Ser Asn Tyr Glu Gln Glu Ser Lys Ile Met Phe Val Val Asn Ala Val Tyr Ala
Met

405 410 415
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Ala His Ala Leu His Lys Met Gln Arg Thr Leu Cys Pro Asn Thr Thr Lys Leu Cys
Asp

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Phe

445 450 455
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Thr Ala Pro Phe Asn Pro Asn Lys Asp Ala Asp Ser Ile Val Lys Phe Asp Thr Phe
Gly

465 470 475
480
Asp Gly Met Gly Arg Tyr Asn Val Phe Asn Phe Gln Asn Val Gly Gly Lys Tyr Ser
Tyr

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Leu Lys Val Gly His Trp Ala Glu Thr Leu Ser Leu Asp Val Asn Ser Ile His Trp
Ser

520 505 510 515
Arg Asn Ser Val Pro Thr Ser Gln Cys Ser Asp Pro Cys Ala Pro Asn Glu Met Lys
Asn

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Met Gln Pro Gly Asp Val Cys Cys Trp Ile Cys Ile Pro Cys Glu Pro Tyr Glu Tyr
Leu

560 545 550 555
Ala Asp Glu Phe Thr Cys Met Asp Cys Gly Ser Gly Gln Trp Pro Thr Ala Asp Leu
Thr

580 565 570 575
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Pro

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Val Thr Ile Ala Cys Leu Gly Phe Met Cys Thr Cys Met Val Val Thr Val Phe Ile
Lys

620 605 610 615
His Asn Asn Thr Pro Leu Val Lys Ala Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu
Phe

640 625 630 635
Gly Val Gly Leu Ser Tyr Cys Met Thr Phe Phe Phe Ile Ala Lys Pro Ser Pro Val
Ile

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Cys Ala Leu Arg Arg Leu Gly Leu Gly Ser Ser Phe Ala Ile Cys Tyr Ser Ala Leu
Leu

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Pro

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Lys Phe Ile Ser Pro Ser Ser Gln Val Phe Ile Cys Leu Gly Leu Ile Leu Val Gln
Ile

720 705 710 715
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Ala

725 730 735
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 760
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 Lys
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 780
 Cys Pro Glu Asn Phe Asn Glu Ala Lys Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys
 Ile
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 Ile Trp Leu Ala Phe Leu Pro Ile Phe Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln
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 Ala
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 Ter

Seq ID 34: V698G, D744N
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 Val
 45 50 55
 60
 Leu Gly Gly Leu Phe Pro Ile Asn Glu Lys Gly Thr Gly Thr Glu Glu Cys Gly Arg
 Ile
 65 70 75
 80
 Asn Glu Asp Arg Gly Ile Gln Arg Leu Glu Ala Met Leu Phe Ala Ile Asp Glu Ile
 Asn

100	85	90	95
Lys Asp Asp Tyr Leu Leu Pro Gly Val Lys Leu Gly Val His Ile Leu Asp Thr Cys Ser			
120	105	110	115
Arg Asp Thr Tyr Ala Leu Glu Gln Ser Leu Glu Phe Val Arg Ala Ser Leu Thr Lys Val			
140	125	130	135
Asp Glu Ala Glu Tyr Met Cys Pro Asp Gly Ser Tyr Ala Ile Gln Glu Asn Ile Pro Leu			
160	145	150	155
Leu Ile Ala Gly Val Ile Gly Gly Ser Tyr Ser Ser Val Ser Ile Gln Val Ala Asn Leu			
180	165	170	175
Leu Arg Leu Phe Gln Ile Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp			
200	185	190	195
Lys Ser Arg Tyr Asp Tyr Phe Ala Arg Thr Val Pro Pro Asp Phe Tyr Gln Ala Lys Ala			
220	205	210	215
Met Ala Glu Ile Leu Arg Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly			
240	225	230	235
Asp Tyr Gly Glu Thr Gly Ile Glu Ala Phe Glu Gln Glu Ala Arg Leu Arg Asn Ile Cys			
260	245	250	255
Ile Ala Thr Ala Glu Lys Val Gly Arg Ser Asn Ile Arg Lys Ser Tyr Asp Ser Val Ile			
280	265	270	275
Arg Glu Leu Leu Gln Lys Pro Asn Ala Arg Val Val Val Leu Phe Met Arg Ser Asp Asp			
300	285	290	295
Ser Arg Glu Leu Ile Ala Ala Ala Ser Arg Ala Asn Ala Ser Phe Thr Trp Val Ala Ser			
320	305	310	315
Asp Gly Trp Gly Ala Gln Glu Ser Ile Ile Lys Gly Ser Glu His Val Ala Tyr Gly Asp			

325 330 335
340
Ile Thr Leu Glu Leu Ala Ser Gln Pro Val Arg Gln Phe Gly Arg Tyr Phe Gln Ser
Leu

345 350 355
360
Asn Pro Tyr Asn Asn His Arg Asn Pro Trp Phe Arg Asp Phe Trp Glu Gln Lys Phe
Gln

365 370 375
380
Cys Ser Leu Gln Asn Lys Arg Asn His Arg Arg Val Cys Glu Lys His Leu Ala Ile
Asp

385 390 395
400
Ser Ser Asn Tyr Glu Gln Glu Ser Lys Ile Met Phe Val Val Asn Ala Val Tyr Ala
Met

405 410 415
420
Ala His Ala Leu His Lys Met Gln Arg Thr Leu Cys Pro Asn Thr Thr Lys Leu Cys
Asp

425 430 435
440
Ala Met Lys Ile Leu Asp Gly Lys Lys Leu Tyr Lys Asp Tyr Leu Leu Lys Ile Asn
Phe

445 450 455
460
Thr Ala Pro Phe Asn Pro Asn Lys Asp Ala Asp Ser Ile Val Lys Phe Asp Thr Phe
Gly

465 470 475
480
Asp Gly Met Gly Arg Tyr Asn Val Phe Asn Phe Gln Asn Val Gly Gly Lys Tyr Ser
Tyr

485 490 495
500
Leu Lys Val Gly His Trp Ala Glu Thr Leu Ser Leu Asp Val Asn Ser Ile His Trp
Ser

505 510 515
520
Arg Asn Ser Val Pro Thr Ser Gln Cys Ser Asp Pro Cys Ala Pro Asn Glu Met Lys
Asn

525 530 535
540
Met Gln Pro Gly Asp Val Cys Cys Trp Ile Cys Ile Pro Cys Glu Pro Tyr Glu Tyr
Leu

545 550 555
560
Ala Asp Glu Phe Thr Cys Met Asp Cys Gly Ser Gly Gln Trp Pro Thr Ala Asp Leu
Thr

	565	570	575
580			
Gly Cys Tyr Asp	Leu Pro Glu Asp Tyr Ile Arg Trp Glu Asp Ala Trp Ala Ile Gly		
Pro			
	585	590	595
600			
Val Thr Ile Ala Cys Leu Gly Phe Met Cys Thr Cys Met Val Val Thr Val Phe Ile			
Lys			
	605	610	615
620			
His Asn Asn Thr Pro Leu Val Lys Ala Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu			
Phe			
	625	630	635
640			
Gly Val Gly Leu Ser Tyr Cys Met Thr Phe Phe Phe Ile Ala Lys Pro Ser Pro Val			
Ile			
	645	650	655
660			
Cys Ala Leu Arg Arg Leu Gly Leu Gly Ser Ser Phe Ala Ile Cys Tyr Ser Ala Leu			
Leu			
	665	670	675
680			
Thr Lys Thr Asn Cys Ile Ala Arg Ile Phe Asp Gly Val Lys Asn Gly Ala Gln Arg			
Pro			
	685	690	695
700			
Lys Phe Ile Ser Pro Ser Ser Gln Val Phe Ile Cys Leu Gly Leu Ile Leu Gly Gln			
Ile			
	705	710	715
720			
Val Met Val Ser Val Trp Leu Ile Leu Glu Ala Pro Gly Thr Arg Arg Tyr Thr Leu			
Ala			
	725	730	735
740			
Glu Lys Arg Glu Thr Val Ile Leu Lys Cys Asn Val Lys Asp Ser Ser Met Leu Ile			
Ser			
	745	750	755
760			
Leu Thr Tyr Asn Val Ile Leu Val Ile Leu Cys Thr Val Tyr Ala Phe Lys Thr Arg			
Lys			
	765	770	775
780			
Cys Pro Glu Asn Phe Asn Glu Ala Lys Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys			
Ile			
	785	790	795
800			
Ile Trp Leu Ala Phe Leu Pro Ile Phe Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln			
Thr			

805 810 815
 820
 Thr Thr Met Cys Ile Ser Val Ser Leu Ser Gly Phe Val Val Leu Gly Cys Leu Phe
 Ala

825 830 835
 840
 Pro Lys Val His Ile Ile Leu Phe Gln Pro Gln Lys Asn Val Val Thr His Arg Leu
 His

845 850 855
 860
 Leu Asn Arg Phe Ser Val Ser Gly Thr Gly Thr Thr Tyr Ser Gln Ser Ser Ala Ser
 Thr

865 870 875
 880
 Tyr Val Pro Thr Val Cys Asn Gly Arg Glu Val Leu Asp Ser Thr Thr Ser Ser Leu
 Ter

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20

5 10 15
 Met Lys Met Leu Thr Arg Leu Gln Val Leu Thr Leu Ala Leu Phe Ser Lys Gly Phe
 Leu

25 30 35
 40
 Leu Ser Leu Gly Asp His Asn Phe Leu Arg Arg Glu Ile Lys Ile Glu Gly Asp Leu
 Val

45 50 55
 60
 Leu Gly Gly Leu Phe Pro Ile Asn Glu Lys Gly Thr Gly Thr Glu Glu Cys Gly Arg
 Ile

65 70 75
 80
 Asn Glu Asp Arg Gly Ile Gln Arg Leu Glu Ala Met Leu Phe Ala Ile Asp Glu Ile
 Asn

85 90 95
 100
 Lys Asp Asp Tyr Leu Leu Pro Gly Val Lys Leu Gly Val His Ile Leu Asp Thr Cys
 Ser

105 110 115
 120
 Arg Asp Thr Tyr Ala Leu Glu Gln Ser Leu Glu Phe Val Arg Ala Ser Leu Thr Lys
 Val

125 130 135
 140
 Asp Glu Ala Glu Tyr Met Cys Pro Asp Gly Ser Tyr Ala Ile Gln Glu Asn Ile Pro
 Leu

145 150 155
 160

Leu Ile Ala Gly Val Ile Gly Gly Ser Tyr Ser Ser Val Ser Ile Gln Val Ala Asn
Leu

165 170 175
180
Leu Arg Leu Phe Gln Ile Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser
Asp

185 190 195
200
Lys Ser Arg Tyr Asp Tyr Phe Ala Arg Thr Val Pro Pro Asp Phe Tyr Gln Ala Lys
Ala

205 210 215
220
Met Ala Glu Ile Leu Arg Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu
Gly

225 230 235
240
Asp Tyr Gly Glu Thr Gly Ile Glu Ala Phe Glu Gln Glu Ala Arg Leu Arg Asn Ile
Cys

245 250 255
260
Ile Ala Thr Ala Glu Lys Val Gly Arg Ser Asn Ile Arg Lys Ser Tyr Asp Ser Val
Ile

265 270 275
280
Arg Glu Leu Leu Gln Lys Pro Asn Ala Arg Val Val Val Leu Phe Met Arg Ser Asp
Asp

285 290 295
300
Ser Arg Glu Leu Ile Ala Ala Ala Ser Arg Ala Asn Ala Ser Phe Thr Trp Val Ala
Ser

305 310 315
320
Asp Gly Trp Gly Ala Gln Glu Ser Ile Ile Lys Gly Ser Glu His Val Ala Tyr Gly
Asp

325 330 335
340
Ile Thr Leu Glu Leu Ala Ser Gln Pro Val Arg Gln Phe Gly Arg Tyr Phe Gln Ser
Leu

345 350 355
360
Asn Pro Tyr Asn Asn His Arg Asn Pro Trp Phe Arg Asp Phe Trp Glu Gln Lys Phe
Gln

365 370 375
380
Cys Ser Leu Gln Asn Lys Arg Asn His Arg Arg Val Cys Glu Lys His Leu Ala Ile
Asp

385 390 395
400

Ser Ser Asn Tyr Glu Gln Glu Ser Lys Ile Met Phe Val Val Asn Ala Val Tyr Ala
Met

405

410

415

420

Ala His Ala Leu His Lys Met Gln Arg Thr Leu Cys Pro Asn Thr Thr Lys Leu Cys
Asp

425

430

435

440

Ala Met Lys Ile Leu Asp Gly Lys Lys Leu Tyr Lys Asp Tyr Leu Leu Lys Ile Asn
Phe

445

450

455

460

Thr Ala Pro Phe Asn Pro Asn Lys Asp Ala Asp Ser Ile Val Lys Phe Asp Thr Phe
Gly

465

470

475

480

Asp Gly Met Gly Arg Tyr Asn Val Phe Asn Phe Gln Asn Val Gly Gly Lys Tyr Ser
Tyr

485

490

495

500

Leu Lys Val Gly His Trp Ala Glu Thr Leu Ser Leu Asp Val Asn Ser Ile His Trp
Ser

505

510

515

520

Arg Asn Ser Val Pro Thr Ser Gln Cys Ser Asp Pro Cys Ala Pro Asn Glu Met Lys
Asn

525

530

535

540

Met Gln Pro Gly Asp Val Cys Cys Trp Ile Cys Ile Pro Cys Glu Pro Tyr Glu Tyr
Leu

545

550

555

560

Ala Asp Glu Phe Thr Cys Met Asp Cys Gly Ser Gly Gln Trp Pro Thr Ala Asp Leu
Thr

565

570

575

580

Gly Cys Tyr Asp Leu Pro Glu Asp Tyr Ile Arg Trp Glu Asp Ala Trp Ala Ile Gly
Pro

585

590

595

600

Val Thr Ile Ala Cys Leu Gly Phe Met Cys Thr Cys Met Val Val Thr Val Phe Ile
Lys

605

610

615

620

His Asn Asn Thr Pro Leu Val Lys Ala Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu
Phe

625

630

635

640

Gly Val Gly Leu Ser Tyr Cys Met Thr Phe Phe Phe Ile Ala Lys Pro Ser Pro Val
Ile

645 650 655
660
Cys Ala Leu Arg Arg Leu Gly Leu Gly Ser Ser Phe Ala Ile Cys Tyr Ser Ala Leu
Leu

665 670 675
680
Thr Lys Thr Asn Cys Ile Ala Arg Ile Phe Asp Gly Val Lys Asn Gly Ala Gln Arg
Pro

685 690 695
700
Lys Phe Ile Ser Pro Ser Ser Gln Val Phe Ile Cys Leu Gly Leu Ile Ser Gly Gln
Ile

705 710 715
720
Val Met Val Ser Val Trp Leu Ile Leu Glu Ala Pro Gly Thr Arg Arg Tyr Thr Leu
Ala

725 730 735
740
Glu Lys Arg Glu Thr Val Ile Leu Lys Cys Asn Val Lys Asp Ser Ser Met Leu Ile
Ser

745 750 755
760
Leu Thr Tyr Asn Val Ile Leu Val Ile Leu Cys Thr Val Tyr Ala Phe Lys Thr Arg
Lys

765 770 775
780
Cys Pro Glu Asn Phe Asn Glu Ala Lys Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys
Ile

785 790 795
800
Ile Trp Leu Ala Phe Leu Pro Ile Phe Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln
Thr

805 810 815
820
Thr Thr Met Cys Ile Ser Val Ser Leu Ser Gly Phe Val Val Leu Gly Cys Leu Phe
Ala

825 830 835
840
Pro Lys Val His Ile Ile Leu Phe Gln Pro Gln Lys Asn Val Val Thr His Arg Leu
His

845 850 855
860
Leu Asn Arg Phe Ser Val Ser Gly Thr Gly Thr Thr Tyr Ser Gln Ser Ser Ala Ser
Thr

865 870 875
880

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	5		10		15
Met Lys Met Leu Thr Arg Leu Gln Val Leu Thr Leu Ala Leu Phe Ser Lys Gly Phe Leu					
	25		30		35
40					
Leu Ser Leu Gly Asp His Asn Phe Leu Arg Arg Glu Ile Lys Ile Glu Gly Asp Leu Val					
	45		50		55
60					
Leu Gly Gly Leu Phe Pro Ile Asn Glu Lys Gly Thr Gly Thr Glu Glu Cys Gly Arg Ile					
	65		70		75
80					
Asn Glu Asp Arg Gly Ile Gln Arg Leu Glu Ala Met Leu Phe Ala Ile Asp Glu Ile Asn					
	85		90		95
100					
Lys Asp Asp Tyr Leu Leu Pro Gly Val Lys Leu Gly Val His Ile Leu Asp Thr Cys Ser					
	105		110		115
120					
Arg Asp Thr Tyr Ala Leu Glu Gln Ser Leu Glu Phe Val Arg Ala Ser Leu Thr Lys Val					
	125		130		135
140					
Asp Glu Ala Glu Tyr Met Cys Pro Asp Gly Ser Tyr Ala Ile Gln Glu Asn Ile Pro Leu					
	145		150		155
160					
Leu Ile Ala Gly Val Ile Gly Gly Ser Tyr Ser Ser Val Ser Ile Gln Val Ala Asn Leu					
	165		170		175
180					
Leu Arg Leu Phe Gln Ile Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser Asp					
	185		190		195
200					
Lys Ser Arg Tyr Asp Tyr Phe Ala Arg Thr Val Pro Pro Asp Phe Tyr Gln Ala Lys Ala					
	205		210		215
220					
Met Ala Glu Ile Leu Arg Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly					

225 230 235
240
Asp Tyr Gly Glu Thr Gly Ile Glu Ala Phe Glu Gln Glu Ala Arg Leu Arg Asn Ile
Cys

245 250 255
260
Ile Ala Thr Ala Glu Lys Val Gly Arg Ser Asn Ile Arg Lys Ser Tyr Asp Ser Val
Ile

265 270 275
280
Arg Glu Leu Leu Gln Lys Pro Asn Ala Arg Val Val Val Leu Phe Met Arg Ser Asp
Asp

285 290 295
300
Ser Arg Glu Leu Ile Ala Ala Ala Ser Arg Ala Asn Ala Ser Phe Thr Trp Val Ala
Ser

305 310 315
320
Asp Gly Trp Gly Ala Gln Glu Ser Ile Ile Lys Gly Ser Glu His Val Ala Tyr Gly
Asp

325 330 335
340
Ile Thr Leu Glu Leu Ala Ser Gln Pro Val Arg Gln Phe Gly Arg Tyr Phe Gln Ser
Leu

345 350 355
360
Asn Pro Tyr Asn Asn His Arg Asn Pro Trp Phe Arg Asp Phe Trp Glu Gln Lys Phe
Gln

365 370 375
380
Cys Ser Leu Gln Asn Lys Arg Asn His Arg Arg Val Cys Glu Lys His Leu Ala Ile
Asp

385 390 395
400
Ser Ser Asn Tyr Glu Gln Glu Ser Lys Ile Met Phe Val Val Asn Ala Val Tyr Ala
Met

405 410 415
420
Ala His Ala Leu His Lys Met Gln Arg Thr Leu Cys Pro Asn Thr Thr Lys Leu Cys
Asp

425 430 435
440
Ala Met Lys Ile Leu Asp Gly Lys Lys Leu Tyr Lys Asp Tyr Leu Leu Lys Ile Asn
Phe

445 450 455
460
Thr Ala Pro Phe Asn Pro Asn Lys Asp Ala Asp Ser Ile Val Lys Phe Asp Thr Phe
Gly

465 470 475
480
Asp Gly Met Gly Arg Tyr Asn Val Phe Asn Phe Gln Asn Val Gly Gly Lys Tyr Ser
Tyr

485 490 495
500
Leu Lys Val Gly His Trp Ala Glu Thr Leu Ser Leu Asp Val Asn Ser Ile His Trp
Ser

505 510 515
520
Arg Asn Ser Val Pro Thr Ser Gln Cys Ser Asp Pro Cys Ala Pro Asn Glu Met Lys
Asn

525 530 535
540
Met Gln Pro Gly Asp Val Cys Cys Trp Ile Cys Ile Pro Cys Glu Pro Tyr Glu Tyr
Leu

545 550 555
560
Ala Asp Glu Phe Thr Cys Met Asp Cys Gly Ser Gly Gln Trp Pro Thr Ala Asp Leu
Thr

565 570 575
580
Gly Cys Tyr Asp Leu Pro Glu Asp Tyr Ile Arg Trp Glu Asp Ala Trp Ala Ile Gly
Pro

585 590 595
600
Val Thr Ile Ala Cys Leu Gly Phe Met Cys Thr Cys Met Val Val Thr Val Phe Ile
Lys

605 610 615
620
His Asn Asn Thr Pro Leu Val Lys Ala Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu
Phe

625 630 635
640
Gly Val Gly Leu Ser Tyr Cys Met Thr Phe Phe Phe Ile Ala Lys Pro Ser Pro Val
Ile

645 650 655
660
Cys Ala Leu Arg Arg Leu Gly Leu Gly Ser Ser Phe Ala Ile Cys Tyr Ser Ala Leu
Leu

665 670 675
680
Thr Lys Thr Asn Cys Ile Ala Arg Ile Phe Asp Gly Val Lys Asn Gly Ala Gln Arg
Pro

685 690 695
700
Lys Phe Ile Ser Pro Ser Ser Gln Val Phe Ile Cys Leu Gly Leu Ile Ser Val Gln
Ile

705 710 715
 720
 Val Met Val Ser Val Trp Leu Ile Leu Glu Ala Pro Gly Thr Arg Arg Tyr Thr Leu
 Ala
 725 730 735
 740
 Glu Lys Arg Glu Thr Val Ile Leu Lys Cys Asn Val Lys Asp Ser Ser Met Leu Ile
 Ser
 745 750 755
 760
 Leu Thr Tyr Asp Val Ile Leu Val Ile Leu Cys Thr Val Tyr Ala Phe Lys Thr Arg
 Lys
 765 770 775
 780
 Cys Pro Glu Asn Phe Asn Glu Ala Lys Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys
 Ile
 785 790 795
 800
 Ile Trp Leu Ala Phe Leu Pro Ile Phe Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln
 Thr
 805 810 815
 820
 Thr Thr Met Cys Ile Ser Val Ser Leu Ser Gly Phe Val Val Leu Gly Cys Leu Phe
 Ala
 825 830 835
 840
 Pro Lys Val His Ile Ile Leu Phe Gln Pro Gln Lys Asn Val Val Thr His Arg Leu
 His
 845 850 855
 860
 Leu Asn Arg Phe Ser Val Ser Gly Thr Gly Thr Thr Tyr Ser Gln Ser Ser Ala Ser
 Thr
 865 870 875
 880
 Tyr Val Pro Thr Val Cys Asn Gly Arg Glu Val Leu Asp Ser Thr Thr Ser Ser Leu
 Ter

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20 5 10 15
 Met Lys Met Leu Thr Arg Leu Gln Val Leu Thr Leu Ala Leu Phe Ser Lys Gly Phe
 Leu
 25 30 35
 40
 Leu Ser Leu Gly Asp His Asn Phe Leu Arg Arg Glu Ile Lys Ile Glu Gly Asp Leu
 Val
 45 50 55
 60
 Leu Gly Gly Leu Phe Pro Ile Asn Glu Lys Gly Thr Gly Thr Glu Glu Cys Gly Arg
 Ile

80 65 70 75
Asn Glu Asp Arg Gly Ile Gln Arg Leu Glu Ala Met Leu Phe Ala Ile Asp Glu Ile
Asn

100 85 90 95
Lys Asp Asp Tyr Leu Leu Pro Gly Val Lys Leu Gly Val His Ile Leu Asp Thr Cys
Ser

120 105 110 115
Arg Asp Thr Tyr Ala Leu Glu Gln Ser Leu Glu Phe Val Arg Ala Ser Leu Thr Lys
Val

140 125 130 135
Asp Glu Ala Glu Tyr Met Cys Pro Asp Gly Ser Tyr Ala Ile Gln Glu Asn Ile Pro
Leu

160 145 150 155
Leu Ile Ala Gly Val Ile Gly Gly Ser Tyr Ser Ser Val Ser Ile Gln Val Ala Asn
Leu

180 165 170 175
Leu Arg Leu Phe Gln Ile Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser
Asp

200 185 190 195
Lys Ser Arg Tyr Asp Tyr Phe Ala Arg Thr Val Pro Pro Asp Phe Tyr Gln Ala Lys
Ala

220 205 210 215
Met Ala Glu Ile Leu Arg Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu
Gly

240 225 230 235
Asp Tyr Gly Glu Thr Gly Ile Glu Ala Phe Glu Gln Glu Ala Arg Leu Arg Asn Ile
Cys

260 245 250 255
Ile Ala Thr Ala Glu Lys Val Gly Arg Ser Asn Ile Arg Lys Ser Tyr Asp Ser Val
Ile

280 265 270 275
Arg Glu Leu Leu Gln Lys Pro Asn Ala Arg Val Val Val Leu Phe Met Arg Ser Asp
Asp

300 285 290 295
Ser Arg Glu Leu Ile Ala Ala Ala Ser Arg Ala Asn Ala Ser Phe Thr Trp Val Ala
Ser

305 310 315
320
Asp Gly Trp Gly Ala Gln Glu Ser Ile Ile Lys Gly Ser Glu His Val Ala Tyr Gly
Asp

325 330 335
340
Ile Thr Leu Glu Leu Ala Ser Gln Pro Val Arg Gln Phe Gly Arg Tyr Phe Gln Ser
Leu

345 350 355
360
Asn Pro Tyr Asn Asn His Arg Asn Pro Trp Phe Arg Asp Phe Trp Glu Gln Lys Phe
Gln

365 370 375
380
Cys Ser Leu Gln Asn Lys Arg Asn His Arg Arg Val Cys Glu Lys His Leu Ala Ile
Asp

385 390 395
400
Ser Ser Asn Tyr Glu Gln Glu Ser Lys Ile Met Phe Val Val Asn Ala Val Tyr Ala
Met

405 410 415
420
Ala His Ala Leu His Lys Met Gln Arg Thr Leu Cys Pro Asn Thr Thr Lys Leu Cys
Asp

425 430 435
440
Ala Met Lys Ile Leu Asp Gly Lys Lys Leu Tyr Lys Asp Tyr Leu Leu Lys Ile Asn
Phe

445 450 455
460
Thr Ala Pro Phe Asn Pro Asn Lys Asp Ala Asp Ser Ile Val Lys Phe Asp Thr Phe
Gly

465 470 475
480
Asp Gly Met Gly Arg Tyr Asn Val Phe Asn Phe Gln Asn Val Gly Gly Lys Tyr Ser
Tyr

485 490 495
500
Leu Lys Val Gly His Trp Ala Glu Thr Leu Ser Leu Asp Val Asn Ser Ile His Trp
Ser

505 510 515
520
Arg Asn Ser Val Pro Thr Ser Gln Cys Ser Asp Pro Cys Ala Pro Asn Glu Met Lys
Asn

525 530 535
540
Met Gln Pro Gly Asp Val Cys Cys Trp Ile Cys Ile Pro Cys Glu Pro Tyr Glu Tyr
Leu

545 550 555
560
Ala Asp Glu Phe Thr Cys Met Asp Cys Gly Ser Gly Gln Trp Pro Thr Ala Asp Leu
Thr

565 570 575
580
Gly Cys Tyr Asp Leu Pro Glu Asp Tyr Ile Arg Trp Glu Asp Ala Trp Ala Ile Gly
Pro

585 590 595
600
Val Thr Ile Ala Cys Leu Gly Phe Met Cys Thr Cys Met Val Val Thr Val Phe Ile
Lys

605 610 615
620
His Asn Asn Thr Pro Leu Val Lys Ala Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu
Phe

625 630 635
640
Gly Val Gly Leu Ser Tyr Cys Met Thr Phe Phe Phe Ile Ala Lys Pro Ser Pro Val
Ile

645 650 655
660
Cys Ala Leu Arg Arg Leu Gly Leu Gly Ser Ser Phe Ala Ile Cys Tyr Ser Ala Leu
Leu

665 670 675
680
Thr Lys Thr Asn Cys Ile Ala Arg Ile Phe Asp Gly Val Lys Asn Gly Ala Gln Arg
Pro

685 690 695
700
Lys Phe Ile Ser Pro Ser Ser Gln Val Phe Ile Cys Leu Gly Leu Ile Ser Gly Gln
Ile

705 710 715
720
Val Met Val Ser Val Trp Leu Ile Leu Glu Ala Pro Gly Thr Arg Arg Tyr Thr Leu
Ala

725 730 735
740
Glu Lys Arg Glu Thr Val Ile Leu Lys Cys Asn Val Lys Asp Ser Ser Met Leu Ile
Ser

745 750 755
760
Leu Thr Tyr Asp Val Ile Leu Val Ile Leu Cys Thr Val Tyr Ala Phe Lys Thr Arg
Lys

765 770 775
780
Cys Pro Glu Asn Phe Asn Glu Ala Lys Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys
Ile

785 790 795
 800
 Ile Trp Leu Ala Phe Leu Pro Ile Phe Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln
 Thr
 805 810 815
 820
 Thr Thr Met Cys Ile Ser Val Ser Leu Ser Gly Phe Val Val Leu Gly Cys Leu Phe
 Ala
 825 830 835
 840
 Pro Lys Val His Ile Ile Leu Phe Gln Pro Gln Lys Asn Val Val Thr His Arg Leu
 His
 845 850 855
 860
 Leu Asn Arg Phe Ser Val Ser Gly Thr Gly Thr Thr Tyr Ser Gln Ser Ser Ala Ser
 Thr
 865 870 875
 880
 Tyr Val Pro Thr Val Cys Asn Gly Arg Glu Val Leu Asp Ser Thr Thr Ser Ser Leu
 Ter

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 5 10 15
 20 Met Lys Met Leu Thr Arg Leu Gln Val Leu Thr Leu Ala Leu Phe Ser Lys Gly Phe
 Leu
 25 30 35
 40
 Leu Ser Leu Gly Asp His Asn Phe Leu Arg Arg Glu Ile Lys Ile Glu Gly Asp Leu
 Val
 45 50 55
 60
 Leu Gly Gly Leu Phe Pro Ile Asn Glu Lys Gly Thr Gly Thr Glu Glu Cys Gly Arg
 Ile
 65 70 75
 80
 Asn Glu Asp Arg Gly Ile Gln Arg Leu Glu Ala Met Leu Phe Ala Ile Asp Glu Ile
 Asn
 85 90 95
 100
 Lys Asp Asp Tyr Leu Leu Pro Gly Val Lys Leu Gly Val His Ile Leu Asp Thr Cys
 Ser
 105 110 115
 120
 Arg Asp Thr Tyr Ala Leu Glu Gln Ser Leu Glu Phe Val Arg Ala Ser Leu Thr Lys
 Val
 125 130 135
 140

Asp Glu Ala Glu Tyr Met Cys Pro Asp Gly Ser Tyr Ala Ile Gln Glu Asn Ile Pro
Leu

145 150 155
160
Leu Ile Ala Gly Val Ile Gly Gly Ser Tyr Ser Ser Val Ser Ile Gln Val Ala Asn
Leu

165 170 175
180
Leu Arg Leu Phe Gln Ile Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser
Asp

185 190 195
200
Lys Ser Arg Tyr Asp Tyr Phe Ala Arg Thr Val Pro Pro Asp Phe Tyr Gln Ala Lys
Ala

205 210 215
220
Met Ala Glu Ile Leu Arg Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu
Gly

225 230 235
240
Asp Tyr Gly Glu Thr Gly Ile Glu Ala Phe Glu Gln Glu Ala Arg Leu Arg Asn Ile
Cys

245 250 255
260
Ile Ala Thr Ala Glu Lys Val Gly Arg Ser Asn Ile Arg Lys Ser Tyr Asp Ser Val
Ile

265 270 275
280
Arg Glu Leu Leu Gln Lys Pro Asn Ala Arg Val Val Val Leu Phe Met Arg Ser Asp
Asp

285 290 295
300
Ser Arg Glu Leu Ile Ala Ala Ala Ser Arg Ala Asn Ala Ser Phe Thr Trp Val Ala
Ser

305 310 315
320
Asp Gly Trp Gly Ala Gln Glu Ser Ile Ile Lys Gly Ser Glu His Val Ala Tyr Gly
Asp

325 330 335
340
Ile Thr Leu Glu Leu Ala Ser Gln Pro Val Arg Gln Phe Gly Arg Tyr Phe Gln Ser
Leu

345 350 355
360
Asn Pro Tyr Asn Asn His Arg Asn Pro Trp Phe Arg Asp Phe Trp Glu Gln Lys Phe
Gln

365 370 375
380

Cys Ser Leu Gln Asn Lys Arg Asn His Arg Arg Val Cys Glu Lys His Leu Ala Ile
Asp

385 390 395
400

Ser Ser Asn Tyr Glu Gln Glu Ser Lys Ile Met Phe Val Val Asn Ala Val Tyr Ala
Met

405 410 415
420

Ala His Ala Leu His Lys Met Gln Arg Thr Leu Cys Pro Asn Thr Thr Lys Leu Cys
Asp

425 430 435
440

Ala Met Lys Ile Leu Asp Gly Lys Lys Leu Tyr Lys Asp Tyr Leu Leu Lys Ile Asn
Phe

445 450 455
460

Thr Ala Pro Phe Asn Pro Asn Lys Asp Ala Asp Ser Ile Val Lys Phe Asp Thr Phe
Gly

465 470 475

480
Asp Gly Met Gly Arg Tyr Asn Val Phe Asn Phe Gln Asn Val Gly Gly Lys Tyr Ser
Tyr

485 490 495
500

Leu Lys Val Gly His Trp Ala Glu Thr Leu Ser Leu Asp Val Asn Ser Ile His Trp
Ser

505 510 515
520

Arg Asn Ser Val Pro Thr Ser Gln Cys Ser Asp Pro Cys Ala Pro Asn Glu Met Lys
Asn

525 530 535
540

Met Gln Pro Gly Asp Val Cys Cys Trp Ile Cys Ile Pro Cys Glu Pro Tyr Glu Tyr
Leu

545 550 555
560

Ala Asp Glu Phe Thr Cys Met Asp Cys Gly Ser Gly Gln Trp Pro Thr Ala Asp Leu
Thr

565 570 575
580

Gly Cys Tyr Asp Leu Pro Glu Asp Tyr Ile Arg Trp Glu Asp Ala Trp Ala Ile Gly
Pro

585 590 595
600

Val Thr Ile Ala Cys Leu Gly Phe Met Cys Thr Cys Met Val Val Thr Val Phe Ile
Lys

605 610 615
620

His Asn Asn Thr Pro Leu Val Lys Ala Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu
Phe

625 630 635
640
Gly Val Gly Leu Ser Tyr Cys Met Thr Phe Phe Phe Ile Ala Lys Pro Ser Pro Val
Ile

645 650 655
660
Cys Ala Leu Arg Arg Leu Gly Leu Gly Ser Ser Phe Ala Ile Cys Tyr Ser Ala Leu
Leu

665 670 675
680
Thr Lys Thr Asn Cys Ile Ala Arg Ile Phe Asp Gly Val Lys Asn Gly Ala Gln Arg
Pro

685 690 695
700
Lys Phe Ile Ser Pro Ser Ser Gln Val Phe Ile Cys Leu Gly Leu Ile Ser Val Gln
Ile

705 710 715
720
Val Met Val Ser Val Trp Leu Ile Leu Glu Ala Pro Gly Thr Arg Arg Tyr Thr Leu
Ala

725 730 735
740
Glu Lys Arg Glu Thr Val Ile Leu Lys Cys Asn Val Lys Asp Ser Ser Met Leu Ile
Ser

745 750 755
760
Leu Thr Tyr Asn Val Ile Leu Val Ile Leu Cys Thr Val Tyr Ala Phe Lys Thr Arg
Lys

765 770 775
780
Cys Pro Glu Asn Phe Asn Glu Ala Lys Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys
Ile

785 790 795
800
Ile Trp Leu Ala Phe Leu Pro Ile Phe Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln
Thr

805 810 815
820
Thr Thr Met Cys Ile Ser Val Ser Leu Ser Gly Phe Val Val Leu Gly Cys Leu Phe
Ala

825 830 835
840
Pro Lys Val His Ile Ile Leu Phe Gln Pro Gln Lys Asn Val Val Thr His Arg Leu
His

845 850 855
860

Leu Asn Arg Phe Ser Val Ser Gly Thr Gly Thr Thr Tyr Ser Gln Ser Ser Ala Ser
Thr

865

870

875

880

Tyr Val Pro Thr Val Cys Asn Gly Arg Glu Val Leu Asp Ser Thr Thr Ser Ser Leu
Ter

Seq ID 39: V698G

5

10

15

20

Met Lys Met Leu Thr Arg Leu Gln Val Leu Thr Leu Ala Leu Phe Ser Lys Gly Phe
Leu

25

30

35

40

Leu Ser Leu Gly Asp His Asn Phe Leu Arg Arg Glu Ile Lys Ile Glu Gly Asp Leu
Val

45

50

55

60

Leu Gly Gly Leu Phe Pro Ile Asn Glu Lys Gly Thr Gly Thr Glu Glu Cys Gly Arg
Ile

65

70

75

80

Asn Glu Asp Arg Gly Ile Gln Arg Leu Glu Ala Met Leu Phe Ala Ile Asp Glu Ile
Asn

85

90

95

100

Lys Asp Asp Tyr Leu Leu Pro Gly Val Lys Leu Gly Val His Ile Leu Asp Thr Cys
Ser

105

110

115

120

Arg Asp Thr Tyr Ala Leu Glu Gln Ser Leu Glu Phe Val Arg Ala Ser Leu Thr Lys
Val

125

130

135

140

Asp Glu Ala Glu Tyr Met Cys Pro Asp Gly Ser Tyr Ala Ile Gln Glu Asn Ile Pro
Leu

145

150

155

160

Leu Ile Ala Gly Val Ile Gly Gly Ser Tyr Ser Ser Val Ser Ile Gln Val Ala Asn
Leu

165

170

175

180

Leu Arg Leu Phe Gln Ile Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser
Asp

185

190

195

200

Lys Ser Arg Tyr Asp Tyr Phe Ala Arg Thr Val Pro Pro Asp Phe Tyr Gln Ala Lys
Ala

220	205	210	215
Met Ala Glu Ile Leu Arg Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu Gly			
240	225	230	235
Asp Tyr Gly Glu Thr Gly Ile Glu Ala Phe Glu Gln Glu Ala Arg Leu Arg Asn Ile Cys			
260	245	250	255
Ile Ala Thr Ala Glu Lys Val Gly Arg Ser Asn Ile Arg Lys Ser Tyr Asp Ser Val Ile			
280	265	270	275
Arg Glu Leu Leu Gln Lys Pro Asn Ala Arg Val Val Val Leu Phe Met Arg Ser Asp Asp			
300	285	290	295
Ser Arg Glu Leu Ile Ala Ala Ala Ser Arg Ala Asn Ala Ser Phe Thr Trp Val Ala Ser			
320	305	310	315
Asp Gly Trp Gly Ala Gln Glu Ser Ile Ile Lys Gly Ser Glu His Val Ala Tyr Gly Asp			
340	325	330	335
Ile Thr Leu Glu Leu Ala Ser Gln Pro Val Arg Gln Phe Gly Arg Tyr Phe Gln Ser Leu			
360	345	350	355
Asn Pro Tyr Asn Asn His Arg Asn Pro Trp Phe Arg Asp Phe Trp Glu Gln Lys Phe Gln			
380	365	370	375
Cys Ser Leu Gln Asn Lys Arg Asn His Arg Arg Val Cys Glu Lys His Leu Ala Ile Asp			
400	385	390	395
Ser Ser Asn Tyr Glu Gln Glu Ser Lys Ile Met Phe Val Val Asn Ala Val Tyr Ala Met			
420	405	410	415
Ala His Ala Leu His Lys Met Gln Arg Thr Leu Cys Pro Asn Thr Thr Lys Leu Cys Asp			
440	425	430	435
Ala Met Lys Ile Leu Asp Gly Lys Lys Leu Tyr Lys Asp Tyr Leu Leu Lys Ile Asn Phe			

445 450 455
460
Thr Ala Pro Phe Asn Pro Asn Lys Asp Ala Asp Ser Ile Val Lys Phe Asp Thr Phe
Gly

465 470 475
480
Asp Gly Met Gly Arg Tyr Asn Val Phe Asn Phe Gln Asn Val Gly Gly Lys Tyr Ser
Tyr

485 490 495
500
Leu Lys Val Gly His Trp Ala Glu Thr Leu Ser Leu Asp Val Asn Ser Ile His Trp
Ser

505 510 515
520
Arg Asn Ser Val Pro Thr Ser Gln Cys Ser Asp Pro Cys Ala Pro Asn Glu Met Lys
Asn

525 530 535
540
Met Gln Pro Gly Asp Val Cys Cys Trp Ile Cys Ile Pro Cys Glu Pro Tyr Glu Tyr
Leu

545 550 555
560
Ala Asp Glu Phe Thr Cys Met Asp Cys Gly Ser Gly Gln Trp Pro Thr Ala Asp Leu
Thr

565 570 575
580
Gly Cys Tyr Asp Leu Pro Glu Asp Tyr Ile Arg Trp Glu Asp Ala Trp Ala Ile Gly
Pro

585 590 595
600
Val Thr Ile Ala Cys Leu Gly Phe Met Cys Thr Cys Met Val Val Thr Val Phe Ile
Lys

605 610 615
620
His Asn Asn Thr Pro Leu Val Lys Ala Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu
Phe

625 630 635
640
Gly Val Gly Leu Ser Tyr Cys Met Thr Phe Phe Phe Ile Ala Lys Pro Ser Pro Val
Ile

645 650 655
660
Cys Ala Leu Arg Arg Leu Gly Leu Gly Ser Ser Phe Ala Ile Cys Tyr Ser Ala Leu
Leu

665 670 675
680
Thr Lys Thr Asn Cys Ile Ala Arg Ile Phe Asp Gly Val Lys Asn Gly Ala Gln Arg
Pro

685 690 695
 700
 Lys Phe Ile Ser Pro Ser Ser Gln Val Phe Ile Cys Leu Gly Leu Ile Leu Gly Gln
 Ile
 705 710 715
 720
 Val Met Val Ser Val Trp Leu Ile Leu Glu Ala Pro Gly Thr Arg Arg Tyr Thr Leu
 Ala
 725 730 735
 740
 Glu Lys Arg Glu Thr Val Ile Leu Lys Cys Asn Val Lys Asp Ser Ser Met Leu Ile
 Ser
 745 750 755
 760
 Leu Thr Tyr Asp Val Ile Leu Val Ile Leu Cys Thr Val Tyr Ala Phe Lys Thr Arg
 Lys
 765 770 775
 780
 Cys Pro Glu Asn Phe Asn Glu Ala Lys Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys
 Ile
 785 790 795
 800
 Ile Trp Leu Ala Phe Leu Pro Ile Phe Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln
 Thr
 805 810 815
 820
 Thr Thr Met Cys Ile Ser Val Ser Leu Ser Gly Phe Val Val Leu Gly Cys Leu Phe
 Ala
 825 830 835
 840
 Pro Lys Val His Ile Ile Leu Phe Gln Pro Gln Lys Asn Val Val Thr His Arg Leu
 His
 845 850 855
 860
 Leu Asn Arg Phe Ser Val Ser Gly Thr Gly Thr Thr Tyr Ser Gln Ser Ser Ala Ser
 Thr
 865 870 875
 880
 Tyr Val Pro Thr Val Cys Asn Gly Arg Glu Val Leu Asp Ser Thr Thr Ser Ser Leu
 Ter

Seq ID 40: L697S, V698G, T742A, D744N

5

10

15

20

Met Lys Met Leu Thr Arg Leu Gln Val Leu Thr Leu Ala Leu Phe Ser Lys Gly Phe
Leu

25

30

35

40

Leu Ser Leu Gly Asp His Asn Phe Leu Arg Arg Glu Ile Lys Ile Glu Gly Asp Leu
Val

45 50 55
60
Leu Gly Gly Leu Phe Pro Ile Asn Glu Lys Gly Thr Gly Thr Glu Glu Cys Gly Arg
Ile

65 70 75
80
Asn Glu Asp Arg Gly Ile Gln Arg Leu Glu Ala Met Leu Phe Ala Ile Asp Glu Ile
Asn

85 90 95
100
Lys Asp Asp Tyr Leu Leu Pro Gly Val Lys Leu Gly Val His Ile Leu Asp Thr Cys
Ser

105 110 115
120
Arg Asp Thr Tyr Ala Leu Glu Gln Ser Leu Glu Phe Val Arg Ala Ser Leu Thr Lys
Val

125 130 135
140
Asp Glu Ala Glu Tyr Met Cys Pro Asp Gly Ser Tyr Ala Ile Gln Glu Asn Ile Pro
Leu

145 150 155
160
Leu Ile Ala Gly Val Ile Gly Gly Ser Tyr Ser Ser Val Ser Ile Gln Val Ala Asn
Leu

165 170 175
180
Leu Arg Leu Phe Gln Ile Pro Gln Ile Ser Tyr Ala Ser Thr Ser Ala Lys Leu Ser
Asp

185 190 195
200
Lys Ser Arg Tyr Asp Tyr Phe Ala Arg Thr Val Pro Pro Asp Phe Tyr Gln Ala Lys
Ala

205 210 215
220
Met Ala Glu Ile Leu Arg Phe Phe Asn Trp Thr Tyr Val Ser Thr Val Ala Ser Glu
Gly

225 230 235
240
Asp Tyr Gly Glu Thr Gly Ile Glu Ala Phe Glu Gln Glu Ala Arg Leu Arg Asn Ile
Cys

245 250 255
260
Ile Ala Thr Ala Glu Lys Val Gly Arg Ser Asn Ile Arg Lys Ser Tyr Asp Ser Val
Ile

265 270 275
280
Arg Glu Leu Leu Gln Lys Pro Asn Ala Arg Val Val Val Leu Phe Met Arg Ser Asp
Asp

285 290 295
300
Ser Arg Glu Leu Ile Ala Ala Ala Ser Arg Ala Asn Ala Ser Phe Thr Trp Val Ala
Ser

305 310 315
320
Asp Gly Trp Gly Ala Gln Glu Ser Ile Ile Lys Gly Ser Glu His Val Ala Tyr Gly
Asp

325 330 335
340
Ile Thr Leu Glu Leu Ala Ser Gln Pro Val Arg Gln Phe Gly Arg Tyr Phe Gln Ser
Leu

345 350 355
360
Asn Pro Tyr Asn Asn His Arg Asn Pro Trp Phe Arg Asp Phe Trp Glu Gln Lys Phe
Gln

365 370 375
380
Cys Ser Leu Gln Asn Lys Arg Asn His Arg Arg Val Cys Glu Lys His Leu Ala Ile
Asp

385 390 395
400
Ser Ser Asn Tyr Glu Gln Glu Ser Lys Ile Met Phe Val Val Asn Ala Val Tyr Ala
Met

405 410 415
420
Ala His Ala Leu His Lys Met Gln Arg Thr Leu Cys Pro Asn Thr Thr Lys Leu Cys
Asp

425 430 435
440
Ala Met Lys Ile Leu Asp Gly Lys Lys Leu Tyr Lys Asp Tyr Leu Leu Lys Ile Asn
Phe

445 450 455
460
Thr Ala Pro Phe Asn Pro Asn Lys Asp Ala Asp Ser Ile Val Lys Phe Asp Thr Phe
Gly

465 470 475
480
Asp Gly Met Gly Arg Tyr Asn Val Phe Asn Phe Gln Asn Val Gly Gly Lys Tyr Ser
Tyr

485 490 495
500
Leu Lys Val Gly His Trp Ala Glu Thr Leu Ser Leu Asp Val Asn Ser Ile His Trp
Ser

505 510 515
520
Arg Asn Ser Val Pro Thr Ser Gln Cys Ser Asp Pro Cys Ala Pro Asn Glu Met Lys
Asn

525 530 535
540
Met Gln Pro Gly Asp Val Cys Cys Trp Ile Cys Ile Pro Cys Glu Pro Tyr Glu Tyr
Leu

545 550 555
560
Ala Asp Glu Phe Thr Cys Met Asp Cys Gly Ser Gly Gln Trp Pro Thr Ala Asp Leu
Thr

565 570 575
580
Gly Cys Tyr Asp Leu Pro Glu Asp Tyr Ile Arg Trp Glu Asp Ala Trp Ala Ile Gly
Pro

585 590 595
600
Val Thr Ile Ala Cys Leu Gly Phe Met Cys Thr Cys Met Val Val Thr Val Phe Ile
Lys

605 610 615
620
His Asn Asn Thr Pro Leu Val Lys Ala Ser Gly Arg Glu Leu Cys Tyr Ile Leu Leu
Phe

625 630 635
640
Gly Val Gly Leu Ser Tyr Cys Met Thr Phe Phe Phe Ile Ala Lys Pro Ser Pro Val
Ile

645 650 655
660
Cys Ala Leu Arg Arg Leu Gly Leu Gly Ser Ser Phe Ala Ile Cys Tyr Ser Ala Leu
Leu

665 670 675
680
Thr Lys Thr Asn Cys Ile Ala Arg Ile Phe Asp Gly Val Lys Asn Gly Ala Gln Arg
Pro

685 690 695
700
Lys Phe Ile Ser Pro Ser Ser Gln Val Phe Ile Cys Leu Gly Leu Ile Ser Gly Gln
Ile

705 710 715
720
Val Met Val Ser Val Trp Leu Ile Leu Glu Ala Pro Gly Thr Arg Arg Tyr Thr Leu
Ala

725 730 735
740
Glu Lys Arg Glu Thr Val Ile Leu Lys Cys Asn Val Lys Asp Ser Ser Met Leu Ile
Ser

745 750 755
760
Leu Ala Tyr Asn Val Ile Leu Val Ile Leu Cys Thr Val Tyr Ala Phe Lys Thr Arg
Lys

	765	770	775
780			
Cys Pro Glu Asn Phe Asn Glu Ala Lys Phe Ile Gly Phe Thr Met Tyr Thr Thr Cys Ile			
	785	790	795
800			
Ile Trp Leu Ala Phe Leu Pro Ile Phe Tyr Val Thr Ser Ser Asp Tyr Arg Val Gln Thr			
	805	810	815
820			
Thr Thr Met Cys Ile Ser Val Ser Leu Ser Gly Phe Val Val Leu Gly Cys Leu Phe Ala			
	825	830	835
840			
Pro Lys Val His Ile Ile Leu Phe Gln Pro Gln Lys Asn Val Val Thr His Arg Leu His			
	845	850	855
860			
Leu Asn Arg Phe Ser Val Ser Gly Thr Gly Thr Thr Tyr Ser Gln Ser Ser Ala Ser Thr			
	865	870	875
880			
Tyr Val Pro Thr Val Cys Asn Gly Arg Glu Val Leu Asp Ser Thr Thr Ser Ser Leu Ter			

Seq ID 41: D744N

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1 atgaagatgt tgacaagact gcaagttctt accttagctt tgttttcaaa
51 gggattttta ctctcttttag gggaccataa ctttctaagg agagagatta
101 aaatagaagg tgaccttggt ttagggggcc tgtttcctat taacgaaaaa
151 ggcactggaa ctgaagaatg tgggcgaatc aatgaagacc gagggattca
201 acgcctggaa gccatgttgt ttgctattga tgaaatcaac aaagatgatt
251 acttgctacc aggagtgaag ttgggtgttc acattttgga tacatgttca
301 agggatacct atgcattgga gcaatcactg gagtttgtca gggcatcttt
351 gacaaaagtg gatgaagctg agtatatgtg tcctgatgga tcctatgcca
501 ttcaagaaaa catcccactt ctcatcgag gggtcattgg tggctcttat
551 agcagtgttt ccatacaggt ggcaaacctg ctgcggtctt tccagatccc
501 tcagatcagc tacgcatcca ccagcgccaa actcagtgat aagtcgcgct
551 atgattactt tgccaggacc gtgccccccg acttctacca ggccaaagcc
601 atggctgaga tcttgcgctt cttcaactgg acctacgtgt ccacagttagc
651 ctccgagggg gattacgggg agacagggat cgaggccttc gagcaggaag
701 cccgcctgcg caacatctgc atcgctacgg cggagaaggt gggccgctcc
751 aacatccgca agtcctacga cagcgtgatc cgagaactgt tgcagaagcc
801 caacgcgcgc gtcgtggtcc tcttcatgcg cagcgacgac tcgcgggagc
851 tcattgcagc cgccagccgc gccaatgcct ccttcacctg ggtggccagc
901 gacggttggg gcgcgcagga gagcatcatc aagggcagcg agcatgtggc
951 ctacggcgac atcacctgg agctggcctc ccagcctgtc cgccagttcg
1001 gccgtactt ccagagcctc aaccctaca acaaccaccg caaccctgg
1051 ttccgggact tctgggagca aaagtctcag tgcagcctcc agaacaacg
1101 caaccacagg cgcgtctgcg aaaagcacct ggccatcgac agcagcaact
1151 acgagcaaga gtccaagatc atgtttgtgg tgaacgcggt gtatgccatg
1201 gccacgctt tgcacaaaat gcagcgacc ctctgtccca acactaccaa
1251 gctttgtgat gctatgaaga tcctggatgg gaagaagttg tacaaggatt
1301 acttgctgaa aatcaacttc acggctccat tcaacccaaa taaagatgca
1351 gatagcatag tcaagtttga cacttttggg gatggaatgg ggcgatacaa
1501 cgtgttcaat ttccaaaatg taggtgggaa gtatttctac ttgaaagttg
1551 gtcactgggc agaaacctta tcgctagatg tcaactctat ccactggtcc
1501 cggaactcag tccccacttc ccagtgcagc gaccctgtg cccccaatga
1551 aatgaagaat atgcaaccag gggatgtctg ctgctggatt tgcatcccct
1601 gtgaacccta cgaatacctg gctgatgagt ttacctgtat ggattgtggg
1651 tctggacagt ggcccactgc agacctaaact ggatgctatg accttctga
1701 ggactacatc aggtgggaag acgcctgggc cattggccca gtcaccattg
1751 cctgtctggg ttttatgtgt acatgcatgg ttgtaactgt ttttatcaag
1801 cacaacaaca cacccttggg caaagcatcg ggccgagaaac tctgctacat
1851 cttattgttt ggggttggcc tgtcatactg catgacattc ttcttcattg
1901 ccaagccatc accagtcatc tgtgcattgc gccgactcgg gctggggagt
1951 tccttcgcta tctgttactc agccctgctg accaagacaa actgcattgc
2001 ccgcatcttc gatggggtca agaattggcg tcagaggcca aaattcatca
2051 gccccagttc tcaggttttc atctgcctgg gtctgatcct ggtgcaaatt
2101 gtgatgggtg ctgtgtgggt catcctggag gccccaggca ccaggaggta
2151 tacccttgca gagaagcggg aaacagtcac cctaaaatgc aatgtcaaag
2201 attccagcat gttgatctct cttacctaca atgtgatcct ggtgatctta
2251 tgcactgtgt acgccttcaa aacgcggaag tgcccagaaa atttcaacga
2301 agctaagttc ataggtttta ccatgtacac cacgtgcac cactggttgg
2351 ccttctctcc tatattttat gtgacatcaa gtgactacag agtgacagc
2501 acaaccatgt gcatctctgt cagcctgagt ggctttgtgg tcttgggctg
2551 tttgtttgca cccaaggttc acatcatcct gtttcaaccc cagaagaatg
2501 ttgtcacaca cagactgcac ctcaacaggt tcagtgtcag tggaactggg
2551 accacatact ctcatgcctc tgcaagcacg tatgtgcaa cgggtgtgcaa
2601 tgggcgggaa gtcctcgact ccaccacctc atctctgtga
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Seq ID 42: V698G, D744N

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1  atgaagatgt  tgacaagact  gcaagttctt  accttagctt  tgttttcaaa
51  gggattttta  ctctcttttag  gggaccataa  ctttctaagg  agagagatta
101  aaatagaagg  tgaccttggt  ttagggggcc  tgtttcctat  taacgaaaaa
151  ggcactggaa  ctgaagaatg  tgggcgaatc  aatgaagacc  gagggattca
201  acgcctggaa  gccatgttgt  ttgctattga  tgaaatcaac  aaagatgatt
251  acttgctacc  aggagtgaag  ttgggtgttc  acattttgga  tacatgttca
301  agggatacct  atgcattgga  gcaatcactg  gagtttgtca  gggcatcttt
351  gacaaaagtg  gatgaagctg  agtatatgtg  tcctgatgga  tcctatgcca
501  ttcaagaaaa  catcccactt  ctcatcgag  gggtcattgg  tggctcttat
551  agcagtgttt  ccatacaggt  ggcaaacctg  ctgcggctct  tccagatccc
501  tcagatcagc  tacgcatcca  ccagcgccaa  actcagtgat  aagtcgcgct
551  atgattactt  tgccaggacc  gtgccccccg  acttctacca  ggccaaagcc
601  atggctgaga  tcttgcgctt  cttcaactgg  acctacgtgt  ccacagtagc
651  ctccgagggg  gattacgggg  agacagggat  cgaggccttc  gagcaggaag
701  cccgcctgcg  caacatctgc  atcgctacgg  cggagaaggt  gggccgctcc
751  aacatccgca  agtcctacga  cagcgtgatc  cgagaactgt  tgcagaagcc
801  caacgcgcgc  gtcgtggtcc  tcttcatgcg  cagcgacgac  tcgcgggagc
851  tcattgcagc  cgccagccgc  gccaatgcct  ccttcacctg  ggtggccagc
901  gacggttggg  gcgcgcagga  gagcatcatc  aagggcagcg  agcatgtggc
951  ctacggcgag  ataccctgg  agctggcctc  ccagcctgtc  cgccagttcg
1001  gccgtacttt  ccagagcctc  aacccttaca  acaaccaccg  caacccttgg
1051  ttccgggact  tctgggagca  aaagtttcag  tgcagcctcc  agaacaaacg
1101  caaccacagg  cgcgtctgcg  aaaagcacct  ggccatcgac  agcagcaact
1151  acgagcaaga  gtccaagatc  atgtttgtgg  tgaacgcggt  gtatgccatg
1201  gcccacgctt  tgcacaaaat  gcagcgaccc  ctctgtccca  acactacca
1251  gctttgtgat  gctatgaaga  tcctggatgg  gaagaagttg  tacaaggatt
1301  acttgctgaa  aatcaacttc  acggctccat  tcaacccaaa  taaagatgca
1351  gatagcatag  tcaagtttga  cacttttggg  gatggaatgg  ggcgatacaa
1501  cgtgttcaat  ttccaaaatg  tagtggggaa  gtattcctac  ttgaaagttg
1551  gtcactgggc  agaaacctta  tcgctagatg  tcaactctat  ccactgggtcc
1501  cggaactcag  tccccacttc  ccagtgcagc  gacccctgtg  cccccaatga
1551  aatgaagaat  atgcaaccag  gggatgtctg  ctgctggatt  tgcatccctt
1601  gtgaacccta  cgaataacctg  gctgatgagt  ttacctgtat  ggattgtggg
1651  tctggacagt  ggcccactgc  agacctaaact  ggatgctatg  accttccctga
1701  ggactacatc  aggtgggaag  acgcctgggc  cattggccca  gtcaccattg
1751  cctgtctggg  ttttatgtgt  acatgcatgg  ttgtaactgt  ttttatcaag
1801  cacaacaaca  cacccttggg  caaagcatcg  ggccgagaac  tctgctacat
1851  cttattgttt  ggggttgcc  tgtcatactg  catgacattc  ttcttcattg
1901  ccaagccatc  accagtcac  tgtgcattgc  gccgactcgg  gctggggagt
1951  tccttcgcta  tctgttactc  agccctgctg  accaagacaa  actgcattgc
2001  ccgcatcttc  gatggggtca  agaatggcgc  tcagaggcca  aaattcatca
2051  gcccagttc  tcaggttttc  atctgcctgg  gtctgatcct  ggggcaaatt
2101  gtgatgggtg  ctgtgtggct  catcctggag  gcccaggca  ccaggaggta
2151  tacccttgca  gagaagcggg  aaacagtcac  cctaaaatgc  aatgtcaaa
2201  attccagcat  gttgatctct  cttacctaca  atgtgatcct  ggtgatctta
2251  tgcactgtgt  acgccttcaa  aacgcggaag  tgcccagaaa  atttcaacga
2301  agctaagttc  ataggtttta  ccatgtacac  cacgtgcac  atctggttgg
2351  ccttctctcc  tatattttat  gtgacatcaa  gtgactacag  agtgcagacg
2501  acaaccatgt  gcatctctgt  cagcctgagt  ggctttgtgg  tcttgggctg
2551  tttgtttgca  cccaagggtc  acatcatcct  gtttcaacct  cagaagaatg
2501  ttgtcacaca  cagactgcac  ctcaacaggt  tcagtgtcag  tggaactggg
2551  accacatact  ctcagtcctc  tgcaagcacg  tatgtgccaa  cgggtgtgcaa
2601  tgggcgggaa  gtcctcgact  ccaccacctc  atctctgtga
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Seq ID 43: L697S, V698G, D744N

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1  atgaagatgt  tgacaagact  gcaagttctt  accttagctt  tgttttcaaa
51  gggattttta  ctctcttttag  gggaccataa  ctttctaagg  agagagatta
101  aatagaagg  tgaccttggt  ttagggggcc  tgtttcctat  taacgaaaaa
151  ggcactggaa  ctgaagaatg  tgggcgaatc  aatgaagacc  gagggattca
201  acgcctggaa  gccatgttgt  ttgctattga  tgaaatcaac  aaagatgatt
251  acttgctacc  aggagtgaag  ttgggtgttc  acattttgga  tacatgttca
301  agggatacct  atgcattgga  gcaatcactg  gagtttgtca  gggcatcttt
351  gacaaaagtg  gatgaagctg  agtatatgtg  tcctgatgga  tcctatgcca
501  ttcaagaaaa  catcccactt  ctcatcag  gggtcattgg  tggctcttat
551  agcagtgttt  ccatacaggt  ggcaaacctg  ctgcggtctt  tccagatccc
501  tcagatcagc  tacgcattca  ccagcgccaa  actcagtgat  aagtcgcgct
551  atgattactt  tgccaggacc  gtgccccccg  acttctacca  ggccaaagcc
601  atggctgaga  tcttgcgctt  cttcaactgg  acctacgtgt  ccacagtagc
651  ctccgagggt  gattacgggg  agacagggat  cgaggccttc  gagcaggaag
701  cccgcctgcg  caacatctgc  atcgctacgg  cggagaaggt  gggccgctcc
751  aacatccgca  agtcctacga  cagcgtgatc  cgagaactgt  tgcagaagcc
801  caacgcgcgc  gtgctggtcc  tcttcatgcg  cagcgacgac  tcgcgggagc
851  tcattgcagc  cgccagccgc  gccaatgcct  ccttcacctg  ggtggccagc
901  gacggtggg  atcaccctgg  agctggcctc  ccagcctgtc  cgccagttcg
951  ctacggcgac  atcaccctgg  agctggcctc  ccagcctgtc  cgccagttcg
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Seq ID 44: L697S

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Seq ID 45: L697S, V698G

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Seq ID 46: L697S, D744N

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Seq ID 47: V698G

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Seq ID 48: L697S, V698G, T742A, D744N

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